2014 FINAL REPORT VIRGINIA ~ CHESAPEAKE BAY FINFISH AGEING AND POPULATION ANALYSIS

VMRC ~ ODU Age and Growth Laboratory Center for Quantitative Fisheries Ecology Old Dominion University Norfolk, Virginia 23508

> HONGSHENG LIAO, CYNTHIA M. JONES, & JESSICA L. GILMORE

> > SEPTEMBER 28, 2015





HESAPE

2014 FINAL REPORT VIRGINIA AND CHESAPEAKE BAY FINFISH AGEING AND POPULATION ANALYSIS

Hongsheng Liao, Cynthia M. Jones, & Jessica L. Gilmore

September 28, 2015

Center for Quantitative Fisheries Ecology Old Dominion University 800 West 46th Street Norfolk, VA 23508

Funded by contract No. F-126-R-12 from the Virginia Saltwater Recreational Development Fund through the Virginia Marine Resources Commission

Contents

EXECUTIVE SUMMARY

ACKNOWLEDGEMENTS

1	AT	LANTIC	CROAKER Micropogonias undulatus	1
	1.1	INTRO	DUCTION	2
	1.2	METH	DDS	2
		1.2.1	ample size for ageing	2
		1.2.2	Iandling of collections	2
		1.2.3	Preparation	2
		1.2.4	Readings	3
		1.2.5	Comparison tests	4
	1.3	RESUL	rs	4
		1.3.1	ample size	4
		1.3.2	Reading precision	4
		1.3.3	Tear class	5
		1.3.4	Age-length key	5
	1.4	REFER	ENCES	5
2	$\mathbf{B}\mathbf{L}$	ACK DI	RUM Pogonias cromis	11
	2.1	INTRO	DUCTION	12
	$2.1 \\ 2.2$	INTRO METHO	DUCTION DDS	12 12
	$2.1 \\ 2.2$	INTRO METHO 2.2.1	DUCTION	12 12 12
	$2.1 \\ 2.2$	INTRO METHO 2.2.1 2.2.2	DUCTION	12 12 12 12
	2.1 2.2	INTRO METHO 2.2.1 2.2.2 2.2.3	DUCTION	12 12 12 12 12
	2.1 2.2	INTRO METHO 2.2.1 2.2.2 2.2.3 2.2.4	DUCTION	12 12 12 12 12 13
	2.12.22.3	INTRO METHO 2.2.1 2.2.2 2.2.3 2.2.4 RESUL	DUCTION	12 12 12 12 12 13 14
	2.12.22.3	INTRO METHO 2.2.1 2.2.2 2.2.3 2.2.4 RESUL 2.3.1	DUCTION	12 12 12 12 12 13 14 14
	2.12.22.3	INTRO METHO 2.2.1 2.2.2 2.2.3 2.2.4 RESUL 2.3.1 2.3.2	DUCTION	12 12 12 12 12 13 14 14 14
	2.12.22.3	INTRO METHO 2.2.1 2.2.2 2.2.3 2.2.4 RESUL 2.3.1 2.3.2 2.3.3	DUCTION	$12 \\ 12 \\ 12 \\ 12 \\ 13 \\ 14 \\ 14 \\ 14 \\ 14 \\ 14 \\ 14 \\ 14$
	2.12.22.32.4	INTRO METHO 2.2.1 2.2.2 2.2.3 2.2.4 RESUL 2.3.1 2.3.2 2.3.3 REFER	DUCTION	$12 \\ 12 \\ 12 \\ 12 \\ 13 \\ 14 \\ 14 \\ 14 \\ 15 \\ 15 \\ 12 \\ 12 \\ 12 \\ 12 \\ 12 \\ 12$
2	 2.1 2.2 2.3 2.4 BLU 	INTRO METHO 2.2.1 2.2.2 2.2.3 2.2.4 RESUL 2.3.1 2.3.2 2.3.3 REFER	DUCTION	12 12 12 12 13 14 14 14 14 15
3	 2.1 2.2 2.3 2.4 BLN 3.1 	INTRO METHO 2.2.1 2.2.2 2.2.3 2.2.4 RESUL 2.3.1 2.3.2 2.3.3 REFER	DUCTION	12 12 12 12 13 14 14 14 15 19 20

vii

viii

	3.2	METHODS	J
		3.2.1 Sample size for ageing)
		3.2.2 Handling of collections)
		3.2.3 Preparation)
		3.2.4 Readings	
		3.2.5 Comparison tests	1
	3.3	RESULTS	;
		3.3.1 Sample size	,
		3.3.2 Reading precision	;
		3.3.3 Year class	;
		3.3.4 Age-length key	-
	3.4	REFERENCES	:
4	CO	DIA Deshuardaran sanadaran 91	
4	4.1	$\begin{array}{ccc} \text{DIA} & \textbf{Rachycentron} & \textbf{canadam} & & \textbf{DI} \\ \text{INTRODUCTION} & & 29 \end{array}$	
	4.1	METHODS 22	2)
	4.2	METHODS 32 4.2.1 Handling of collections 22	2)
		4.2.1 Handling of collections	2)
		$4.2.2 \text{Freparation} \qquad \qquad$	2)
		$4.2.5 \text{Readings} \dots \dots \dots \dots \dots \dots \dots \dots \dots $	2 N
	19	4.2.4 Comparison tests	
	4.0	$\begin{array}{cccc} $:
		4.5.1 Reading precision $\dots \dots \dots$:
		$4.3.2 \text{feal class} \dots \dots \dots \dots \dots \dots \dots \dots \dots $:
	<u> </u>	PEFERENCES 35	
	4.4		
5	REI	D DRUM Sciaenops ocellatus 39)
	5.1	INTRODUCTION)
	5.2	METHODS)
		5.2.1 Handling of collections)
		5.2.2 Preparation)
		5.2.3 Readings)
		5.2.4 Comparison tests	
	5.3	RESULTS	1
		5.3.1 Reading precision $\ldots \ldots 42$	1
		5.3.2 Year class	1
		5.3.3 Age-length key	1
	5.4	REFERENCES	
6	SHI	FEDSHEAD Anchosanaus probatogenhalus	,
U	61	INTRODUCTION 48	
	6.2	METHODS 48	
	0.2	6.2.1 Handling of collections 48	
		6.2.2 Preparation /8	
		6.2.3 Beadings 48	
		6.2.4 Comparison tests 40	,
	63	RESULTS	,
	0.0	6.3.1 Reading precision 50	
		out reading production	

		6.3.2	Year class	. 50
		6.3.3	Age-length key	. 50
	6.4	REFE	RENCES	. 50
7	AT]	LANT	IC SPADEFISH Chaetodipterus faber	55
	7.1	INTR	ODUCTION	. 56
	7.2	METI	HODS	. 56
		7.2.1	Sample size for ageing	. 56
		7.2.2	Handling of collections	. 56
		7.2.3	Preparation	. 56
		7.2.4	Readings	. 57
		7.2.5	Comparison tests	. 58
	7.3	RESU	ITS	. 58
		7.3.1	Sample size	. 58
		732	Reading precision	. 58
		733	Vear class	. 50
		7.3.4	Age length low	. 50
	74	7.3.4 DEEE		. 59
	1.4	REF E		. 59
8	SPA	NISH	MACKEBEL Scomberomorous maculatus	63
Ŭ	81	INTR	ODUCTION	64
	8.2	METI	HODS	. 64
	0.2	891	Sample size for agoing	. 01
		822	Handling of collections	. 04
		0.2.2	Propagation	. 04
		0.2.3		. 04 65
		0.2.4		. 00
	0.0	8.2.5	Comparison tests	. 00
	8.3	RESU	LTS	. 66
		8.3.1	Sample size	. 66
		8.3.2	Reading precision	. 66
		8.3.3	Year class	. 67
		8.3.4	Age-length key	. 67
	8.4	REFE	RENCES	. 67
_				
9	SPC	JT Let	nostomus xanthurus	71
	9.1	INTR	ODUCTION	. 72
	9.2	METI	HODS	. 72
		9.2.1	Sample size for ageing	. 72
		9.2.2	Handling of collections	. 72
		9.2.3	Preparation	. 72
		9.2.4	Readings	. 73
		9.2.5	Comparison tests	. 74
	9.3	RESU	ILTS	. 74
		9.3.1	Sample size	. 74
		9.3.2	Reading precision	. 74
		9.3.3	Year class	. 75
		9.3.4	Age-length key	. 75
	9.4	REFE	RENCES	. 75

10 SPC	OTTED SEATROUT Cynoscion nebulosus	79
10.1	INTRODUCTION	80
10.2	METHODS	80
	10.2.1 Sample size for ageing	80
	10.2.2 Handling of collections	80
	10.2.3 Preparation	80
	10.2.4 Readings	81
	10.2.5 Comparison tests	82
10.3	RESULTS	82
	10.3.1 Sample size	82
	10.3.2 Reading precision	82
	10.3.3 Year class	82
	10.3.4 Age-length key	83
10.4	REFERENCES	83
11 STF	RIPED BASS Morone saxatilis	87
11.1	INTRODUCTION	88
11.2	METHODS	88
	11.2.1 Sample size for ageing	88
	11.2.2 Handling of collection	88
	11.2.3 Preparation	89
	11.2.4 Readings	89
11.0	11.2.5 Comparison Tests	92
11.3	RESULTS	92
	11.3.1 Sample size	92
	11.3.2 Scales	93
	11.3.5 Otolitilis	94
	11.3.4 Comparison of scale and otonth ages	94 05
11 /	PECOMMENDATIONS	95
11.4	REFERENCES	95
11.0	TEPERENCES	90
12 SUI	MMER FLOUNDER Paralichthus dentatus	105
12.1	INTRODUCTION	106
12.2	METHODS	106
	12.2.1 Sample size for ageing	106
	12.2.2 Handling of collection	106
	12.2.3 Preparation	107
	12.2.4 Readings	107
	12.2.5 Comparison Tests	110
12.3	RESULTS	110
	12.3.1 Sample size	110
	12.3.2 Scales	111
	12.3.3 Otoliths	112
	12.3.4 Comparison of scale and otolith ages	112
	12.3.5 Age-Length-Key (ALK)	113
12.4	REFERENCES	113

ΤΑι	TOG Tautoga onitis	121
13.1	INTRODUCTION	122
13.2	METHODS	122
	13.2.1 Sample size for ageing	122
	13.2.2 Handling of collection	122
	13.2.3 Preparation	123
	13.2.4 Readings	123
	13.2.5 Comparison Tests	125
13.3	RESULTS	125
	13.3.1 Sample size	125
	13.3.2 Opercula	125
	13.3.3 Otoliths	126
	13.3.4 Comparison of operculum and otolith ages	127
	13.3.5 Age-Length-Key (ALK)	128
13.4	REFERENCES	128
WF	KEISH Composition regulis	127
1/1	INTRODUCTION	138
14.1	METHODS	138
14.2	14.2.1 Sample size for agoing	138
	14.2.2 Handling of collections	138
	14.2.2 Preparation	138
	14.2.4 Readings	130
	11.2.1 Iteaungs	100
	14.2.5 Comparison tests	140
14.3	14.2.5 Comparison tests	$140 \\ 140$
14.3	14.2.5 Comparison tests	$140 \\ 140 \\ 140$
14.3	14.2.5 Comparison tests	140 140 140 140
14.3	14.2.5 Comparison tests	140 140 140 140 141
14.3	14.2.5 Comparison testsRESULTS14.3.1 Sample size14.3.2 Reading precision14.3.3 Year class14.3.4 Age-length key	140 140 140 140 141 141
	 TAU 13.1 13.2 13.3 13.4 WEA 14.1 14.2 	TAUTOG Tautoga onitis Image: State Sta

EXECUTIVE SUMMARY

In this report we present the ageing results of 14 finfish species collected from commercial and recreational catches made in the Chesapeake Bay and Virginia waters of the Atlantic Ocean, U.S.A. in 2014. All fish were collected by the Virginia Marine Resources Commission's (VMRC) Stock Assessment Program and the Center for Quantitative Fisheries Ecology (CQFE) at Old Dominion University in 2014 and aged in 2015 at the Age and Growth Laboratory of CQFE. This report is broken into chapters, one for each of the 14 species. We present measures of ageing precision, graphs of year-class distributions, and age-length keys for each species.

Three calcified structures (hard-parts) are used in age determination. Specifically, two calcified structures were used for determining fish ages of the following three species: striped bass, Morone saxatilis, (n = 856); summer flounder, Paralichthys dentatus, (n = 856); 819): and tautog. Tautoga onitis, (n = 308). Scales and otoliths were used to age striped bass and summer flounder, opercula and otoliths were used to age tautog. Comparing alternative hard-parts allowed us to assess their usefulness in determining fish age as well as the relative precision of each structure. Ages were determined from otoliths only for the following species: Atlantic croaker, Micropogonias undulatus, (n = 364); black drum, Pogonias cromis, (n = 123); bluefish, Pomatomus saltatrix, (n = 366); cobia, Rachycentron canadum, (n = 287); red drum, Sciaenops ocellatus, (n = 80); sheepshead, Archosargus probatocephalus, (n = 120); Atlantic spadefish, Chaetodipterus faber, (n = 288); Spanish mackerel, Scomberomorous maculates, (n = 236); spot, Leiostomus xanthurus, (n = 276); spotted seatrout, Cynoscion nebulosus, (n = 273); and weakfish, Cynoscion regalis, (n = 273)295). In total, we made 10,926 age readings from scales, otoliths and opercula collected during 2014. A summary of the age ranges for all species aged is presented in Table 1.

In this report, we also present sample sizes and coefficient of variation (CV) for estimates of age composition for the following species: Atlantic croaker, bluefish, spadefish, Spanish mackerel, spot, spotted seatrout, striped bass, summer flounder, tautog, and weakfish. The sample sizes and the CVs enabled us to determine how many fish we needed to age in each length interval and to measure the precision for estimates of major age classes in each species, respectively, enhancing our efficiency and effectiveness on ageing those species.

To support environmental and wildlife agencies, and charities, we donated more than 2,529 pounds of dissected fish to the Salvation Army to feed the homeless, and the Wildlife Response, Inc., a local wildlife rescue agency which is responsible for saving injured animals found by the public.

In 2014, we continued to upgrade our Age and Growth Laboratory website, which can be accessed at http://odu.edu/sci/research/cqfe/research/ageing-lab. The website includes an electronic version of this document and our previous VMRC final reports from 2001 to 2013. The site also provides more detailed explanations of the methods and structures we use in age determination.

Table 1: The minimum and maximum ages, number of fish and their hard-parts collected, number of fish aged, and age readings for the 14 finfish species in 2014. The hard-parts and age readings include both scales and otoliths for striped bass and summer flounder, and both opercula and otoliths for tautog.

Species	Nun	nber	Number	Nun	nber	Num	ber	Minimum	Maximum
	of	fish	of hard-	of	fish	of re	ead-	age	age
	colle	ected	parts	ageo	l	ings			
Atlantic Croaker	514		514	364		728		1	10
Black Drum	124		123	123		246		3	47
Bluefish	577		577	366		732		0	11
Cobia	296		287	287		574		2	12
Red Drum	80		80	80		160		0	2
Sheepshead	123		120	120		240		2	28
Spadefish	325		323	288		576		0	7
Spanish Mackerel	253		253	236		472		0	6
Spotted Seatrout	362		362	273		546		0	8
Spot	336		336	276		552		0	3
Striped Bass	$1,\!10$	5	$1,\!297$	856		2,194		2	23
Summer Flounder	$1,\!10$	1	$1,\!307$	819		2,098		0	14
Tautog	309		615	308		1,218		3	30
Weakfish	377		377	295		590		1	6
Totals	$5,\!88$	2	$6,\!571$	4,69	1	$10,\!92$	6		

ACKNOWLEDGEMENTS

We thank Alicia Brown, Aris Horsey, and Allison Roberts for their technical expertise in preparing otoliths, scales, and opercula for age determination. They all put in long hours processing "tons" of fish in our lab. A special note of appreciation is extended to Joe Grist, Joe Cimino, Adam Kenyon, and their technicians at the VMRC, including Richard Hancock, Myra Thompson, and Chris Williams for their many efforts in this cooperative project. We would like also to thank our graduate students Kathleen Kirch and Mike Schmidtke for their help in processing fish whenever we were short of hands.

Chapter 1

ATLANTIC CROAKER Micropogonias undulatus



1.1 INTRODUCTION

We aged a total of 364 Atlantic croaker, *Micropogonias undulatus*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2014. Croaker ages ranged from 1to 10 years old with an average age of 4.7, a standard deviation of 1.8, and a standard error of 0.09. Ten age classes (1 to 10) were represented, comprising fish of the 2004 to 2013 year-classes. The sample was dominated by fish from the year-classes of 2008 and 2010 with 28.8% and 26.6%, respectively.

1.2 METHODS

1.2.1 Sample size for ageing

We estimated sample size for ageing croaker in 2014 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 C V^2 + B_a/L} \tag{1.1}$$

where A is the sample size for ageing croaker in 2014; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; Lwas the total number of croaker used by VMRC to estimate length distribution of the catches from 2008 to 2012. θ_a , V_a , B_a , and CV were calculated using pooled agelength data of croaker collected from 2008 to 2012 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (1.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion

to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by ageing an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2008 to 2012 catch. A_l is number of fish to be aged for length interval l in 2014.

1.2.2 Handling of collections

Otoliths were received by the Age & and Growth Laboratory in labeled coin envelopes, were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

1.2.3 Preparation

Sagittal otoliths, hereafter, referred to as "otoliths", were processed for age determination following the methods described in Barbieri et al. (1994) with a few The left or right otolith modifications. was randomly selected and attached, distal side down, to a glass slide with clear CrystalbondTM 509 adhesive or imbedded in epoxy. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a pencil across the otolith surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMetTM lowspeed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). Thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thinsections.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing Atlantic croaker.

1.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the speciesspecific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, Atlantic croaker otolith annulus formation occurs between April and May (Barbieri et al. 1994a and b). A croaker captured between January 1 and May 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification. Each reader aged all of the otolith samples.

Due to discrepancy on identification of the first annulus of Atlantic croaker among Atlantic states, Atlantic States Marine Fisheries Commission (ASMFC) has decided not to count the smallest annulus at the center of the thin-section as the first annulus. Following ASMFC's instruction, we didn't count the smallest annulus at the center as the first annulus in 2014 (Figure 1.1).

All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers'



Figure 1.1: Otolith thin-sections of a 8 yearold croaker without counting the smallest ring and with the last annulus on the edge of the thin-section

ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age Atlantic croaker using their otolith thinsections.

1.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

1.3 RESULTS

1.3.1 Sample size

We estimated a sample size of 463 Atlantic croaker in 2014, ranging in length interval from 6 to 20 inches (Table 1.1). This sample size provided a range in (CV) for age composition approximately from the smallest (CV) of 9% for age 4 to the largest (CV) of 21% for age 2. In 2014, we randomly selected and aged 364 fish from 514 croaker collected by VMRC. We fell short in our over-all collections for this optimal length-class sampling estimate by 104 fish. However, we were short of many fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would definitely be influenced significantly. Therefore, precaution should be used when developing ALK using these age data.

1.3.2 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 98% and a CV of 0.3% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 94% and a CV of 1.4% (test of symmetry: $\chi^2 = 3$, df = 2, P = 0.2231). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 96.7% and a CV of 0.6% (test of symmetry: $\chi^2 = 5.2$, df = 5, P = 0.392) (Figure 1.2).



Figure 1.2: Between-reader comparison of otolith age estimates for Atlantic croaker collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

There was no time-series bias for both readers. Reader 1 had an agreement of 98% with ages of fish aged in 2003 with a CV of 0.1% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173). Reader 2 had an agreement of 100%.

1.3.3 Year class

Of the 364 fish aged with otoliths, 10 age classes (1 to 10) were represented (Table 1.2). The average age was 4.7 years, and the standard deviation and standard error were 1.8 and 0.09, respectively. Year-class data show that the fishery was comprised of 10 year-classes: fish from the 2004 to 2013 year-classes, with fish primarily from the year classes of 2008 and 2010 with 28.8% and 26.6%, respectively. The ratio of males to females was 1:1.84 in the sample collected (Figure 1.3).

1.3.4 Age-length key

We developed an age-length-key (Table 1.3) that can be used in the conversion of numbers-at-length in the estimated catch



Figure 1.3: Year-class frequency distribution for Atlantic croaker collected for ageing in 2014. Distribution is broken down by sex. 'Unknown' is for gonads that were not available for examination or were not examined for sex during sampling.

to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

1.4 REFERENCES

- Barbieri, L. R., Chittenden, M.E. Jr., Lowerre-Barbieri, S.K. 1994a. Maturity, spawning, and ovarian cycle of Atlantic croaker, Micropogonias undulatus, in the Chesapeake bay and adjacent coastal waters. Fishery Bulletin 92:671-685.
- Barbieri, L. R., Chittenden, M.E. Jr., Lowerre-Barbieri, S.K. 1994b. Age, growth, and mortality of Atlantic croaker, Micropogonias undulatus, in the Chesapeake bay region, with a discussion of apparent geographic changes in population dynamics. Fishery Bulletin 92:1-12.
- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.

- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analysing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- Quinn, T. J. II, and R. B. Deriso. 1999. Quantitative Fish Dynamics. Oxford University Press. New York.
- R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.

Table 1.1: Number of Atlantic croaker collected and aged in each 1-inch length interval in 2014.
'Target' represents the sample size for ageing estimated for 2014, and 'Need' represents number of
fish shorted in each length interval compared to the optimum sample size for ageing and number of
fish aged.

Interval	Target	Collected	Aged	Need
6 - 6.99	5	1	1	4
7 - 7.99	7	24	8	0
8 - 8.99	7	27	8	0
9 - 9.99	19	29	20	0
10 - 10.99	30	52	30	0
11 - 11.99	54	78	54	0
12 - 12.99	97	158	98	0
13 - 13.99	72	73	73	0
14 - 14.99	59	39	39	20
15 - 15.99	44	24	24	20
16 - 16.99	29	8	8	21
17 - 17.99	23	1	1	22
18 - 18.99	7	0	0	7
19 - 19.99	5	0	0	5
20 - 20.99	5	0	0	5
Totals	463	514	364	104

					Age						
Interval	1	2	3	4	5	6	7	8	9	10	Totals
6 - 6.99	0	1	0	0	0	0	0	0	0	0	1
7 - 7.99	5	3	0	0	0	0	0	0	0	0	8
8 - 8.99	0	7	0	1	0	0	0	0	0	0	8
9 - 9.99	0	12	1	6	0	1	0	0	0	0	20
10 - 10.99	0	9	0	12	6	3	0	0	0	0	30
11 - 11.99	0	5	1	27	7	13	0	1	0	0	54
12 - 12.99	0	9	1	23	17	37	6	3	2	0	98
13 - 13.99	0	8	1	15	12	26	3	6	2	0	73
14 - 14.99	0	4	0	5	10	13	2	3	1	1	39
15 - 15.99	0	2	0	7	5	8	1	0	0	1	24
16 - 16.99	0	0	0	1	1	3	0	3	0	0	8
17 - 17.99	0	0	0	0	0	1	0	0	0	0	1
Totals	5	60	4	97	58	105	12	16	5	2	364

Table 1.2: The number of Atlantic croaker assigned to each total length-at-age category for 364 fish sampled for otolith age determination in Virginia during 2014.

					Age					
Interval	1	2	3	4	5	6	7	8	9	10
6 - 6.99	0	1	0	0	0	0	0	0	0	0
7 - 7.99	0.62	0.38	0	0	0	0	0	0	0	0
8 - 8.99	0	0.88	0	0.12	0	0	0	0	0	0
9 - 9.99	0	0.6	0.05	0.3	0	0.05	0	0	0	0
10 - 10.99	0	0.3	0	0.4	0.2	0.1	0	0	0	0
11 - 11.99	0	0.09	0.02	0.5	0.13	0.24	0	0.02	0	0
12 - 12.99	0	0.09	0.01	0.23	0.17	0.38	0.06	0.03	0.02	0
13 - 13.99	0	0.11	0.01	0.21	0.16	0.36	0.04	0.08	0.03	0
14 - 14.99	0	0.1	0	0.13	0.26	0.33	0.05	0.08	0.03	0.03
15 - 15.99	0	0.08	0	0.29	0.21	0.33	0.04	0	0	0.04
16 - 16.99	0	0	0	0.12	0.12	0.38	0	0.38	0	0
17 - 17.99	0	0	0	0	0	1	0	0	0	0

Table 1.3: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for Atlantic croaker sampled for age determination in Virginia during 2014.

Chapter 2

BLACK DRUM Pogonias cromis



2.1 INTRODUCTION

We aged a total of 123 black drum, *Pogonias cromis*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2014. Black drum ages ranged from 3 to 47 years old with an average age of 11.2, a standard deviation of 9.2, and a standard error of 0.83. Twentysix age classes (3 to 17, 22, 24 to 26, 29, 33 to 35, 42 to 43, 47) were represented, comprising fish of the 1967, 1971 to 1972, 1979 to 1981, 1985, 1988 to 1990, 1992, 1997 to 2011 year-classes. The sample was dominated by fish from the year-classes of 2009 and 2007 with 17.1% and 15.4%, respectively.

2.2 METHODS

2.2.1 Handling of collections

Sagittal otoliths, hereafter, referred to as "otoliths", were received by the Age and Growth Laboratory in labeled coin envelopes. In the lab they were sorted by date of capture, their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

2.2.2 Preparation

Otoliths were processed for age determination following the methods described in Bobko (1991) and Jones and Wells (1998). The left or right sagittal otolith was randomly selected and attached, distal side down, to a glass slide with CrystalbondTM 509 adhesive or embedded in epoxy. The otoliths were viewed by eye, and when necessary, under a stereo microscope to identify the location of the core, and the position of the core marked using a pencil across the otolith surface. At least one transverse cross-section (hereafter "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMetTM low-speed saw equipped with two, three inch diameter, Norton Diamond Grinding Wheels, separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The position of the marked core fell within the 0.5 mm space between the blades, such that the core was included in the removed thinsection. Otolith thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the sections.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing black drum.

2.2.3 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the speciesspecific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, black drum otolith annulus formation occurs between May and June (Beckman et al. 1990; Bobko 1991; Jones and Wells 1998). A black drum captured between January 1 and June 30, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 2.1). Each reader aged all of the otolith samples.

All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or



Figure 2.1: Otolith thin-section of a 3 year-old black drum

the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age black drum using their otolith thin-sections.

2.2.4 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

2.3 RESULTS

2.3.1 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 98% and a CV of 0.2% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 94% and a CV of 0.7% (test of symmetry: $\chi^2 = 3$, df = 3, P = 0.3916). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 93.5% and a CV of 0.3% (test of symmetry: $\chi^2 = 8$, df = 8, P = 0.4335) (Figure 2.2).



Figure 2.2: Between-reader comparison of otolith age estimates for black drum collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

There was no time-series bias for both readers. Reader 1 had an agreement of 78% with ages of fish aged in 2000 with a CV of 0.5% (test of symmetry: $\chi^2 = 11$, df = 10, P = 0.3575). Reader 2 had an agreement of 90% with a CV of 0.3% (test of symmetry: $\chi^2 = 5$, df = 5, P = 0.4159).

2.3.2 Year class

Of the 123 fish aged with otoliths, 26 age classes (3 to 17, 22, 24 to 26, 29, 33 to 35, 42 to 43, 47) were represented (Table 2.1). The average age was 11.2 years, and the standard deviation and standard error were 9.2 and 0.83, respectively. Year-class data show that the fishery was comprised of 26 year-classes: fish from the 1967, 1971 to 1972, 1979 to 1981, 1985, 1988 to 1990, 1992, 1997 to 2011 year-classes, with fish primarily from the year classes of 2009 and 2007 with 17.1% and 15.4%, respectively. The ratio of males to females was 1:0.83 in the sample collected (Figure 2.3).



Figure 2.3: Year-class frequency distribution for black drum collected for ageing in 2014. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

2.3.3 Age-length key

We developed an age-length-key (Table 2.2) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

2.4 REFERENCES

- Beckman, D. W., C. A. Wilson, and A. L. Stanley. 1990. Age and growth of black drum in Louisiana waters of the Gulf of Mexico. Transactions of American Fisheries Society 19:537-544.
- Bobko, S. J. 1991. Age, growth, and reproduction of black drum, Pogonias cromis, in Virginia. M.S. thesis. Old Dominion University, Norfolk, VA.
- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age eterminations. Trans. Am. Fish. Soc. 124:131-138.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analysing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- Jones, C.J. and B.K. Wells. 1998. Age, growth, and mortality of black drum, Pogonias cromis, in the Chesapeake Bay region. Fish. Bull. 96:451-461.
- R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.

2																												
5		48 -	47 -	46 -	44 -	43 -	42 -	39 -	38 -	37 -	36 -	35 -	34 -	ယ္လ္လ ယ	32 -	31 -	30 -	29 -	28 -	27 -	25 -	24 -	23 -	22 -	0	Ιı		0
₹ †	Tota	- 48.	- 47.	- 46.	- 44.	- 43.	- 42.	- 39.	- 38.	- 37.	- 36.	35.	- 34.	 	- 32.	- 31.	- 30.	- 29.	- 28.	- 27.	25.	- 24.	- 23.	- 22.	- 0.	nterv		
	als	66	66	66	66	66	66	66	66	66	66	66	66	66	66	66	66	66	66	66	66	66	66	66	66	7al		
-	10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	2	ယ	ယ	Ľ	0	ω		
	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	0	4		
	21	0	0	0	0	0	0	0	0	0	0	0	0	1	0	4	4	6	4	2	0	0	0	0	0	5		
	4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	2	1	0	0	0	0	0	0	0	6		
	19	0	0	0	0	0	0	0	0	0	0	0	1	ω	9	ယ	⊢	2	0	0	0	0	0	0	0	7		
	11	0	0	0	0	0	0	0	0	0	0	2	1	6	0	2	0	0	0	0	0	0	0	0	0	8		
	∞	0	0	0	0	0	0	0	1	0	ယ	1	1	2	0	0	0	0	0	0	0	0	0	0	0	9		
	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	10		
	2	0	0	0	0	0	0	0	0	0	0	⊢	0	0	Ц	0	0	0	0	0	0	0	0	0	0	11		
	∞	0	0	0	0	0	0	1	Ц	4	μ	0	Ц	0	0	0	0	0	0	0	0	0	0	0	0	12		
	13	0	0	0	0	0	0	1	ယ	1	2	ы	1	0	0	0	0	0	0	0	0	0	0	0	0	13		
	ц	0	0	0	0	0	0	0	0	0		0	0	0	0	0	0	0	0	0	0	0	0	0	0	14		
	_	_	_	_	_	_	_					_	_	_	_	_	_	_	_			_	_	_		1	Ag	
	6	0	0	0	0	0	0	0	1	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	5 1	e	
	μ	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	6 1		
	2	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	7		
	2	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	22		
	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	24		
	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	25		
	н	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	26		
	Ц	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	29		
	Ц	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	33		
	1	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	34		
	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	35		
	ယ	0	2	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	42		
		-		-	_	-	-	-	-	-	-	-	-	-	-	-	-	-	-	- -		-	-	-	-	45		
			_	_	_		_		_	_	_	_	_		_	_	_	_	_	_	_	_	_	_	_	3 4		
	-	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	7 J		
	123	1	2	1	СЛ	1	ယ	6	6	7	7	9	6	12	10	10	7	9	Ċī	4	2	ယ	ట	1	ట	otals		

Table 2.1: The number of black drum assigned to each total length (inch)-at-age category for 123 fish sampled for otolith age determination in Virginia

Table 2.2: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for black drum sampled for age determination in Virginia during 2014.

												0												
nterval	3	4	5	5 7	8	6	10	11	12	13	14	15	16	17	22	24	25	26	29	33	34	35	42	13
0.99	0	0	0	0	0	0	0	0	0	0	0	0.33	0	0.33	0.33	0	0	0	0	0	0	0	0	0
- 22.99	1	0	0	0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 23.99	1	0	0	0 (0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 24.99	1	0	0	0 (0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 25.99	1	0	0	0 (0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 27.99 0.5	25 0.:	25 0.	5	0 (0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 28.99	0 0	.2 0.	8	0 (0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 29.99	0	0 0.6	7 0.1	1 0.22	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 30.99	0	0 0.5	7 0.29	9 0.14	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 31.99	0	0 0.	4 0.	1 0.3	0.2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 32.99	0	0	0	0.0	0	0	0	0.1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 33.99	0	0.0.0	8	0.25	0.5	0.17	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 34.99	0	0	0	0.17 O	0.17	0.17	0.17	0	0.17	0.17	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 35.99	0	0	0	0	0.22	0.11	0	0.11	0	0.56	0	0	0	0	0	0	0	0	0	0	0	0	0	0
- 36.99	0	0	0	0 0	0	0.43	0	0	0.14	0.29	0.14	0	0	0	0	0	0	0	0	0	0	0	0	0
- 37.99	0	0	0	0 0	0	0	0	0	0.57	0.14	0	0.29	0	0	0	0	0	0	0	0	0	0	0	0
- 38.99	0	0	0	0	0	0.17	0	0	0.17	0.5	0	0.17	0	0	0	0	0	0	0	0	0	0	0	0
- 39.99	0	0	0	0	0	0	0	0	0.17	0.17	0	0.33	0.17	0.17	0	0	0	0	0	0	0	0	0	0
- 42.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0.33	0.33	0	0.33	0	0	0	0	0
- 43.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
- 44.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0.2	0	0	0.2	0	0.2	0	0.2	0.2	0
- 46.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
- 47.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
- 48.99	0	0	0	0 0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0

Chapter 3

BLUEFISH Pomatomus saltatrix



3.1 INTRODUCTION

We aged a total of 366 bluefish, *Pomatomus* saltatrix, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2014. Bluefish ages ranged from 0 to 11 years old with an average age of 2.4, a standard deviation of 2, and a standard error of 0.1. Twelve age classes (0 to 11) were represented, comprising fish of the 2003 to 2014 year-classes. The sample was dominated by fish from the year-classes of 2012 and 2013 with 32.8% and 31.4%, respectively.

3.2 METHODS

3.2.1 Sample size for ageing

We estimated sample size for ageing bluefish in 2014 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 C V^2 + B_a/L} \tag{3.1}$$

where A is the sample size for ageing bluefish in 2014; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CVis the coefficient of variation; L was the total number of bluefish used by VMRC to estimate length distribution of the catches from 2008 to 2012. θ_a , V_a , B_a , and CV were calculated using pooled age-length data of bluefish collected from 2008 to 2012 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (3.1) indicates that the more fish that are aged, the smaller the CV(or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above

which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2008 to 2012 catch. A_l is number of fish to be aged for length interval l in 2014. Based on VMRC's request in 2010, we used 1-cm length interval for bluefish, which differed from other species (1-inch).

3.2.2 Handling of collections

Otoliths were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

3.2.3 Preparation

We used our thin-section and bake technique to process bluefish sagittal otoliths (hereafter, referred to as "otoliths") for age determination (Robillard et al. 2009). Otolith preparation began by randomly selecting either the right or left otolith. Each whole otolith was placed in a ceramic "Coors" spot plate well and baked in a Thermolyne 1400 furnace at 400 °C. Baking time was dependent on the otolith's size and gauged by color, with a light caramel color desired. Once a suitable color was achieved the baked otolith was embedded in epoxy resin with its distal surface orientated downwards. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core. Then, the position of the core was marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter, referred to

as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMetTM low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The otolith was positioned so the blades straddled each side of the otolith focus. It was crucial that this cut be perpendicular to the long axis of the otolith. Failure to do so resulted in broad and distorted winter growth zones. A proper cut resulted in annuli that were clearly defined and delineated. Once cut, thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thinsection.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing bluefish.

3.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the speciesspecific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, bluefish otolith deposition occurs between March and June (Robillard et al. 2009). A bluefish captured between January 1 and May 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 3.1). Each reader aged all of the otolith samples.



Figure 3.1: Otolith thin-section of a 5 year-old bluefish with the last annulus on the edge of the thin-section

If an otolith was properly sectioned, the sulcal groove came to a sharp point within the middle of the focus. Typically the first year's annulus was found by locating the focus of the otolith, which was characterized as a visually distinct dark, oblong region found in the center of the otolith. The first year's annulus had the highest visibility proximal to the focus along the edge of the sulcal groove. Once located, the first year's annulus was followed outward from the sulcal groove towards the dorsal perimeter of the otolith. Often, but not always, the first year was associated with a very distinct crenellation on the dorsal surface and a prominent protrusion on the ventral surface. Both of these landmarks had a tendency to become less prominent in older fish.

Even with the bake and thin-section technique, interpretation of the growth zones from the otoliths of young bluefish was difficult. Rapid growth within the first year of life prevents a sharp delineation between opaque and translucent zones. When the exact location of the first year was not clearly evident, and the otolith had been sectioned accurately, a combination of surface landscape (1st year crenellation) and the position of the second annuli were used to help determine the position of the first annulus.

What appeared to be "double annuli" were occasionally observed in bluefish 4-7 years

of age and older. This double-annulus formation was typically characterized by distinct and separate annuli in extremely close proximity to each other. We do not know if the formation of these double annuli were two separate annuli, or in fact only one, but they seemed to occur during times of reduced growth after maturation. "Double annuli" were considered to be one annulus when both marks joined to form a central origin (the origin being the sulcal groove and the outer peripheral edge of the otolith). If these annuli did not meet to form a central origin they were considered two distinct annuli, and were counted as such.

All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age bluefish using their otolith thin-sections.

3.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

3.3 RESULTS

3.3.1 Sample size

We estimated a sample size of 449 bluefish in 2014, ranging in length interval from 16 to 97 centimeters (Table 3.1). This sample size provided a range in (CV) for age composition approximately from the smallest (CV) of 5% for age 2 to the largest (CV)of 21% for age 5. In 2014, we randomly selected and aged 366 fish from 577 bluefish collected by VMRC. We fell short in our over-all collections for this optimal lengthclass sampling estimate by 115 fish. However, we were short of no fish from the major length intervals (The interval requires more than 5 fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

3.3.2 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 94% and a CV of 1.7% (test of symmetry: $\chi^2 = 3$, df = 3, P = 0.3916), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 90% and a CV of 4.5% (test of symmetry: $\chi^2 = 5$, df = 4, P = 0.2873). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 84.15% and a CV of 6.4% (test of symmetry: $\chi^2 = 15.47$, df =10, P = 0.1157) (Figure 3.2).



Figure 3.2: Between-reader comparison of otolith age estimates for bluefish collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

There was no time-series bias for both readers. Reader 1 had an agreement of 96% with ages of fish aged in 2000 with a CV of 3.8% (test of symmetry: $\chi^2 = 2$, df = 2, P = 0.3679). Reader 2 had an agreement of 98% with a CV of 0.9% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173).

3.3.3 Year class

Of the 366 fish aged with otoliths, 12 age classes (0 to 11) were represented (Table 3.2). The average age was 2.4 years, and the standard deviation and standard error were 2 and 0.1, respectively. Year-class data show that the fishery was comprised of 12 year-classes: fish from the 2003 to 2014 year-classes, with fish primarily from the year classes of 2012 and 2013 with 32.8% and 31.4%, respectively. The ratio of males to females was 1:2.39 in the sample collected (Figure 3.3).



Figure 3.3: Year-class frequency distribution for bluefish collected for ageing in 2014. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

3.3.4 Age-length key

We developed an age-length-key (Table 3.3) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length cm intervals.

3.4 REFERENCES

- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analysing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- Quinn, T. J. II, and R. B. Deriso. 1999. Quantitative Fish Dynamics. Oxford University Press. New York.
- R Core Team (2012). R: A lan-

guage and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www. R-project.org.

Robillard, E. M., C. S. Reiss, and C. M. Jones. 2009. Age-validation and growth of bluefish (Pomatomus saltatrix) along the East Coast of the United States. Fisheries Research 95:65-75.
Table 3.1:	Number of	blue fish	collecte	d and	aged in	n each	$1\text{-}\mathrm{cm}$	length	interval i	in 2014.	'Tar	get'
represents	the sample	size for	ageing	estima	ted for	2014,	and	'Need'	represent	s numbe	er of	fish
shorted in	each length	interval o	compare	d to th	ne optir	num sa	mple	size for	ageing a	nd numb	er of	fish
aged.												

(Go back to text)				
Interval	Target	Collected	Aged	Need
16 - 16.99	5	1	1	4
17 - 17.99	5	0	0	5
18 - 18.99	5	0	0	5
19 - 19.99	5	6	6	0
20 - 20.99	5	20	6	0
21 - 21.99	5	14	5	0
22 - 22.99	5	18	5	0
23 - 23.99	5	13	6	0
24 - 24.99	5	13	6	0
25 - 25.99	5	15	6	0
26 - 26.99	5	12	6	0
27 - 27.99	5	11	6	0
28 - 28.99	5	10	6	0
29 - 29.99	5	12	6	0
30 - 30.99	5	12	8	0
31 - 31.99	5	8	6	0
32 - 32.99	5	14	6	0
33 - 33.99	6	11	6	0
34 - 34.99	6	6	6	0
35 - 35.99	6	9	6	0
36 - 36.99	8	11	8	0
37 - 37.99	8	14	8	0
38 - 38.99	7	16	7	0
39 - 39.99	9	11	9	0
40 - 40.99	8	14	8	0
41 - 41.99	8	12	8	0
42 - 42.99	8	18	8	0
43 - 43.99	8	14	8	0
44 - 44.99	7	9	7	0
45 - 45.99	9	12	9	0
46 - 46.99	7	8	7	0
47 - 47.99	8	15	8	0
48 - 48.99	6	10	6	0
49 - 49.99	5	13	6	0
50 - 50.99	5	11	6	0
51 - 51.99	5	4	4	1
52 - 52.99	5	10	6	0
53 - 53.99	5	5	5	0
54 - 54.99	5	5	5	0
55 - 55.99	5	6	6	0

10	1				
((i	οt	ac	k 1	tO 1	text

(To continue)

_

Interval	Target	Collected	Aged	Need
56 - 56.99	5	11	6	0
57 - 57.99	5	6	6	0
58 - 58.99	5	13	6	0
59 - 59.99	5	15	6	0
60 - 60.99	5	7	6	0
61 - 61.99	5	14	6	0
62 - 62.99	5	5	5	0
63 - 63.99	5	11	11	0
64 - 64.99	5	5	5	0
65 - 65.99	5	4	4	1
66 - 66.99	5	2	2	3
67 - 67.99	5	1	1	4
68 - 68.99	5	3	3	2
69 - 69.99	5	4	4	1
70 - 70.99	5	5	5	0
71 - 71.99	5	4	4	1
72 - 72.99	5	3	3	2
73 - 73.99	5	3	3	2
74 - 74.99	5	3	3	2
75 - 75.99	5	0	0	5
76 - 76.99	5	4	4	1
77 - 77.99	5	5	5	0
78 - 78.99	5	2	2	3
79 - 79.99	5	2	2	3
80 - 80.99	5	1	1	4
81 - 81.99	5	7	7	0
82 - 82.99	5	4	4	1
83 - 83.99	5	1	1	4
84 - 84.99	5	1	1	4
85 - 85.99	5	0	0	5
86 - 86.99	5	3	3	2
87 - 87.99	5	0	0	5
88 - 88.99	5	1	1	4
89 - 89.99	5	0	0	5
90 - 90.99	5	1	1	4
91 - 91.99	5	2	2	3
92 - 92.99	5	0	0	5
93 - 93.99	5	0	0	5
94 - 94.99	5	0	0	5
95 - 95.99	5	0	0	5
96 - 96.99	5	1	1	4
97 - 97.99	5	0	0	5
Totals	449	577	366	115

Table 3.1 (Continued)

Table 3.2: The number of bluefish assigned to each total length (cm)-at-age category for 366 fish sampled for otolith age determination in Virginia during 2014.

(Go back to text)

						Age							
Interval	0	1	2	3	4	5	6	7	8	9	10	11	Totals
16 - 16.99	1	0	0	0	0	0	0	0	0	0	0	0	1
19 - 19.99	6	0	0	0	0	0	0	0	0	0	0	0	6
20 - 20.99	3	3	0	0	0	0	0	0	0	0	0	0	6
21 - 21.99	2	3	0	0	0	0	0	0	0	0	0	0	5
22 - 22.99	3	2	0	0	0	0	0	0	0	0	0	0	5
23 - 23.99	3	3	0	0	0	0	0	0	0	0	0	0	6
24 - 24.99	2	4	0	0	0	0	0	0	0	0	0	0	6
25 - 25.99	0	6	0	0	0	0	0	0	0	0	0	0	6
26 - 26.99	0	6	0	0	0	0	0	0	0	0	0	0	6
27 - 27.99	0	6	0	0	0	0	0	0	0	0	0	0	6
28 - 28.99	0	6	0	0	0	0	0	0	0	0	0	0	6
29 - 29.99	0	6	0	0	0	0	0	0	0	0	0	0	6
30 - 30.99	0	8	0	0	0	0	0	0	0	0	0	0	8
31 - 31.99	0	6	0	0	0	0	0	0	0	0	0	0	6
32 - 32.99	0	6	0	0	0	0	0	0	0	0	0	0	6
33 - 33.99	0	4	2	0	0	0	0	0	0	0	0	0	6
34 - 34.99	0	5	1	0	0	0	0	0	0	0	0	0	6
35 - 35.99	0	4	2	0	0	0	0	0	0	0	0	0	6
36 - 36.99	0	7	1	0	0	0	0	0	0	0	0	0	8
37 - 37.99	0	5	3	0	0	0	0	0	0	0	0	0	8
38 - 38.99	0	3	4	0	0	0	0	0	0	0	0	0	7
39 - 39.99	0	3	6	0	0	0	0	0	0	0	0	0	9
40 - 40.99	0	1	7	0	0	0	0	0	0	0	0	0	8
41 - 41.99	0	2	6	0	0	0	0	0	0	0	0	0	8
42 - 42.99	0	1	7	0	0	0	0	0	0	0	0	0	8
43 - 43.99	0	0	6	2	0	0	0	0	0	0	0	0	8
44 - 44.99	0	2	4	1	0	0	0	0	0	0	0	0	7
45 - 45.99	0	2	7	0	0	0	0	0	0	0	0	0	9
46 - 46.99	0	1	6	0	0	0	0	0	0	0	0	0	7
47 - 47.99	0	2	5	1	0	0	0	0	0	0	0	0	8
48 - 48.99	0	1	5	0	0	0	0	0	0	0	0	0	6
49 - 49.99	0	1	4	1	0	0	0	0	0	0	0	0	6
50 - 50.99	0	1	3	2	0	0	0	0	0	0	0	0	6
51 - 51.99	0	1	2	1	0	0	0	0	0	0	0	0	4
52 - 52.99	0	1	3	2	0	0	0	0	0	0	0	0	6
53 - 53.99	0	1	3	1	0	0	0	0	0	0	0	0	5
54 - 54.99	0	2	2	1	0	0	0	0	0	0	0	0	5
55 - 55.99	0	0	4	2	0	0	0	0	0	0	0	0	6
56 - 56.99	0	0	3	3	0	0	0	0	0	0	0	0	6
57 - 57.99	0	0	2	3	1	0	0	0	0	0	0	0	6
58 - 58.99	0	0	3	2	1	0	0	0	0	0	0	0	6

(To continue)

Table 3.2 (Continued)

						Age							
Interval	0	1	2	3	4	5	6	7	8	9	10	11	Totals
59 - 59.99	0	0	2	3	1	0	0	0	0	0	0	0	6
60 - 60.99	0	0	3	3	0	0	0	0	0	0	0	0	6
61 - 61.99	0	0	3	3	0	0	0	0	0	0	0	0	6
62 - 62.99	0	0	4	0	1	0	0	0	0	0	0	0	5
63 - 63.99	0	0	5	5	0	0	1	0	0	0	0	0	11
64 - 64.99	0	0	1	0	3	0	1	0	0	0	0	0	5
65 - 65.99	0	0	1	0	2	1	0	0	0	0	0	0	4
66 - 66.99	0	0	0	1	1	0	0	0	0	0	0	0	2
67 - 67.99	0	0	0	0	0	1	0	0	0	0	0	0	1
68 - 68.99	0	0	0	0	3	0	0	0	0	0	0	0	3
69 - 69.99	0	0	0	0	4	0	0	0	0	0	0	0	4
70 - 70.99	0	0	0	0	3	2	0	0	0	0	0	0	5
71 - 71.99	0	0	0	0	2	1	1	0	0	0	0	0	4
72 - 72.99	0	0	0	1	1	0	1	0	0	0	0	0	3
73 - 73.99	0	0	0	0	1	2	0	0	0	0	0	0	3
74 - 74.99	0	0	0	0	0	0	2	0	1	0	0	0	3
76 - 76.99	0	0	0	0	0	2	1	1	0	0	0	0	4
77 - 77.99	0	0	0	0	1	1	3	0	0	0	0	0	5
78 - 78.99	0	0	0	0	0	1	1	0	0	0	0	0	2
79 - 79.99	0	0	0	0	0	0	2	0	0	0	0	0	2
80 - 80.99	0	0	0	0	0	0	0	1	0	0	0	0	1
81 - 81.99	0	0	0	0	0	1	3	1	0	1	0	1	7
82 - 82.99	0	0	0	0	0	0	2	1	0	0	1	0	4
83 - 83.99	0	0	0	0	0	0	0	0	0	1	0	0	1
84 - 84.99	0	0	0	0	0	0	0	0	0	1	0	0	1
86 - 86.99	0	0	0	0	0	0	0	0	2	0	1	0	3
88 - 88.99	0	0	0	0	0	0	0	1	0	0	0	0	1
90 - 90.99	0	0	0	0	0	0	0	0	1	0	0	0	1
91 - 91.99	0	0	0	0	0	0	0	0	2	0	0	0	2
96 - 96.99	0	0	0	0	0	0	0	0	0	1	0	0	1
Totals	20	115	120	38	25	12	18	5	6	4	2	1	366

Table 3.3: Age-Length key, as proportion-at-age in each 1-cm length interval, based on otolith ages for bluefish sampled for age determination in Virginia during 2014.

(Go back to text)

						Age						
Interval	0	1	2	3	4	5	6	7	8	9	10	11
16 - 16.99	1	0	0	0	0	0	0	0	0	0	0	0
19 - 19.99	1	0	0	0	0	0	0	0	0	0	0	0
20 - 20.99	0.5	0.5	0	0	0	0	0	0	0	0	0	0
21 - 21.99	0.4	0.6	0	0	0	0	0	0	0	0	0	0
22 - 22.99	0.6	0.4	0	0	0	0	0	0	0	0	0	0
23 - 23.99	0.5	0.5	0	0	0	0	0	0	0	0	0	0
24 - 24.99	0.33	0.67	0	0	0	0	0	0	0	0	0	0
25 - 25.99	0	1	0	0	0	0	0	0	0	0	0	0
26 - 26.99	0	1	0	0	0	0	0	0	0	0	0	0
27 - 27.99	0	1	0	0	0	0	0	0	0	0	0	0
28 - 28.99	0	1	0	0	0	0	0	0	0	0	0	0
29 - 29.99	0	1	0	0	0	0	0	0	0	0	0	0
30 - 30.99	0	1	0	0	0	0	0	0	0	0	0	0
31 - 31.99	0	1	0	0	0	0	0	0	0	0	0	0
32 - 32.99	0	1	0	0	0	0	0	0	0	0	0	0
33 - 33.99	0	0.67	0.33	0	0	0	0	0	0	0	0	0
34 - 34.99	0	0.83	0.17	0	0	0	0	0	0	0	0	0
35 - 35.99	0	0.67	0.33	0	0	0	0	0	0	0	0	0
36 - 36.99	0	0.88	0.12	0	0	0	0	0	0	0	0	0
37 - 37.99	0	0.62	0.38	0	0	0	0	0	0	0	0	0
38 - 38.99	0	0.43	0.57	0	0	0	0	0	0	0	0	0
39 - 39.99	0	0.33	0.67	0	0	0	0	0	0	0	0	0
40 - 40.99	0	0.12	0.88	0	0	0	0	0	0	0	0	0
41 - 41.99	0	0.25	0.75	0	0	0	0	0	0	0	0	0
42 - 42.99	0	0.12	0.88	0	0	0	0	0	0	0	0	0
43 - 43.99	0	0	0.75	0.25	0	0	0	0	0	0	0	0
44 - 44.99	0	0.29	0.57	0.14	0	0	0	0	0	0	0	0
45 - 45.99	0	0.22	0.78	0	0	0	0	0	0	0	0	0
46 - 46.99	0	0.14	0.86	0	0	0	0	0	0	0	0	0
47 - 47.99	0	0.25	0.62	0.12	0	0	0	0	0	0	0	0
48 - 48.99	0	0.17	0.83	0	0	0	0	0	0	0	0	0
49 - 49.99	0	0.17	0.67	0.17	0	0	0	0	0	0	0	0
50 - 50.99	0	0.17	0.5	0.33	0	0	0	0	0	0	0	0
51 - 51.99	0	0.25	0.5	0.25	0	0	0	0	0	0	0	0
52 - 52.99	0	0.17	0.5	0.33	0	0	0	0	0	0	0	0
53 - 53.99	0	0.2	0.6	0.2	0	0	0	0	0	0	0	0
54 - 54.99	0	0.4	0.4	0.2	0	0	0	0	0	0	0	0
55 - 55.99	0	0	0.67	0.33	0	0	0	0	0	0	0	0
56 - 56.99	0	0	0.5	0.5	0	0	0	0	0	0	0	0
57 - 57.99	0	0	0.33	0.5	0.17	0	0	0	0	0	0	0
58 - 58.99	0	0	0.5	0.33	0.17	0	0	0	0	0	0	0

(To continue)

Table 3.3 (Continued)

						Age						
Interval	0	1	2	3	4	5	6	7	8	9	10	11
59 - 59.99	0	0	0.33	0.5	0.17	0	0	0	0	0	0	0
60 - 60.99	0	0	0.5	0.5	0	0	0	0	0	0	0	0
61 - 61.99	0	0	0.5	0.5	0	0	0	0	0	0	0	0
62 - 62.99	0	0	0.8	0	0.2	0	0	0	0	0	0	0
63 - 63.99	0	0	0.45	0.45	0	0	0.09	0	0	0	0	0
64 - 64.99	0	0	0.2	0	0.6	0	0.2	0	0	0	0	0
65 - 65.99	0	0	0.25	0	0.5	0.25	0	0	0	0	0	0
66 - 66.99	0	0	0	0.5	0.5	0	0	0	0	0	0	0
67 - 67.99	0	0	0	0	0	1	0	0	0	0	0	0
68 - 68.99	0	0	0	0	1	0	0	0	0	0	0	0
69 - 69.99	0	0	0	0	1	0	0	0	0	0	0	0
70 - 70.99	0	0	0	0	0.6	0.4	0	0	0	0	0	0
71 - 71.99	0	0	0	0	0.5	0.25	0.25	0	0	0	0	0
72 - 72.99	0	0	0	0.33	0.33	0	0.33	0	0	0	0	0
73 - 73.99	0	0	0	0	0.33	0.67	0	0	0	0	0	0
74 - 74.99	0	0	0	0	0	0	0.67	0	0.33	0	0	0
76 - 76.99	0	0	0	0	0	0.5	0.25	0.25	0	0	0	0
77 - 77.99	0	0	0	0	0.2	0.2	0.6	0	0	0	0	0
78 - 78.99	0	0	0	0	0	0.5	0.5	0	0	0	0	0
79 - 79.99	0	0	0	0	0	0	1	0	0	0	0	0
80 - 80.99	0	0	0	0	0	0	0	1	0	0	0	0
81 - 81.99	0	0	0	0	0	0.14	0.43	0.14	0	0.14	0	0.14
82 - 82.99	0	0	0	0	0	0	0.5	0.25	0	0	0.25	0
83 - 83.99	0	0	0	0	0	0	0	0	0	1	0	0
84 - 84.99	0	0	0	0	0	0	0	0	0	1	0	0
86 - 86.99	0	0	0	0	0	0	0	0	0.67	0	0.33	0
88 - 88.99	0	0	0	0	0	0	0	1	0	0	0	0
90 - 90.99	0	0	0	0	0	0	0	0	1	0	0	0
91 - 91.99	0	0	0	0	0	0	0	0	1	0	0	0
96 - 96.99	0	0	0	0	0	0	0	0	0	1	0	0

Chapter 4

COBIA Rachycentron canadum



4.1 INTRODUCTION

We aged a total of 287 cobia, *Rachycentron* canadum, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2014. Cobia ages ranged from 2 to 12 years old with an average age of 4.7, a standard deviation of 1.8, and a standard error of 0.11. Ten age classes (2 to 10, 12) were represented, comprising fish of the 2002, 2004 to 2012 year-classes. The sample was dominated by fish from the yearclass of 2010 with 54.4%.

4.2 METHODS

4.2.1 Handling of collections

Sagittal otoliths, hereafter, referred to as "otoliths", were received by the Age and Growth Laboratory in labeled coin envelopes and were sorted by date of capture, their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored inside of protective Axygen 2 ml micro-tubes within their original labeled coin envelopes.

4.2.2 Preparation

Otoliths were processed for age determination. The left or right otolith was randomly selected and embedded, distal side down, in epoxy resin and allowed to harden overnight. The otoliths were viewed by eye, and when necessary, under a stereo microscope to identify the location of the core, and the position of the core marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMetTM lowspeed saw equipped with two, three inch diameter, Norton Diamond Grinding Wheels, separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The position of the marked core fell within the 0.5 mm space between the blades, such that the core was included in the removed thin sec-Otolith thin-sections were placed tion. on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing cobia.

4.2.3 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the speciesspecific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, cobia otolith annulus deposition occurs between June and July (Richards 1967 and modified by CQFE). A cobia captured between January 1 and July 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of July 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 4.1). Each reader aged all of the otolith samples. All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again



Figure 4.1: Otolith thin-section of a 4 year-old cobia.

without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age cobia using their otolith thin-sections.

4.2.4 Comparison tests

A symmetry test (Hoenig et al. 1995)and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

4.3 RESULTS

4.3.1 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 94% and a CV of 1.7% (test of symmetry: $\chi^2 = 3$, df = 3, P = 0.3916), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 96% and a CV of 1% (test of symmetry: $\chi^2 = 2$, df = 2, P = 0.3679). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 95.12% and a CV of 0.9% (test of symmetry: $\chi^2 = 4.67$, df = 7, P = 0.7006) (Figure 4.2).



Figure 4.2: Between-reader comparison of otolith age estimates for cobia collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

There was no time-series bias for both readers. Reader 1 had an agreement of 86% with ages of fish aged in 2000 with a CV of 1.2% (test of symmetry: $\chi^2 = 5$, df = 6, P = 0.5438). Reader 2 had an agreement of 86% with a CV of 1.7% (test of symmetry: $\chi^2 = 7$, df = 7, P = 0.4289).

4.3.2 Year class

Of the 287 fish aged with otoliths, 10 age classes (2 to 10, 12) were represented (Table 4.1). The average age was 4.7 years, and the standard deviation and standard error were 1.8 and 0.11, respectively. Yearclass data show that the fishery was comprised of 10 year-classes: fish from the 2002, 2004 to 2012 year-classes, with fish primarily from the year class of 2010 with 54.4%. The ratio of males to females was 1:1.15 in the sample collected (Figure 4.3).



Figure 4.3: Year-class frequency distribution for cobia collected for ageing in 2014. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

4.3.3 Age-length key

We developed an age-length-key (Table 4.2) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

4.4 REFERENCES

- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analysing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.
- Richards, C.E. 1967. Age, Growth, and Fecundity of the Cobia, Rachycentron canadum, from Chesapeake Bay and Adjacent Waters. Contribution No. 252, Virginia Institute of Marine Science, Gloucester Point, Virginia, 343-350.

					Age						
Interval	2	3	4	5	6	7	8	9	10	12	Totals
35 - 35.99	3	2	1	0	0	0	0	0	0	0	6
36 - 36.99	1	1	4	0	0	0	0	0	0	0	6
37 - 37.99	0	8	13	1	0	0	0	0	0	0	22
38 - 38.99	1	10	15	0	0	0	0	0	0	0	26
39 - 39.99	0	5	17	1	0	1	0	0	0	0	24
40 - 40.99	0	7	15	0	0	1	0	0	0	0	23
41 - 41.99	0	2	14	1	1	0	0	0	0	0	18
42 - 42.99	0	2	15	2	2	3	0	0	0	0	24
43 - 43.99	0	2	6	1	4	3	0	0	0	0	16
44 - 44.99	0	1	8	0	0	2	1	0	0	0	12
45 - 45.99	0	0	14	1	1	2	1	1	0	0	20
46 - 46.99	0	2	11	2	1	3	1	2	0	1	23
47 - 47.99	0	0	9	2	1	1	0	0	1	0	14
48 - 48.99	0	0	7	0	0	0	0	0	1	1	9
49 - 49.99	0	0	4	0	1	1	0	0	0	0	6
50 - 50.99	0	0	1	3	1	1	1	0	0	0	7
51 - 51.99	0	0	1	0	0	0	0	0	1	0	2
52 - 52.99	0	0	0	1	1	1	0	0	0	0	3
53 - 53.99	0	0	0	1	1	0	0	0	0	0	2
54 - 54.99	0	0	0	0	0	5	0	0	0	0	5
55 - 55.99	0	0	0	0	2	3	1	0	0	0	6
56 - 56.99	0	0	0	0	1	1	0	1	0	0	3
57 - 57.99	0	0	0	0	0	0	1	1	1	0	3
58 - 58.99	0	0	0	0	0	0	0	1	1	0	2
59 - 59.99	0	0	1	0	0	0	0	1	0	0	2
60 - 60.99	0	0	0	0	0	0	1	1	0	0	2
62 - 62.99	0	0	0	0	0	0	0	0	1	0	1
Totals	5	42	156	16	17	28	7	8	6	2	287

Table 4.1: The number of cobia assigned to each total length (inch)-at-age category for 287 fish sampled for otolith age determination in Virginia during 2014.

(Go back to text)

					Age					
Interval	2	3	4	5	6	7	8	9	10	12
35 - 35.99	0.5	0.33	0.17	0	0	0	0	0	0	0
36 - 36.99	0.17	0.17	0.67	0	0	0	0	0	0	0
37 - 37.99	0	0.36	0.59	0.05	0	0	0	0	0	0
38 - 38.99	0.04	0.38	0.58	0	0	0	0	0	0	0
39 - 39.99	0	0.21	0.71	0.04	0	0.04	0	0	0	0
40 - 40.99	0	0.3	0.65	0	0	0.04	0	0	0	0
41 - 41.99	0	0.11	0.78	0.06	0.06	0	0	0	0	0
42 - 42.99	0	0.08	0.62	0.08	0.08	0.12	0	0	0	0
43 - 43.99	0	0.12	0.38	0.06	0.25	0.19	0	0	0	0
44 - 44.99	0	0.08	0.67	0	0	0.17	0.08	0	0	0
45 - 45.99	0	0	0.7	0.05	0.05	0.1	0.05	0.05	0	0
46 - 46.99	0	0.09	0.48	0.09	0.04	0.13	0.04	0.09	0	0.04
47 - 47.99	0	0	0.64	0.14	0.07	0.07	0	0	0.07	0
48 - 48.99	0	0	0.78	0	0	0	0	0	0.11	0.11
49 - 49.99	0	0	0.67	0	0.17	0.17	0	0	0	0
50 - 50.99	0	0	0.14	0.43	0.14	0.14	0.14	0	0	0
51 - 51.99	0	0	0.5	0	0	0	0	0	0.5	0
52 - 52.99	0	0	0	0.33	0.33	0.33	0	0	0	0
53 - 53.99	0	0	0	0.5	0.5	0	0	0	0	0
54 - 54.99	0	0	0	0	0	1	0	0	0	0
55 - 55.99	0	0	0	0	0.33	0.5	0.17	0	0	0
56 - 56.99	0	0	0	0	0.33	0.33	0	0.33	0	0
57 - 57.99	0	0	0	0	0	0	0.33	0.33	0.33	0
58 - 58.99	0	0	0	0	0	0	0	0.5	0.5	0
59 - 59.99	0	0	0.5	0	0	0	0	0.5	0	0
60 - 60.99	0	0	0	0	0	0	0.5	0.5	0	0
62 - 62.99	0	0	0	0	0	0	0	0	1	0

Table 4.2: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for cobia sampled for age determination in Virginia during 2014.

(Go back to text)

Chapter 5

RED DRUM Sciaenops ocellatus



5.1 INTRODUCTION

We aged a total of 80 red drum, *Sciaenops* ocellatus, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2014. Red drum ages ranged from 1 to 3 years old with an average age of 2.8, a standard deviation of 0.5, and a standard error of 0.06. Three age classes (1 to 3) were represented, comprising fish of the 2011 to 2013 year-classes. The sample was dominated by fish from the year-class of 2011 with 83.8%.

5.2 METHODS

5.2.1 Handling of collections

Sagittal otoliths, hereafter, referred to as "otoliths", were received by the Age and Growth Laboratory in labeled coin envelopes, and were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

5.2.2 Preparation

Otoliths were processed for age determination following the methods described in Ross et al. (1993) and Jones and Wells (1998) for red drum. The left or right sagittal otolith was randomly selected and attached, distal side down, to a glass slide with CrystalbondTM 509 adhesive. The otoliths were viewed by eye, and when necessary, under a stereo microscope to identify the location of the core, and the position of the core marked using a pencil across the otolith surface. At least one transverse cross-section (hereafter "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMetTM low-speed saw equipped with two, three inch diameter, Norton Diamond Grinding Wheels, separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The position of the marked core fell within the 0.5 mm space between the blades, such that the core was included in the removed thinsection. Otolith thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the sections.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing red drum.

5.2.3 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the speciesspecific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, red drum annulus formation occurs between March and June (Ross et al. 1995 and modified by CQFE). A red drum captured between January 1 and June 30, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of June 30, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 5.1). Each reader aged all of the otolith samples.

All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of pre-



Figure 5.1: Otolith thin-section of a 4 year-old red drum with the last annulus on the edge of the thin-section

viously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age red drum using their otolith thin-sections.

5.2.4 Comparison tests

A symmetry test (Hoenig et al. 1995)and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

5.3 RESULTS

5.3.1 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 100%, and there was no significant difference between the first and second readings for Reader 2 with an agreement of 100%. There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 100% (Figure 5.2).



Figure 5.2: Between-reader comparison of otolith age estimates for red drum collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

There was no time-series bias for both readers. Reader 1 had an agreement of 98% with ages of fish aged in 2000 with a CV of 0.4% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173). Reader 2 had an agreement of 98% with a CV of 0.1% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173).

5.3.2 Year class

Of the 80 fish aged with otoliths, 3 age classes (1 to 3) were represented (Table 5.1). The average age was 2.8 years, and the standard deviation and standard error

were 0.5 and 0.06, respectively. Year-class data show that the fishery was comprised of 3 year-classes: fish from the 2011 to 2013 year-classes, with fish primarily from the year class of 2011 with 83.8%. The ratio of males to females was 1:0.42 in the sample collected (Figure 5.3).



Figure 5.3: Year-class frequency distribution for red drum collected for ageing in 2014. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

5.3.3 Age-length key

We developed an age-length-key (Table 5.2) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

5.4 **REFERENCES**

- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analyzing differences be-

tween two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.

- R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.
- Ross, J., Stevens, T., Vaughan, D. 1995. Age, Growth, Mortality, and Reproductive Biology of Red Drums in North Carolina Waters. Trans. Amer. Fish. Soc. 124:37-54.

		Age		
Interval	1	2	3	Totals
17 - 17.99	1	0	0	1
18 - 18.99	2	0	1	3
19 - 19.99	1	0	1	2
20 - 20.99	0	0	2	2
21 - 21.99	0	0	9	9
22 - 22.99	0	1	11	12
23 - 23.99	0	4	9	13
24 - 24.99	0	3	13	16
25 - 25.99	0	0	8	8
26 - 26.99	0	1	10	11
27 - 27.99	0	0	2	2
29 - 29.99	0	0	1	1
Totals	4	9	67	80

Table 5.1: The number of red drum assigned to each total length (inch)-at-age category for 80 fish sampled for otolith age determination in Virginia during 2014.

(Go back to text)

	Age		
Interval	1	2	3
17 - 17.99	1	0	0
18 - 18.99	0.67	0	0.33
19 - 19.99	0.5	0	0.5
20 - 20.99	0	0	1
21 - 21.99	0	0	1
22 - 22.99	0	0.08	0.92
23 - 23.99	0	0.31	0.69
24 - 24.99	0	0.19	0.81
25 - 25.99	0	0	1
26 - 26.99	0	0.09	0.91
27 - 27.99	0	0	1
29 - 29.99	0	0	1

Table 5.2: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for red drum sampled for age determination in Virginia during 2014.

(Go back to text)

Chapter 6

SHEEPSHEAD Archosargus probatocephalus



6.1 INTRODUCTION

We aged a total of 120 sheepshead, Archosargus probatocephalus, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2014. Sheepshead ages ranged from 2 to 28 years old with an average age of 8.5, a standard deviation of 6.6, and a standard error of 0.6. Twenty age classes (2 to 9, 12 to 13, 15 to 18, 20 to 21, 23 to 24, 26, 28) were represented, comprising fish of the 1986, 1988, 1990 to 1991, 1993 to 1994, 1996 to 1999, 2001 to 2002, 2005 to 2012 year-classes. The sample was dominated by fish from the year-class of 2011 with 30.8%.

6.2 METHODS

6.2.1 Handling of collections

Sagittal otoliths, hereafter, referred to as "otoliths", were received by the Age and Growth Laboratory in labeled coin envelopes, and were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

6.2.2 Preparation

Otoliths were processed for age determination following the methods described in Ballenger et al. (in progress). The left or right otolith was randomly selected and embedded, distal side down, in epoxy resin and allowed to harden overnight. The otoliths were viewed by eye, and when necessary, under a stereo microscope to identify the location of the core, and the position of the core marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMetTM low-speed saw equipped with two, three inch diameter, Norton Diamond Grinding Wheels, separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The position of the marked core fell within the 0.5 mm space between the blades, such that the core was included in the removed thin section. Otolith thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing sheepshead.

6.2.3 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the speciesspecific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, sheepshead otolith annulus formation occurs between May and June (Ballenger et al. in progress). A sheepshead captured between January 1 and July 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 6.1). Each reader aged all of the otolith samples. All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When



Figure 6.1: Otolith thin-section of a 5 year-old sheepshead

the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age sheepshead using their otolith thin-sections.

6.2.4 Comparison tests

A symmetry test (Hoenig et al. 1995)and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

6.3 RESULTS

6.3.1 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 88% and a CV of 1.3% (test of symmetry: $\chi^2 = 4$, df = 5, P = 0.5494), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 76% and a CV of 3% (test of symmetry: $\chi^2 = 9$, df = 7, P = 0.2527). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 75.83% and a CV of 2.1% (test of symmetry: $\chi^2 = 21.67$, df = 15, P = 0.1168) (Figure 6.2).



Figure 6.2: Between-reader comparison of otolith age estimates for sheepshead collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

There was no time-series bias for both readers. Reader 1 had an agreement of 98% with ages of fish aged in 2008 with a CV of 0.1% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173). Reader 2 had an agreement of 100%.

6.3.2 Year class

Of the 120 fish aged with otoliths, 20 age classes (2 to 9, 12 to 13, 15 to 18, 20 to 21,

23 to 24, 26, 28) were represented (Table 6.1). The average age was 8.5 years, and the standard deviation and standard error were 6.6 and 0.6, respectively. Year-class data show that the fishery was comprised of 20 year-classes: fish from the 1986, 1988, 1990 to 1991, 1993 to 1994, 1996 to 1999, 2001 to 2002, 2005 to 2012 year-classes, with fish primarily from the year class of 2011 with 30.8%. The ratio of males to females was 1:0.65 in the sample collected (Figure 6.3).



Figure 6.3: Year-class frequency distribution for sheepshead collected for ageing in 2014. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

6.3.3 Age-length key

We developed an age-length-key (Table 6.2) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

6.4 REFERENCES

Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.

- Ballenger, J. H. Liao, and C.M. Jones. (in progress). Length-weight, Age and Growth of Sheepshead, Archosargus probatocephalus (Pisces: Sparidae), from the Chesapeake Bay region, Virginia: A Comparison to other Areas. Fisheries Bulletin.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analyzing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.

										Age											
Interval	2	ω	4	сл	6	7	∞	9	12	13	15	16	17	18	20	21	23	24	26	28	Total
10 - 10.99	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
11 - 11.99	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
12 - 12.99	0	4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2
13 - 13.99	0	ы	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
14 - 14.99	0	7	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
15 - 15.99	0	12	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
16 - 16.99	0	4	⊢	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
17 - 17.99	0	4	0	0	ω	0		0	0	0	0	0	0	0	0	0	0	0	0	0	
18 - 18.99	0	1	Ļ	ಲು	0	4	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1
19 - 19.99	0	0	0	Ļ	1	Ċī	ယ	ယ	0	0	0	0	0	0	0	0	0	0	0	0	1
20 - 20.99	0	0	0	0	0	4	4	1	0	0	0	0	0		0	0	0	0	0	0	1
21 - 21.99	0	0	0	0	2	2	1	1	0	0	0	0	0	0	0	0	0	0	0	0	
22 - 22.99	0	0	0	0	1	1	2	ယ	1	ယ	0	1	1	0	0	0	0	1	1	0	1
23 - 23.99	0	0	0	0	0	0	0	⊢	0	0	1	Ľ	ಬ	0	2	Ļ	0	⊢	0	⊢	1
24 - 24.99	0	0	0	0	0	0	0	0	0		0	1	0	0	0	0	0	1	1	⊢	
25 - 25.99	0	0			>			D	C	þ	>	-	2	-	>	>	-				
		0	(\subset	\subset		\subset	\subset	\bigcirc		C	⊢	N	F			L	0	0		

н.	
ä	
tic	
ina	
E.	
te	
de	
e Ge	
r a	
fo	
ed	
lqı	
an	
с С	
lea	
dsc	
eel	
$^{\mathrm{sh}}$	
or	
ŝ	
ge	
Ъ	
lit	
oto	
ų	
0 7	
sec	
ba	
al,	
IV	
nte	
л. Л	
gt]	
len	
ų	
й.	
4	
ch	
eа	
п.	
ge	
t-a	
l-a	
ior	
ort	
do	
\mathbf{pr}	
as	
y,	
\mathbf{ke}	
th	÷
gue	01_{-1}
-Ľ	5
é	ing
A	lur
.2	a c
e G	ini
ldt	irg
Ĥ	\geq

	28	0	0	0	0	0	0	0	0	0	0	0	0	0	0.09	0.2	0	
	26	0	0	0	0	0	0	0	0	0	0	0	0	0.07	0	0.2	0	
	24	0	0	0	0	0	0	0	0	0	0	0	0	0.07	0.09	0.2	0	
	23	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0.2	
	21	0	0	0	0	0	0	0	0	0	0	0	0	0	0.09	0	0	
	20	0	0	0	0	0	0	0	0	0	0	0	0	0	0.18	0	0	
	18	0	0	0	0	0	0	0	0	0	0	0.1	0	0	0	0	0.2	
	17	0	0	0	0	0	0	0	0	0	0	0	0	0.07	0.27	0	0.4	
	16	0	0	0	0	0	0	0	0	0	0	0	0	0.07	0.09	0.2	0.2	
	15	0	0	0	0	0	0	0	0	0	0	0	0	0	0.09	0	0	
Age	13	0	0	0	0	0	0	0	0	0	0	0	0	0.2	0	0.2	0	
	12	0	0	0	0	0	0	0	0	0	0	0	0	0.07	0	0	0	
	6	0	0	0	0	0	0	0	0	0	0.23	0.1	0.17	0.2	0.09	0	0	
	∞	0	0	0	0	0	0	0	0.12	0.1	0.23	0.4	0.17	0.13	0	0	0	
	2	0	0	0	0	0	0	0	0	0.4	0.38	0.4	0.33	0.07	0	0	0	
	9	0	0	0	0	0	0	0	0.38	0	0.08	0	0.33	0.07	0	0	0	
	ъ	0	0	0	0	0	0	0	0	0.3	0.08	0	0	0	0	0	0	
	4	0	0	0	0	0	0	0.2	0	0.1	0	0	0	0	0	0	0	
	က	0	0	1	1	1	1	0.8	0.5	0.1	0	0	0	0	0	0	0	
	7		μ	0	0	0	0	0	0	0	0	0	0	0	0	0	0	ext)
	Interval	10 - 10.99	11 - 11.99	12 - 12.99	13 - 13.99	14 - 14.99	15 - 15.99	16 - 16.99	17 - 17.99	18 - 18.99	19 - 19.99	20 - 20.99	21 - 21.99	22 - 22.99	23 - 23.99	24 - 24.99	25 - 25.99	Go back to t

CHAPTER 6. SHEEPSHEAD ARCHOSARGUS PROBATOCEPHALUS

Chapter 7

ATLANTIC SPADEFISH Chaetodipterus faber



7.1 INTRODUCTION

We aged a total of 288 spadefish, *Chaetodipterus faber*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2014. Spadefish ages ranged from 0 to 7 years old with an average age of 3, a standard deviation of 1.4, and a standard error of 0.08. Eight age classes (0 to 7) were represented, comprising fish of the 2007 to 2014 year-classes. The sample was dominated by fish from the year-classes of 2012 and 2010 with 48.6% and 30.2%, respectively.

7.2 METHODS

7.2.1 Sample size for ageing

We estimated sample size for ageing spadefish in 2014 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 C V^2 + B_a/L} \tag{7.1}$$

where A is the sample size for ageing spadefish in 2014; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of spadefish used by VMRC to estimate length distribution of the catches from 2008 to 2012. θ_a , V_a , B_a , and CV were calculated using pooled agelength data of spadefish collected from 2008 to 2012 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (7.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A

should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2008 to 2012 catch. A_l is number of fish to be aged for length interval l in 2014.

7.2.2 Handling of collections

Otoliths were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

7.2.3 Preparation

We used our thin-section and bake technique to process spadefish sagittal otoliths (hereafter, referred to as "otoliths") for age determination. Otolith preparation began by randomly selecting either the right or left otolith. Each whole otolith was placed in a ceramic "Coors" spot plate well and baked in a Thermolyne 1400 furnace at 400 °C. Baking time was dependent on the otolith's size and gauged by color, with a light caramel color desired. Once a suitable color was achieved the baked otolith was embedded in epoxy resin with its distal surface orientated downwards and allowed to harden overnight. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith

using a Buehler IsoMetTM low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The otolith was positioned so the blades straddled each side of the otolith focus. It was crucial that this cut be perpendicular to the long axis of the otolith. Failure to do so resulted in broadening and distored winter growth zones. A proper cut resulted in annuli that were clearly defined and delineated. Once cut, thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thinsection.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing Atlantic spade-fish.

7.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the speciesspecific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, spadefish otolith annulus formation occurs between December and July (Hayse 1989 and modified by CQFE). A spadefish captured between January 1 and July 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of July 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 7.1). Each reader aged all of the otolith samples. All samples were aged in chronological order, based on collection date, without knowledge of



Figure 7.1: Otolith thin-section of a 2 year-old spadefish

previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age Atlantic spadefish using their otolith thinsections.

7.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995)and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

7.3 RESULTS

7.3.1 Sample size

We estimated a sample size of 337 spadefish in 2014, ranging in length interval from 3 to 25 inches (Table 7.1). This sample size provided a range in (CV) for age composition approximately from the smallest (CV) of 6% for age 2 to the largest (CV)of 21% for age 6. In 2014, we aged 288 of 323 spadefish (The rest of fish were either without otoliths or over-collected for certain length interval(s)) collected by VMRC. We fell short in our over-all collections for this optimal length-class sampling estimate by 64 fish. However, we were short of some fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would possibly be influenced significantly.

7.3.2 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 98% and a CV of 0.3% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 98% and a CV of 0.3% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement between Reader 1 and Reader 2 with an agreement of 92.01% and a CV of 2.4% (test of symmetry: $\chi^2 = 7.53$, df = 8, P = 0.4803) (Figure 7.2).

There was no time-series bias for both readers. Reader 1 had an agreement of 96% with ages of fish aged in 2003 with a CV of 0.3% (test of symmetry: $\chi^2 = 2$, df = 2, P = 0.3679). Reader 2 had an agreement of



Figure 7.2: Between-reader comparison of otolith age estimates for spadefish collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

98% with a *CV* of 0.2% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173).

7.3.3 Year class

Of the 288 fish aged with otoliths, 8 age classes (0 to 7) were represented (Table 7.2). The average age was 3 years, and the standard deviation and standard error were 1.4 and 0.08, respectively. Year-class data show that the fishery was comprised of 8 year-classes: fish from the 2007 to 2014 year-classes, with fish primarily from the year classes of 2012 and 2010 with 48.6% and 30.2%, respectively. The ratio of males to females was 1:1.1 in the sample collected (Figure 7.3).

7.3.4 Age-length key

We developed an age-length-key (Table 7.3) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.



Figure 7.3: Year-class frequency distribution for spadefish collected for ageing in 2014. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

7.4 REFERENCES

- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.
- Hayse, J. W. 1989. Feeding habits, age, growth, and reproduction of Atlantic spadefish Chaetodipterus faber (Piscies: Ephippidae) in South Carolina. Fishery Bulletin 88: 67-83.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analyzing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- Quinn, T. J. II, and R. B. Deriso. 1999. Quantitative Fish Dynamics. Oxford University Press. New York.
- R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.

Table 7.1: Number of Atlantic spadefish collected and aged in each 1-inch length interval in 2014.
'Target' represents the sample size for ageing estimated for 2014, and 'Need' represents number of
fish shorted in each length interval compared to the optimum sample size for ageing and number of
fish aged.

Interval	Target	Collected	Aged	Need
3 - 3.99	5	0	0	5
4 - 4.99	5	6	6	0
5 - 5.99	10	22	14	0
6 - 6.99	39	53	39	0
7 - 7.99	46	51	51	0
8 - 8.99	34	30	30	4
9 - 9.99	25	20	20	5
10 - 10.99	20	13	13	7
11 - 11.99	14	16	16	0
12 - 12.99	21	20	20	1
13 - 13.99	22	10	10	12
14 - 14.99	14	14	14	0
15 - 15.99	14	18	14	0
16 - 16.99	10	18	10	0
17 - 17.99	15	19	18	0
18 - 18.99	10	8	8	2
19 - 19.99	8	4	4	4
20 - 20.99	5	1	1	4
21 - 21.99	5	0	0	5
22 - 22.99	5	0	0	5
23 - 23.99	5	0	0	5
25 - 25.99	5	0	0	5
Totals	337	323	288	64

(Go back to text)
				Age					
Interval	0	1	2	3	4	5	6	7	Totals
4 - 4.99	5	1	0	0	0	0	0	0	6
5 - 5.99	1	5	8	0	0	0	0	0	14
6 - 6.99	0	0	39	0	0	0	0	0	39
7 - 7.99	0	0	49	2	0	0	0	0	51
8 - 8.99	0	0	26	4	0	0	0	0	30
9 - 9.99	0	0	14	5	1	0	0	0	20
10 - 10.99	0	0	4	2	7	0	0	0	13
11 - 11.99	0	0	0	4	12	0	0	0	16
12 - 12.99	0	0	0	4	16	0	0	0	20
13 - 13.99	0	0	0	2	8	0	0	0	10
14 - 14.99	0	0	0	2	12	0	0	0	14
15 - 15.99	0	0	0	0	12	1	1	0	14
16 - 16.99	0	0	0	0	9	0	0	1	10
17 - 17.99	0	0	0	0	10	3	2	3	18
18 - 18.99	0	0	0	0	0	2	4	2	8
19 - 19.99	0	0	0	0	0	2	2	0	4
20 - 20.99	0	0	0	0	0	0	0	1	1
Totals	6	6	140	25	87	8	9	7	288

Table 7.2: The number of Atlantic spadefish assigned to each total length-at-age category for 288 fish sampled for otolith age determination in Virginia during 2014.

(Go back to text)⁼

				Age				
Interval	0	1	2	3	4	5	6	7
4 - 4.99	0.83	0.17	0	0	0	0	0	0
5 - 5.99	0.07	0.36	0.57	0	0	0	0	0
6 - 6.99	0	0	1	0	0	0	0	0
7 - 7.99	0	0	0.96	0.04	0	0	0	0
8 - 8.99	0	0	0.87	0.13	0	0	0	0
9 - 9.99	0	0	0.7	0.25	0.05	0	0	0
10 - 10.99	0	0	0.31	0.15	0.54	0	0	0
11 - 11.99	0	0	0	0.25	0.75	0	0	0
12 - 12.99	0	0	0	0.2	0.8	0	0	0
13 - 13.99	0	0	0	0.2	0.8	0	0	0
14 - 14.99	0	0	0	0.14	0.86	0	0	0
15 - 15.99	0	0	0	0	0.86	0.07	0.07	0
16 - 16.99	0	0	0	0	0.9	0	0	0.1
17 - 17.99	0	0	0	0	0.56	0.17	0.11	0.17
18 - 18.99	0	0	0	0	0	0.25	0.5	0.25
19 - 19.99	0	0	0	0	0	0.5	0.5	0
20 - 20.99	0	0	0	0	0	0	0	1

Table 7.3: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for Atlantic spadefish sampled for age determination in Virginia during 2014.

(Go back to $\overline{\text{text}}$)

Chapter 8

SPANISH MACKEREL Scomberomorous maculatus



8.1 INTRODUCTION

We aged a total of 236 Spanish mackerel, Scomberomorous maculatus, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2014. Spanish mackerel ages ranged from 0 to 6 years old with an average age of 1.5, a standard deviation of 0.8, and a standard error of 0.05. Seven age classes (0 to 6) were represented, comprising fish of the 2008 to 2014 year-classes. The sample was dominated by fish from the year-classes of 2013 and 2012 with 63.6% and 25.4%, respectively.

8.2 METHODS

8.2.1 Sample size for ageing

We estimated sample size for ageing Spanish mackerel in 2014 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 C V^2 + B_a/L} \tag{8.1}$$

where A is the sample size for ageing Spanish mackerel in 2014; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; Lwas the total number of Spanish mackerel used by VMRC to estimate length distribution of the catches from 2008 to 2012. θ_a , V_a , B_a , and CV were calculated using pooled age-length data of Spanish mackerel collected from 2008 to 2012 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (8.1) indicates that the more fish that are aged, the smaller the CV(or higher precision) that will be obtained.

Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2008 to 2012 catch. A_l is number of fish to be aged for length interval l in 2014.

8.2.2 Handling of collections

Otoliths were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

8.2.3 Preparation

Sagittal otoliths, hereafter, referred to as "otolith", were processed for age determination. The left or right otolith was randomly selected and embedded, distal side down, in epoxy resin and allowed to harden overnight. The otoliths were viewed by eye, and when necessary, under a stereo microscope to identify the location of the core, and the position of the core marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter "thin-section") was then removed from the marked core of each otolith using a Buehler $IsoMet^{TM}$ lowspeed saw equipped with two, three inch diameter, Norton Diamond Grinding Wheels, separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The position of the marked core fell within the 0.5 mm space between the blades, such that the core was included in the removed thin sec-Otolith thin-sections were placed tion.

on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing Spanish mackerel.

8.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the speciesspecific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, Spanish mackerel annulus formation occurs between May and June (Schmidt et al. 1993). A Spanish mackerel captured between January 1 and June 30, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 8.1). Each reader aged all of the otolith samples. All sam-



Figure 8.1: Otolith thin-section of a 3 year-old Spanish mackerel with the last annulus on the edge of the thin-section

ples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age Spanish mackerel using their otolith thinsections.

8.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995)and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

8.3 RESULTS

8.3.1 Sample size

We estimated a sample size of 304 Spanish mackerel in 2014, ranging in length interval from 8 to 31 inches (Table 8.1). This sample size provided a range in (CV) for age

composition approximately from the smallest (CV) of 4% for age 1 to the largest (CV) of 23% for age 0. In 2014, we aged 236 of 253 Spanish mackerel (The rest of fish were either without otoliths or overcollected for certain length interval(s)) collected by VMRC. We fell short in our overall collections for this optimal length-class sampling estimate by 76 fish. However, we were short of some fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would possibly be influenced significantly.

8.3.2 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 98% and a CV of 0.6% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 100%. There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 95.76% and a CV of 1.4% (test of symmetry: $\chi^2 = 6.8$, df = 3, P = 0.0786) (Figure 8.2).



Figure 8.2: Between-reader comparison of otolith age estimates for Spanish mackerel collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

There was no time-series bias for both readers. Reader 1 had an agreement of 96% with fish aged in 2003 with a CV of 1.9% (test of symmetry: $\chi^2 = 0$, df = 1, P = 1). Reader 2 had an agreement of 98% with a CV of 0.3% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173).

8.3.3 Year class

Of the 236 fish aged with otoliths, 7 age classes (0 to 6) were represented (Table 8.2). The average age was 1.5 years, and the standard deviation and standard error were 0.8 and 0.05, respectively. Year-class data show that the fishery was comprised of 7 year-classes: fish from the 2008 to 2014 year-classes, with fish primarily from the year classes of 2013 and 2012 with 63.6% and 25.4%, respectively. The ratio of males to females was 1:1.96 in the sample collected (Figure 8.3).



Figure 8.3: Year-class frequency distribution for Spanish mackerel collected for ageing in 2014. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

8.3.4 Age-length key

We developed an age-length-key (Table 8.3) that can be used in the conversion of numbers-at-length in the estimated catch

to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

8.4 REFERENCES

- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for etermining the consistency of age terminations. Trans. Am. Fish. Soc. 124:131-138.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analyzing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- Quinn, T. J. II, and R. B. Deriso. 1999. Quantitative Fish Dynamics. Oxford University Press. New York.
- R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.
- Schmidt, D., Collins, M., Wyanski, D. 1993. Age, growth, maturity, and spawning of Spanish mackerel from the Atlantic coast of the southeastern United States. Fishery Bulletin 91s.

Table 8.1: Number of Spanish mackerel collected and aged in each 1-inch length interval in 2014.
'Target' represents the sample size for ageing estimated for 2014, and 'Need' represents number of
fish shorted in each length interval compared to the optimum sample size for ageing and number of
fish aged.

Interval	Target	Collected	Aged	Need
8 - 8.99	5	0	0	5
9 - 9.99	5	0	0	5
10 - 10.99	5	0	0	5
11 - 11.99	5	0	0	5
12 - 12.99	5	0	0	5
13 - 13.99	6	3	3	3
14 - 14.99	20	11	11	9
15 - 15.99	42	39	39	3
16 - 16.99	45	48	48	0
17 - 17.99	40	41	41	0
18 - 18.99	22	26	22	0
19 - 19.99	18	23	18	0
20 - 20.99	15	19	16	0
21 - 21.99	15	21	16	0
22 - 22.99	11	7	7	4
23 - 23.99	5	6	6	0
24 - 24.99	5	6	6	0
25 - 25.99	5	2	2	3
26 - 26.99	5	0	0	5
27 - 27.99	5	0	0	5
28 - 28.99	5	0	0	5
29 - 29.99	5	1	1	4
30 - 30.99	5	0	0	5
31 - 31.99	5	0	0	5
Totals	304	253	236	76

				Age				
Interval	0	1	2	3	4	5	6	Totals
13 - 13.99	1	2	0	0	0	0	0	3
14 - 14.99	0	11	0	0	0	0	0	11
15 - 15.99	0	38	1	0	0	0	0	39
16 - 16.99	1	47	0	0	0	0	0	48
17 - 17.99	0	37	4	0	0	0	0	41
18 - 18.99	0	14	8	0	0	0	0	22
19 - 19.99	0	1	16	1	0	0	0	18
20 - 20.99	0	0	14	1	1	0	0	16
21 - 21.99	0	0	10	6	0	0	0	16
22 - 22.99	0	0	6	0	0	1	0	7
23 - 23.99	0	0	1	2	3	0	0	6
24 - 24.99	0	0	0	5	1	0	0	6
25 - 25.99	0	0	0	2	0	0	0	2
29 - 29.99	0	0	0	0	0	0	1	1
Totals	2	150	60	17	5	1	1	236

Table 8.2: The number of Spanish mackerel assigned to each total length-at-age category for 236 fish sampled for otolith age determination in Virginia during 2014.

			Age				
Interval	0	1	2	3	4	5	6
13 - 13.99	0.33	0.67	0	0	0	0	0
14 - 14.99	0	1	0	0	0	0	0
15 - 15.99	0	0.97	0.03	0	0	0	0
16 - 16.99	0.02	0.98	0	0	0	0	0
17 - 17.99	0	0.9	0.1	0	0	0	0
18 - 18.99	0	0.64	0.36	0	0	0	0
19 - 19.99	0	0.06	0.89	0.06	0	0	0
20 - 20.99	0	0	0.88	0.06	0.06	0	0
21 - 21.99	0	0	0.62	0.38	0	0	0
22 - 22.99	0	0	0.86	0	0	0.14	0
23 - 23.99	0	0	0.17	0.33	0.5	0	0
24 - 24.99	0	0	0	0.83	0.17	0	0
25 - 25.99	0	0	0	1	0	0	0
29 - 29.99	0	0	0	0	0	0	1

Table 8.3: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for Spanish mackerel sampled for age determination in Virginia during 2014.

(Go back to text)⁼

Chapter 9

SPOT Leiostomus xanthurus



9.1 INTRODUCTION

We aged a total of 276 spot, *Leiostomus xanthurus*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2014. spot ages ranged from 0 to 3 years old with an average age of 1.7, a standard deviation of 0.5, and a standard error of 0.03. Four age classes (0 to 3) were represented, comprising fish of the 2011 to 2014 year-classes. The sample was dominated by fish from the year-class of 2012 with 65.6%.

9.2 METHODS

9.2.1 Sample size for ageing

We estimated sample size for ageing spot in 2014 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 C V^2 + B_a/L} \tag{9.1}$$

where A is the sample size for ageing spot in 2014; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CVis the coefficient of variation; L was the total number of spot used by VMRC to estimate length distribution of the catches from 2008 to 2012. θ_a , V_a , B_a , and CV were calculated using pooled age-length data of spot collected from 2008 to 2012 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (1.1) indicates that the more fish that are aged, the smaller the CV(or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for

the most major age in catch by ageing an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2008 to 2012 catch. A_l is number of fish to be aged for length interval l in 2014.

9.2.2 Handling of collections

Otoliths were received by the Age & and Growth Laboratory in labeled coin envelopes, were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

9.2.3 Preparation

Sagittal otoliths, hereafter, referred to as "otoliths", were processed for age determination following the methods described in Barbieri et al. (1994) with a few modifications. The left or right otolith was randomly selected and embedded (distal side down) in epoxy resin and allowed to harden overnight. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMetTM low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). Thinsections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-sections.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing spot.

9.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured be-

tween January 1 and the end of the speciesspecific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, spot otolith annulus formation occurs between May and July (Piner and Jones 2004). A spot captured between January 1 and May 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 9.1). Each reader aged all of the otolith samples. Due to dis-



Figure 9.1: Otolith thin-section of a 2 year-old spot

crepancy on identification of the first annulus of spot among Atlantic states, Atlantic States Marine Fisheries Commission (ASMFC) has decided not to count the smallest annulus at the center of the thinsection as the first annulus. Following ASMFC's instruction, we didn't count the smallest annulus at the center as the first annulus in 2014. All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age spot using their otolith thin-sections.

9.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995)and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

9.3 RESULTS

9.3.1 Sample size

We estimated a sample size of 271 spot in 2014, ranging in length interval from 5 to 12 inches (Table 9.1). This sample size provided a range in (CV) for age composition approximately from the smallest (CV) of 5% for age 1 to the largest (CV) of 8% for age 2. In 2014, we randomly selected and aged 276 fish from 336 spot collected by VMRC. We fell short in our over-all collections for this optimal length-class sampling estimate by 5 fish. However, we were short of no fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

9.3.2 Reading precision

Reader 1 had moderate self-precision and Read 2 had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 88% and a CV of 8.7% (test of symmetry: $\chi^2 = 4$, df= 3, P = 0.2615), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 98% and a CV of 0.9% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173). There was an evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 88.77% and a CV of 4.3% (test of symmetry: $\chi^2 = 25.8, df = 3,$ P = 0 (Figure 9.2). This was due to the disagreement between two readers on the ring between the first and the second annulus. Reader 1 considered some of them as actual annuli while Reader 2 didn't.

There was no time-series bias for both readers. Reader 1 had an agreement of 98% with ages of fish aged in 2003 with a CV of



Figure 9.2: Between-reader comparison of otolith age estimates for spot collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

0.9% (test of symmetry: $\chi^2=1,\;df=1,\;P=0.3173).$ Reader 2 had an agreement of 100% .

9.3.3 Year class

Of the 276 fish aged with otoliths, 4 age classes (0 to 3) were represented (Table 9.2). The average age was 1.7 years, and the standard deviation and standard error were 0.5 and 0.03, respectively. Year-class data show that the fishery was comprised of 4 year-classes: fish from the 2011 to 2014 year-classes, with fish primarily from the year class of 2012 with 65.6%. The ratio of males to females was 1:5.71 in the sample collected (Figure 9.3).

9.3.4 Age-length key

We developed an age-length-key (Table 9.3) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.



Figure 9.3: Year-class frequency distribution for spot collected for ageing in 2014. Distribution is broken down by sex. 'Unknown' is for gonads that were not available for examination or were not examined for sex during sampling.

9.4 REFERENCES

- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analyzing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- Piner, K. R., and C. M. Jones. 2004. Age, growth and the potential for growth overfishing of spot (Leiostomus xanthurus) from the Chesapeake Bay, eastern USA. Marine and Freshwater Research 55: 553-560.
- Quinn, T. J. II, and R. B. Deriso. 1999. Quantitative Fish Dynamics. Oxford University Press. New York.
- R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.

Table 9.1: Number of spot collected and aged in each 1-inch length interval in 2014. 'Target' represents the sample size for ageing estimated for 2014, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
5 - 5.99	6	7	6	0
6 - 6.99	6	19	6	0
7 - 7.99	31	36	32	0
8 - 8.99	59	68	60	0
9 - 9.99	85	110	86	0
10 - 10.99	61	72	63	0
11 - 11.99	18	24	23	0
12 - 12.99	5	0	0	5
Totals	271	336	276	5

		Age			
Interval	0	1	2	3	Totals
5 - 5.99	5	1	0	0	6
6 - 6.99	0	6	0	0	6
7 - 7.99	0	23	9	0	32
8 - 8.99	0	15	43	2	60
9 - 9.99	0	13	73	0	86
10 - 10.99	0	27	36	0	63
11 - 11.99	0	3	20	0	23
Totals	5	88	181	2	276

Table 9.2: The number of spot assigned to each total length-at-age category for 276 fish sampled for otolith age determination in Virginia during 2014.

		Age		
Interval	0	1	2	3
5 - 5.99	0.83	0.17	0	0
6 - 6.99	0	1	0	0
7 - 7.99	0	0.72	0.28	0
8 - 8.99	0	0.25	0.72	0.03
9 - 9.99	0	0.15	0.85	0
10 - 10.99	0	0.43	0.57	0
11 - 11.99	0	0.13	0.87	0

Table 9.3: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for spot sampled for age determination in Virginia during 2014.

Chapter 10

SPOTTED SEATROUT Cynoscion nebulosus



10.1 INTRODUCTION

We aged a total of 273 spotted seatrout, *Cynoscion nebulosus*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2014. Spotted seatrout ages ranged from 0 to 8 years old with an average age of 2.1, a standard deviation of 1.5, and a standard error of 0.09. Eight age classes (0 to 6, 8) were represented, comprising fish of the 2006, 2008 to 2014 year-classes. The sample was dominated by fish from the year-classes of 2013 and 2011 with 30% and 23.1%, respectively.

10.2 METHODS

10.2.1 Sample size for ageing

We estimated sample size for ageing spotted seatrout in 2014 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 C V^2 + B_a/L} \tag{10.1}$$

where A is the sample size for ageing spotted seatrout in 2014; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of spotted seatrout used by VMRC to estimate length distribution of the catches from 2008 to 2012. θ_a , V_a , B_a , and CV were calculated using pooled age-length data of spotted seatrout collected from 2008 to 2012 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (10.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a $1\% \ CV$ reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2008 to 2012 catch. A_l is number of fish to be aged for length interval l in 2014.

10.2.2 Handling of collections

Otoliths were received by the Age & Growth Laboratory in labeled coin envelopes. In the lab they were sorted by date of capture, their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

10.2.3 Preparation

Sagittal otoliths, hereafter, referred to as "otoliths", were processed for age determination. The left or right otolith was randomly selected and attached, distal side down, to a glass slide with clear CrystalbondTM 509 adhesive. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a pencil across the otolith surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMetTM low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). Thinsections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-sections.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing spotted seatrout.

10.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the speciesspecific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, spotted seatrout otolith annulus formation occurs between March and May (Ihde and Chittenden 2003). A spotted seatrout captured between January 1 and May 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 10.1). Each reader aged all of the otolith samples. All sam-



Figure 10.1: Otolith thin-section of a 4 yearold spotted seatrout with the last annulus on the edge of the thin-section

ples were aged in chronological order, based

on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age spotted seatrout using their otolith thinsections.

10.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995)and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

10.3 RESULTS

10.3.1 Sample size

We estimated a sample size of 319 spotted seatrout in 2014, ranging in length interval from 6 to 34 inches (Table 10.1). This sample size provided a range in (CV) for age composition approximately from the smallest (CV) of 5% for age 1 to the largest (CV)of 25% for age 4. In 2014, we randomly selected and aged 273 fish from 362 spotted seatrout collected by VMRC. We fell short in our over-all collections for this optimal length-class sampling estimate by 57 fish. However, we were short of few fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

10.3.2 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 100%, and there was no significant difference between the first and second readings for Reader 2 with an agreement of 100%. There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 99.63% and a CV of 0.2% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173) (Figure 10.2).

There was no time-series bias for both readers. Reader 1 had an agreement of 100% with ages of fish aged in 2003. Reader 2 had an agreement of 100%.

10.3.3 Year class

Of the 273 fish aged with otoliths, 8 age classes (0 to 6, 8) were represented (Table 10.2). The average age was 2.1 years,



Figure 10.2: Between-reader comparison of otolith age estimates for spotted seatrout collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

and the standard deviation and standard error were 1.5 and 0.09, respectively. Yearclass data show that the fishery was comprised of 8 year-classes: fish from the 2006, 2008 to 2014 year-classes, with fish primarily from the year classes of 2013 and 2011 with 30% and 23.1%, respectively. The ratio of males to females was 1:0.99 in the sample collected (Figure 10.3).



Figure 10.3: Year-class frequency distribution for spotted seatrout collected for ageing in 2014. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

10.3.4 Age-length key

We developed an age-length-key (Table 10.3) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

10.4 REFERENCES

- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analyzing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- Ihde, T., Chittenden, M. 2003. Validation of presumed annual marks on sectioned otoliths of spotted seatrout, Cynoscion nebulosus, in the Chesapeake Bay region. Bulletin of Marine Science, 72(1):77-87, 2003.
- Quinn, T. J. II, and R. B. Deriso. 1999. Quantitative Fish Dynamics. Oxford University Press. New York.
- R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www. R-project.org.

Table 10.1: Number of spotted seatrout collected and aged in each 1-inch length interval in 2014
'Target' represents the sample size for ageing estimated for 2014, and 'Need' represents number of
fish shorted in each length interval compared to the optimum sample size for ageing and number of
fish aged.

Interval	Target	Collected	Aged	Need
6 - 6.99	5	0	0	5
7 - 7.99	5	0	0	5
8 - 8.99	5	0	0	5
9 - 9.99	5	0	0	5
10 - 10.99	6	0	0	6
11 - 11.99	10	6	6	4
12 - 12.99	21	20	20	1
13 - 13.99	16	11	11	5
14 - 14.99	14	16	14	0
15 - 15.99	19	27	20	0
16 - 16.99	29	46	31	0
17 - 17.99	32	41	32	0
18 - 18.99	22	34	22	0
19 - 19.99	24	33	24	0
20 - 20.99	19	26	20	0
21 - 21.99	9	16	10	0
22 - 22.99	10	14	10	0
23 - 23.99	9	15	10	0
24 - 24.99	8	9	8	0
25 - 25.99	6	10	7	0
26 - 26.99	5	7	6	0
27 - 27.99	5	11	6	0
28 - 28.99	5	10	6	0
29 - 29.99	5	6	6	0
30 - 30.99	5	2	2	3
31 - 31.99	5	1	1	4
32 - 32.99	5	0	0	5
33 - 33.99	5	1	1	4
34 - 34.99	5	0	0	5
Totals	319	362	273	57

				Age					
Interval	0	1	2	3	4	5	6	8	Totals
11 - 11.99	6	0	0	0	0	0	0	0	6
12 - 12.99	20	0	0	0	0	0	0	0	20
13 - 13.99	6	5	0	0	0	0	0	0	11
14 - 14.99	0	12	1	1	0	0	0	0	14
15 - 15.99	0	16	4	0	0	0	0	0	20
16 - 16.99	0	16	15	0	0	0	0	0	31
17 - 17.99	0	15	14	3	0	0	0	0	32
18 - 18.99	0	8	9	4	0	1	0	0	22
19 - 19.99	0	9	4	10	1	0	0	0	24
20 - 20.99	0	1	4	12	3	0	0	0	20
21 - 21.99	0	0	0	6	4	0	0	0	10
22 - 22.99	0	0	0	9	1	0	0	0	10
23 - 23.99	0	0	1	6	3	0	0	0	10
24 - 24.99	0	0	0	6	1	0	1	0	8
25 - 25.99	0	0	0	4	2	0	1	0	7
26 - 26.99	0	0	0	1	4	0	1	0	6
27 - 27.99	0	0	0	1	5	0	0	0	6
28 - 28.99	0	0	0	0	4	1	1	0	6
29 - 29.99	0	0	0	0	2	1	1	2	6
30 - 30.99	0	0	0	0	2	0	0	0	2
31 - 31.99	0	0	0	0	0	0	0	1	1
33 - 33.99	0	0	0	0	0	0	1	0	1
Totals	32	82	52	63	32	3	6	3	273

Table 10.2: The number of spotted seatrout assigned to each total length-at-age category for 273 fish sampled for otolith age determination in Virginia during 2014.

(Go back to text) $\overline{\overline{)}}$

				Age				
Interval	0	1	2	3	4	5	6	8
11 - 11.99	1	0	0	0	0	0	0	0
12 - 12.99	1	0	0	0	0	0	0	0
13 - 13.99	0.55	0.45	0	0	0	0	0	0
14 - 14.99	0	0.86	0.07	0.07	0	0	0	0
15 - 15.99	0	0.8	0.2	0	0	0	0	0
16 - 16.99	0	0.52	0.48	0	0	0	0	0
17 - 17.99	0	0.47	0.44	0.09	0	0	0	0
18 - 18.99	0	0.36	0.41	0.18	0	0.05	0	0
19 - 19.99	0	0.38	0.17	0.42	0.04	0	0	0
20 - 20.99	0	0.05	0.2	0.6	0.15	0	0	0
21 - 21.99	0	0	0	0.6	0.4	0	0	0
22 - 22.99	0	0	0	0.9	0.1	0	0	0
23 - 23.99	0	0	0.1	0.6	0.3	0	0	0
24 - 24.99	0	0	0	0.75	0.12	0	0.12	0
25 - 25.99	0	0	0	0.57	0.29	0	0.14	0
26 - 26.99	0	0	0	0.17	0.67	0	0.17	0
27 - 27.99	0	0	0	0.17	0.83	0	0	0
28 - 28.99	0	0	0	0	0.67	0.17	0.17	0
29 - 29.99	0	0	0	0	0.33	0.17	0.17	0.33
30 - 30.99	0	0	0	0	1	0	0	0
31 - 31.99	0	0	0	0	0	0	0	1
33 - 33.99	0	0	0	0	0	0	1	0

Table 10.3: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for spotted seatrout sampled for age determination in Virginia during 2014.

(Go back to $\overline{\overline{\text{text}})}$

Chapter 11

STRIPED BASS Morone saxatilis



11.1 INTRODUCTION

We aged a total of 856 striped bass, Morone saxatilis, using their scales collected by the VMRC's Biological Sampling Program in 2014. Of 856 aged fish, 563 and 293 fish were collected in Chesapeake Bay (bay fish) and Virginia waters of the Atlantic Ocean (ocean fish), respectively. The average bay fish age was 8.3 years with a standard deviation of 4 and a standard error of 0.17. Twenty-one age classes (2 to 21, 23) were represented in the bay fish, comprising fish from the 1991, 1993 to 2012 year classes. The bay fish sample in 2014 was dominated by the year classes of 2009 2008 2004 2010 2007 2005 and 2003 with 13% 12% 12% 9% 9% 9% and 9%, respectively. The average ocean fish age was 11.2 years with a standard deviation of 2.8 and a standard error of 0.16. Sixteen age classes (6 to 21) were represented in the ocean fish, comprising fish from the 1993 to 2008 year classes. The ocean fish sample in 2014 was dominated by the year class of 2004 with 24%. We also aged a total of 235 fish using their otoliths in addition to ageing their scales. The otolith ages were compared to the scale ages to examine how close both ages were to one another (see details in Results).

11.2 METHODS

11.2.1 Sample size for ageing

We estimated sample sizes for ageing striped bass collected in both Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014, respectively, using a twostage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 C V^2 + B_a/L} \tag{11.1}$$

where A is the sample size for ageing striped bass in 2014; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; Lwas the total number of striped bass used by VMRC to estimate length distribution of the catches from 2008 to 2012. θ_a , V_a , B_a , and CV were calculated using pooled age-length data of striped bass collected from 2008 to 2012 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (11.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2008 to 2012 catch. A_l is number of fish to be aged for length interval l in 2014.

11.2.2 Handling of collection

Sagittal otoliths (hereafter, referred to as "otoliths") and scales were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted based on date of capture. Their envelope labels were verified against VMRC's collection data, and each fish assigned a unique Age and Growth Laboratory identification number. All otoliths and scales were stored dry within their original labeled coin envelopes; otoliths were contained inside protective Axygen 2.0 ml microtubes.

11.2.3 Preparation

Scales

Striped bass scales were prepared for age and growth analysis by making acetate impressions of the scale microstructure. Due to extreme variation in the size and shape of scales from individual fish, we selected only those scales that had even margins and which were of uniform size. We selected a range of four to six preferred scales (based on overall scale size) from each fish, making sure that only non-regenerated scales were used. Scale impressions were made on extruded clear acetate sheets (25 mm x 75 mm) with a Carver Laboratory Heated Press (model "C"). The scales were pressed with the following settings:

Pressure: 15000 psi Temperature: 77 °C (170 °F) Time: 5 to 10 min

Striped bass scales that were the size of a quarter (coin) or larger, were pressed individually for up to twenty minutes. After pressing, the impressions were viewed with a Bell and Howell microfiche reader and checked again for regeneration and incomplete margins. Impressions that were too light, or when all scales were regenerated a new impression was made using different scales from the same fish.

Click here to obtain the protocol at the CQFE website on how to prepare scale impression for ageing striped bass.

Otoliths

We used our thin-section and bake technique to process spadefish sagittal otoliths (hereafter, referred to as "otoliths") for age determination. Otolith preparation began by randomly selecting either the right or left otolith. Each whole otolith was placed in a ceramic "Coors" spot plate well and

baked in a Thermolyne 1400 furnace at 400 °C. Baking time was dependent on the otolith's size and gauged by color, with a light caramel color desired. Once a suitable color was achieved the baked otolith was embedded in epoxy resin with its distal surface orientated downwards and allowed to harden overnight. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMetTM low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The otolith was positioned so the blades straddled each side of the otolith focus. It was crucial that this cut be perpendicular to the long axis of the otolith. Failure to do so resulted in broadening and distorted winter growth zones. A proper cut resulted in annuli that were clearly defined and delineated. Once cut, thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing striped bass.

11.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of reading the information contained in its otolith, the date of its capture, and the species-specific period when it deposits its annulus. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called annuli. Technically, an otolith annulus is the combination of both the opaque and the translucent bands. In practice, only the opaque bands are counted as annuli. The number of these visible dark bands replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the otolith section is examined for translucent growth. If no translucent growth is visible beyond the last dark annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last dark annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the dark band of the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of dark bands in the otolith. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday", but before the dark band deposition period, is interpreted as being toward the next age class.

For example, striped bass otolith deposition occurs between April and June (Secor et al. 1995). A striped bass captured between January 1 and June 30, before the end of the species' annulus formation period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of June 30, the period of annulus formation, which would be noted as 4+4.

Striped bass scales are also considered to have a deposition between April and June (Secor et al. 1995), and age class assignment using these hard-parts is conducted in the same way as otoliths.

All striped bass samples (scale pressings and sectioned otoliths) were aged by two different readers in chronological order based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish again without any knowledge of previously estimated ages or lengths, then assigned a final age to the fish. When the age readers were unable to agree on a final age, the fish was excluded from further analysis.

Scales

We determined fish age by viewing acetate impressions of scales (Figure 11.1) with a standard Bell and Howell R-735 microfiche reader equipped with 20 and 29 mm lenses. Annuli on striped bass scales are identified based on two scale microstructure features, "crossing over" and circuli disrup-Primarily, "crossing over" in the tion. lateral margins near the posterior/anterior interface of the scale is used to determine the origin of the annulus. Here compressed circuli (annulus) "cross-over" the previously deposited circuli of the previous year's growth. Typically annuli of the first three years can be observed transversing this interface as dark bands. These bands



Figure 11.1: Scale impression of a 3 year-old striped bass.

remain consistent throughout the posterior field and rejoin the posterior/anterior interface on the opposite side of the focus. Annuli can also be observed in the anterior lateral field of the scale. Here the annuli typically reveal a pattern of discontinuous and suddenly breaking segmented circuli. This event can also be distinguished by the presence of concentric white lines, which are typically associated with the disruption of circuli.

Annuli can also be observed bisecting the perpendicular plain of the radial striations in the anterior field of the scale. Radii emanate out from the focus of the scale towards the outer corner margins of the anterior field. These radial striations consist mainly of segmented concave circuli. The point of intersection between radii and annuli results in a "straightening out" of the concave circuli. This straightening of the circuli should be consistent throughout the entire anterior field of the scale. This event is further amplified by the presence of concave circuli neighboring both directly above and below the annulus. The first year's annulus can be difficult to locate on some scales. It is typically best identified in the lateral field of the anterior portion of the scale. The distance from the focus to the first year's annulus is typically larger with respect to the following annuli. For the annuli two through six, summer growth generally decreases proportionally. For ages greater than six, a crowding effect of the annuli near the outer margins of the scale is observed. This crowding effect creates difficulties in edge interpretation. At this point it is best to focus on the straightening of the circuli at the anterior margins of the scale.

When ageing young striped bass, zero through age two, extreme caution must be taken as not to over age the structure. In young fish there is no point of reference to aid in the determination of the first year; this invariably results in over examination of the scale and such events as hatching or saltwater incursion marks (checks) may be interpreted as the first year.

Otoliths

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 11.2). Each reader aged all of the otolith samples. By conven-



Figure 11.2: Otolith thin-section of a 4 yearold striped bass with the last annulus on the edge of the thin-section

tion an annulus is identified as the narrow opaque zone, or winter growth. Typically the first year's annulus can be determined by first locating the focus of the otolith. The focus is generally located, depending on preparation, in the center of the otolith, and is visually well defined as a dark oblong region. The first year's annulus can be located directly below the focus, along the outer ridge of the sulcal groove on the ventral and dorsal sides of the otolith. This insertion point along the sulcal ridge resembles a check mark (not to be confused with a false annulus). Here the annulus can be followed outwards along the ventral and dorsal surfaces where it encircles the focus. Subsequent annuli also emanate from the sulcal ridge; however, they do not encircle the focus, but rather travel outwards to the distal surface of the otolith. To be considered a true annulus, each annulus must be rooted in the sulcus and travel without interruption to the distal surface of the otolith. The annuli in striped bass have a tendency to split as they advance towards the distal surface. As a result, it is critical that reading path proceed in a direction down the sulcal ridge and outwards to the distal surface.

Click here to obtain the protocol at the CQFE website on how to age striped bass using their otolith thin-sections.

11.2.5 Comparison Tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for following comparisons: 1) between the two readers in the current year; 2) within each reader in the current year; 3) timeseries bias between the current and previous years within each reader; and 4) between scale and otoliths ages. The readings from the entire sample for the current year were used to examine the difference between two readers. A random sub-sample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2000 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

11.3 RESULTS

11.3.1 Sample size

We estimated a sample size of 550 bay striped bass in 2014, ranging in length interval from 17 to 52 inches (Table 11.1). This sample size provided a range in CVfor age composition approximately from the smallest CV of 11% for age 7 to the largest CV of 23% for age 14 of the bay fish. We randomly selected and aged 563 fish from 769 striped bass collected by VMRC in Chesapeake Bay in 2014. We fell short in our over-all collections for this optimal length-class sampling estimate by 59 fish. However, we were short of some fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would possibly be influenced significantly.

We estimated a sample size of 548 ocean striped bass in 2014, ranging in length interval from 26 to 56 inches (Table 11.2). This sample size provided a range in CV for age composition approximately from the smallest CV of 10% for age 7, 8, and 9 to the largest CV of 22% for age 6 of the ocean fish. We aged all 293 striped bass collected by VMRC in Virginia waters of the Atlantic Ocean in 2014. We fell short in our over-all collections for this optimal length-class sampling estimate by 268 fish. However, we were short of many fish from in the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would definitely be influenced significantly. Therefore, precaution should be used when developing ALK using these age data.

11.3.2 Scales

Reader 1 had high self-precision and Read 2 had moderate self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 64% (1 year or less agreement of 86%) and a CV of 3.9%(test of symmetry: $\chi^2 = 16$, df = 13, P =0.2491), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 44% (1) year or less agreement of 70%) and a CV of 7.5% (test of symmetry: $\chi^2 = 19.67, df =$ 15, P = 0.1851). There was an evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 45% (1) year or less agreement of 83%) and a CVof 6.1% (test of symmetry: $\chi^2 = 239.86$, df = 57, P < 0.0001) (Figure 11.3).



Figure 11.3: Between-reader comparison of scale age estimates for striped bass collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

There was no time-series bias for both readers. Reader 1 had an agreement of 57% (1 year or less agreement of 92%) with ages of fish aged in 2000 with a CV of 5.3% (test of symmetry: $\chi^2 = 15.67$, df = 14, P =0.3342). Reader 2 had an agreement of 47% (1 year or less agreement of 87%) with a CV of 7.2% (test of symmetry: $\chi^2 = 17.53$, df = 15, P = 0.288).

Of the 563 bay striped bass aged with

scales, 21 age classes (2 to 21, 23) were represented (Table 11.3). The average age for the sample was 8.3 years. The standard deviation and standard error were 4 and 0.17, respectively. Year-class data (Figure 11.4) indicates that recruitment into the fishery in Chesapeake Bay begins at age 2, which corresponds to the 2012 year-class for striped bass caught in 2014. Striped bass in the sample in 2014 was dominated by the year classes of 2009 2008 2004 2010 2007 2005 and 2003 with 13% 12% 12% 9% 9% 9% and 9%, respectively. The sex ratio of male to female was 1:1.59 for the bay fish.



Figure 11.4: Year-class frequency distribution for striped bass collected in Chesapeake Bay, Virginia for ageing in 2014. Distribution is broken down by sex and estimated using scale ages. 'Unknown' represents the fish gonads that were not available for examination or were not examined for sex during sampling.

Of the 293 ocean striped bass aged with scales, 16 age classes (6 to 21) were represented (Table 11.4). The average age for the sample was 11.2 years. The standard deviation and standard error were 2.8 and 0.16, respectively. Year-class data (Figure 11.5) indicates that recruitment into the fishery in Virginia waters of Atlantic ocean begins at age 6, which corresponds to the 2008 year-class for striped bass caught in 2014. Striped bass in the sample in 2014 was dominated by the year class of 2004 with 24%. The sex ratio of male to female



was 1:3.33 for the ocean fish.

Figure 11.5: Year-class frequency distribution for striped bass collected in Virginia waters of the Atlantic Ocean for ageing in 2014. Distribution is broken down by sex and estimated using scale ages. 'Unknown' represents the fish gonads that were not available for examination or were not examined for sex during sampling.

11.3.3 Otoliths

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 83% and a CV of 0.8% (test of symmetry: χ^2 = 5.33, df = 5, P = 0.3766), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 88% and a CV of 0.6% (test of symmetry: $\chi^2 = 6$, df = 5, P = 0.3062). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 87% (1 year or less agreement of 99%) and a CV of 0.6% (test of symmetry: $\chi^2 = 18.67, df = 16, P =$ 0.2863) (Figure 11.6).

There was no time-series bias for both readers. Reader 1 had an agreement of 78% with ages of fish aged in 2003 with a CV of 2% (test of symmetry: $\chi^2 = 13$, df = 10, P = 0.2237). Reader 2 had an agreement of 67% with a CV of 3.4% (test of symmetry:



Figure 11.6: Between-reader comparison of otolith age estimates for striped bass collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

 $\chi^2 = 20, df = 12, P = 0.0671).$

Of the 235 striped bass aged with otoliths, 23 age classes (2 to 22, 25, 31) were represented (Table 11.5). The average age for the sample was 10.4 years. The standard deviation and standard error were 5.9 and 0.38, respectively.

11.3.4 Comparison of scale and otolith ages

We aged 235 striped bass using scales and otoliths. There was an evidence of systematic disagreement between otolith and scale ages (test of symmetry: $\chi^2 = 92.63$, df = 54, P = 0.0008) with an average CV of 6.8%. There was an agreement of 45% between scale and otoliths ages whereas scales were assigned a lower and higher age than otoliths for 41% and 14% of the fish, respectively (Figure 11.7). There was also an evidence of bias between otolith and scale ages using an age bias plot (Figure 11.8), with scale generally assigned higher ages for younger fish and lower ages for older fish than otolith age estimates.



Figure 11.7: Comparison of scale and otolith age estimates for striped bass collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.



Figure 11.8: Age-bias plot for striped bass scale and otolith age estimates in 2014.

11.3.5 Age-Length-Key (ALK)

We developed an age-length-key for both bay (Table 11.6) and ocean fish (Table 11.7) using scale ages, separately. The ALK can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using scale ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

11.4 RECOMMENDATIONS

We recommend that VMRC and ASMFC use otoliths for ageing striped bass. Although preparation time is greater for otoliths compared to scales, nonetheless as the mean age of striped bass increases in the recovering fishery, otoliths should provide more reliable estimates of age (Liao et al. 2013; Secor et al. 1995). We will continue to compare the age estimates between otoliths and scales.

11.5 REFERENCES

- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age eterminations. Trans. Am. Fish. Soc. 124:131-138.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analyzing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- Liao, H., A. F. Sharov, C. M. Jones, and G. A. Nelson. 2013. Quantifying the Effects of Aging Bias in Atlantic Striped Bass Stock Assessment, Transactions of the American Fisheries Society, 142:1, 193-207.
- Secor, D. H., T. M. Trice, and H. T. Hornick. 1995. Validation of otolith-based ageing and a comparison of otolith and scale-based ageing in mark-recaptured Chesapeake Bay summer flounder, Morone saxatilis. Fishery Bulletin 93: 186-190.
- Quinn, T. J. II, and R. B. Deriso. 1999. Quantitative Fish Dynamics. Oxford University Press. New York.
- R Core Team (2012). R: A language and environment for statistical

computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www. R-project.org.
Table 11.1: Number of bay striped bass collected and aged in each 1-inch length interval in 201	4.
'Target' represents the sample size for ageing estimated for 2014, and 'Need' represents number	of
fish shorted in each length interval compared to the optimum sample size for ageing and number	of
fish aged.	

Interval	Target	Collected	Aged	Need
17 - 17.99	5	2	2	3
18 - 18.99	9	16	10	0
19 - 19.99	21	32	23	0
20 - 20.99	20	42	26	0
21 - 21.99	26	49	32	0
22 - 22.99	26	48	33	0
23 - 23.99	28	51	35	0
24 - 24.99	30	54	32	0
25 - 25.99	26	53	27	0
26 - 26.99	25	42	27	0
27 - 27.99	20	31	23	0
28 - 28.99	18	28	19	0
29 - 29.99	14	26	18	0
30 - 30.99	13	19	16	0
31 - 31.99	15	21	18	0
32 - 32.99	18	28	18	0
33 - 33.99	19	21	20	0
34 - 34.99	23	23	23	0
35 - 35.99	26	18	18	8
36 - 36.99	34	30	30	4
37 - 37.99	31	30	30	1
38 - 38.99	16	11	11	5
39 - 39.99	11	6	6	5
40 - 40.99	11	3	3	8
41 - 41.99	7	7	7	0
42 - 42.99	7	13	9	0
43 - 43.99	6	12	8	0
44 - 44.99	5	21	13	0
45 - 45.99	5	17	11	0
46 - 46.99	5	10	10	0
47 - 47.99	5	3	3	2
48 - 48.99	5	1	1	4
49 - 49.99	5	0	0	5
50 - 50.99	5	0	0	5
51 - 51.99	5	1	1	4
52 - 52.99	5	0	0	5
Totals	550	769	563	59

Table 11.2: Number of ocean striped bass collected and aged in each 1-inch length interval in 2014. 'Target' represents the sample size for ageing estimated for 2014, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
26 - 26.99	5	0	0	5
27 - 27.99	5	0	0	5
28 - 28.99	20	2	2	18
29 - 29.99	37	5	5	32
30 - 30.99	38	13	13	25
31 - 31.99	38	7	7	31
32 - 32.99	35	15	15	20
33 - 33.99	37	29	29	8
34 - 34.99	40	29	29	11
35 - 35.99	50	33	33	17
36 - 36.99	48	32	32	16
37 - 37.99	54	33	33	21
38 - 38.99	33	22	22	11
39 - 39.99	22	14	14	8
40 - 40.99	16	10	10	6
41 - 41.99	7	14	14	0
42 - 42.99	8	5	5	3
43 - 43.99	5	8	8	0
44 - 44.99	5	8	8	0
45 - 45.99	5	5	5	0
46 - 46.99	5	2	2	3
47 - 47.99	5	3	3	2
48 - 48.99	5	2	2	3
49 - 49.99	5	1	1	4
50 - 50.99	5	1	1	4
51 - 51.99	5	0	0	5
52 - 52.99	5	0	0	5
56 - 56.99	5	0	0	5
Totals	548	293	293	268

										Age											
	3	4	5	9	7	∞	6	10	11	12	13	14	15	16	17	18	19	20	21	23	Totals
	-		0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2
	10	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	10
	12	7	2	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	23
	2	12	9	μ	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	26
	4	12	12	2	Η	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	32
	ŝ	IJ	17	9	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	33
	Η	IJ	12	2	9	7	2	0	0	0	0	0	0	0	0	0	0	0	0	0	35
	Η	4	10	∞	9	e C	0	0	0	0	0	0	0	0	0	0	0	0	0	0	32
	0	Η	9	9	7	Ŋ	Η	Η	0	0	0	0	0	0	0	0	0	0	0	0	27
_	0	0	4	13	x	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	27
_	0	0	ŋ	4	x	က	2	Ξ	0	0	0	0	0	0	0	0	0	0	0	0	23
	0	0	Η	9	2	Ŋ	2	2	0	μ	0	0	0	0	0	0	0	0	0	0	19
_	0	Η	0	e S	μ	Ŋ	Η	က	4	0	0	0	0	0	0	0	0	0	0	0	18
	0	0	0	3	2	2	ŋ	2	0	2	0	0	0	0	0	0	0	0	0	0	16
	0	0	Η	2	2	2	0	4	°	3	μ	0	0	0	0	0	0	0	0	0	18
_	0	0	0	2	2	Η	4	4	3	0	μ	0	Η	0	0	0	0	0	0	0	18
_	0	0	0	0	0	Η	4	9	9	1	μ	-	0	0	0	0	0	0	0	0	20
_	0	0	0	0	Η	μ	10	2	0	μ	0	0	0	0	μ	0	0	0	0	0	23
_	0	0	0	0	0	0	9	ю	4	0	μ	0	0	0	0	0	0	0	0	0	18
_	0	0	0	0	0	Η	5	13	11	0	0	0	0	0	0	0	0	0	0	0	30
_	0	0	0	0	0	0	10	14	9	0	0	0	0	0	0	0	0	0	0	0	30
_	0	0	0	0	0	0	1	က	9	0	1	0	0	0	0	0	0	0	0	0	11
	0	0	0	0	0	0	0	4	Ļ	0	0	-	0	0	0	0	0	0	0	0	9
_	0	0	0	0	0	0	0	0	0	0	Ξ	0	0	0	Η	0	0	Η	0	0	c,
	0	0	0	0	0	0	0	0	μ	0	μ	7	μ	0	0	0	0	0	0	0	7
	0	0	0	0	0	0	0	0	μ	0	0	0	0	0	0	0	0	0	0	0	6
_	0	0	0	0	0	0	0	0	0	0	μ	μ	0	က	2	0	-	0	0	0	∞
_	0	0	0	0	0	0	0	0	0	0	0	0	0	Η	4	Η	μ	0	0	0	13
	0	0	0	0	0	0	0	0	0	0	0		0	-	Ξ	4		Ξ	2	0	11
_	0	0	0	0	0	0	0	0	0	0	0	-	Η	0	e S	0	ŝ	Η	Η	0	10
_	0	0	0	0	0	0	0	0	0	0	0	μ	0	μ	0	0	μ	0	0	0	33 C
	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	Η	Π
	0	0	0	0	0	0	0	0	0	0	0	0	0	0	μ	0	0	0	0	0	1
	39	48	76	65	48	35	53	69	48	x	12	12	2	10	13	ŋ	7	က	3 S	Ч	563

Table 11.3: The number of striped bass assigned to each total length-at-age category for 563 fish sampled for scale age determination in Chesapeake Bay, Virginia during 2014.

								Age									
Interval	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	Totals
28 - 28.99	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	2
29 - 29.99	2	0	2	0	1	0	0	0	0	0	0	0	0	0	0	0	5
30 - 30.99	2	9	1	1	0	0	0	0	0	0	0	0	0	0	0	0	13
31 - 31.99	0	1	1	2	1	1	0	1	0	0	0	0	0	0	0	0	7
32 - 32.99	0	4	1	5	4	0	1	0	0	0	0	0	0	0	0	0	15
33 - 33.99	1	2	4	8	11	3	0	0	0	0	0	0	0	0	0	0	29
34 - 34.99	0	1	0	7	13	5	2	1	0	0	0	0	0	0	0	0	29
35 - 35.99	0	0	0	10	11	6	5	1	0	0	0	0	0	0	0	0	33
36 - 36.99	0	0	0	6	12	7	3	2	2	0	0	0	0	0	0	0	32
37 - 37.99	0	0	0	1	13	10	4	3	1	0	0	1	0	0	0	0	33
38 - 38.99	0	0	0	2	1	7	6	2	0	3	0	1	0	0	0	0	22
39 - 39.99	0	0	0	0	1	3	5	4	0	1	0	0	0	0	0	0	14
40 - 40.99	0	0	0	0	0	3	1	2	1	3	0	0	0	0	0	0	10
41 - 41.99	0	0	0	0	0	0	4	4	2	0	1	3	0	0	0	0	14
42 - 42.99	0	0	0	0	0	0	0	3	0	1	1	0	0	0	0	0	5
43 - 43.99	0	0	0	0	0	0	0	1	1	2	3	0	1	0	0	0	8
44 - 44.99	0	0	0	0	0	0	0	0	2	2	2	0	1	1	0	0	8
45 - 45.99	0	0	0	0	0	0	0	1	0	0	1	2	1	0	0	0	5
46 - 46.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	2
47 - 47.99	0	0	0	0	0	0	0	1	0	0	0	0	1	1	0	0	3
48 - 48.99	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	2
49 - 49.99	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1
50 - 50.99	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1
Totals	5	18	9	42	69	45	31	26	9	12	8	9	5	3	1	1	293

Table 11.4: The number of striped bass assigned to each total length-at-age category for 293 fish sampled for scale age determination in Virginia waters of Atlantic ocean during 2014.

Totals	9	6	12	10	9	11	11	9	7	x	7	6	9	12	x	5	8	9	2	5	2	с С	Ω.	4	7	x	14	10	8	3	2	1	٣
31	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0		0	0	0	0	μ	0	(
25	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	Η		0	0	1
22	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2	0	0	0	(
21	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	-	0	4	3	Η	0	0	Ļ	,
20	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	5	0	-		-	0	0	
19	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	Ļ	0			2	0	0	0	,
8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0		Ļ		0	0	ŝ	4	4	0	1	1	0	,
2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	Ļ	0	0	0	0	0	0	0	0	0	0		0	0	Ļ	0	0	0	
6 1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	H.	0	0	0	0	0	
5 1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	Ļ	0	0	1	1	0	0	Ч	0	0	0	0	0	
4 1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	Ļ	0	0	1	0	0	0	1	2		Г	0	0	0	0	0	
3 e	0	0	0	0	0	0	0	0	0	0	0	0	0	5	2	1	1	0	1	1	2	Ļ	1	5	1	0	0	1	0	0	0	0	
12	0	0	0	0	0	0	0	0	0	0	0	0	0	0	Η	0	0	0	0	0	0	1	0	0	0	0	1	0	0	0	0	0	
11	0	0	0	0	0	Η	0	Η	0	0	Η	ŝ	0	Η	0	2	Η	c,	က		2	0	Η	0	1	0	-	0	0	0	0	0	
10	0	0	0	0	0	0	0	0	0	0	0	0	μ	2	Η	0	2	2	က	Ξ	Ξ	0	0	0	0	0	0	0	0	0	0	0	
6	0	0	0	0	0	0	0	4	0	0	0	7	μ	e S	Η	Η	μ	μ	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
∞	0	0	0	0	0	0	0	0	0	0	Η	Η	-	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
2	0	0	0	0	0	0	Ŋ	0	μ	4	0	0	0	n	0	μ	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
9	0	0	Η	2	Η	Η	2	0	4	2	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
ro	0	2	7	2	IJ	4	2	1	0	0	1	0	Ļ	Η	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
4	0	4	ŋ	4	က	μ	Η	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
e S	S	က	4	0	0	0	Η	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
7		0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
terval	18.99	19.99	20.99	21.99	- 22.99	- 23.99	- 24.99	- 25.99	- 26.99	- 27.99	- 28.99	- 29.99	- 30.99	- 31.99	- 32.99	- 33.99	- 34.99	- 35.99	- 36.99	- 37.99	- 38.99	- 39.99	- 40.99	- 41.99	- 42.99	- 43.99	- 44.99	- 45.99	- 46.99	- 47.99	- 48.99	- 50.99	

Table 11.5: The number of striped bass assigned to each total length-at-age category for 235 fish sampled for otolith age determination in Chesapeake Bay and Virginia waters of Atlantic Ocean during 2014.

$\begin{tabular}{ llllllllllllllllllllllllllllllllllll$		$\begin{array}{c} & & & & & & & \\ & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & & & \\ & & & & & &$	$\begin{array}{c} & 4\\ & 0.5\\ & 0.3\\ & 0.46\\ & 0.12\\ & 0.14\\ & 0.04\\ & 0.04\\ & 0.04\\ & 0.04\\ & 0.04\\ & 0.04\\ & 0.04\\ & 0\\ & 0\\ & 0\\ & 0\\ & 0\\ & 0\\ & 0\\ &$	$\begin{array}{c} & & & & & & \\ & & & & & & \\ & & & & & $	$\begin{array}{c} 6\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\ 0\\$	$\begin{array}{c} & & & & & & & & \\ & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & \\ & & & & & & & & & \\ & & & & & & & & & \\ & & & & & & & & & \\ & & & & & & & & & \\ & & & & & & & & & \\ & & & & & & & & & \\ & & & & & & & & & \\ & & & & & & & & & \\ & & & & & & & & & \\ & & & & & & & & & & \\ & & & & & & & & & \\ & & & & & & & & & \\ & & & & & & & & \\$	$\begin{array}{c} 0.0\\ 0.0\\ 0.0\\ 0.0\\ 0.0\\ 0.0\\ 0.0\\ 0.0$	$\begin{array}{c} 8 \\ 8 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\$			$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{c ccccc} Age \\ \hline 10 & 11 \\ \hline 0 & 0 \\ 0 & 0 \\ 0 & 0 \\ 0 & 0 \\ 0 & 0 \\ 0 & 0 \\ 0 & 0 \\ 0 & 0 \\ 0 & 0 \\ 0 & 0 \\ 0 & 0 \\ 0 & 0 \\ 11 \\ 0 & 0 \\ 0 \\ 11 \\ 0 \\ 0 \\ 0 \\ 11 \\ 0 \\ 0 \\ 0$	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$	Age10111213001100.0501200.170.06120.170.170.061200.140.09130.2200.061470.20000.1400.1200 <td< th=""><th>Age 10 11 12 13 14 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 11 0 0.05 0 <</th><th>Age 11 12 13 14 15 0 1 12 13 14 15 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 11 0 0.05 0.05 0 0 0 0 12 0.17 0.16 0 0 0 0 0 0 0 0<</th><th>Age10111213141516001100.050000000000001100.050000120.170.060000130.3700.06000140.2200000150.17000001670.17000001670.1400.120.12001670.14000001670.140000016700000016700000016700<</th><th>Age 11 12 13 14 15 16 17 0 <td< th=""><th>Age 11 12 13 14 15 16 17 18 0</th><th>Age 11 12 13 14 15 16 17 18 19 0 <t< th=""><th>Age 11 12 13 14 15 16 17 18 19 20 0</th><th>Age 1</th></t<></th></td<></th></td<>	Age 10 11 12 13 14 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 11 0 0.05 0 <	Age 11 12 13 14 15 0 1 12 13 14 15 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 11 0 0.05 0.05 0 0 0 0 12 0.17 0.16 0 0 0 0 0 0 0 0<	Age10111213141516001100.050000000000001100.050000120.170.060000130.3700.06000140.2200000150.17000001670.17000001670.1400.120.12001670.14000001670.140000016700000016700000016700<	Age 11 12 13 14 15 16 17 0 <td< th=""><th>Age 11 12 13 14 15 16 17 18 0</th><th>Age 11 12 13 14 15 16 17 18 19 0 <t< th=""><th>Age 11 12 13 14 15 16 17 18 19 20 0</th><th>Age 1</th></t<></th></td<>	Age 11 12 13 14 15 16 17 18 0 0	Age 11 12 13 14 15 16 17 18 19 0 0 <t< th=""><th>Age 11 12 13 14 15 16 17 18 19 20 0</th><th>Age 1</th></t<>	Age 11 12 13 14 15 16 17 18 19 20 0	Age 1
1	0	0.5 5	0.5	0 0	00	0 -		00	0	0		0	0 0	0 0 0 0 0 0 0 0	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
3 - 18.99	0	1	0	0	0	0	-	0	0	0	0		0	0 0								
20 - 20.99	0 0	0.27	0.46	0.23	0.04	0 0		0 0	0	0 0	0		0	0 0	0 0 0	0 0 0 0	0 0 0 0 0 0					
21 - 21.99	0.03	0.12	0.38	0.38	0.06	0.03	_	0	0	0	0		0	0 0	0 0 0	0 0 0 0	0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0 0	0 0 0 0 0 0 0 0	0 0 0 0 0 0 0 0 0	0 0 0 0 0 0 0 0 0 0
22 - 22.99	0	0.09	0.15	0.52	0.18	0.06			0	0	0		0	0 0	0 0 0							
23 - 23.99 24 - 24.99		0.03	0.14	0.34	0.25	0.17	0.0	9 0.0	⊃ ō.													
25 - 25.99	0	0	0.04	0.22	0.22	0.26	0.1	9 0.0	40.	04		0	0	0 0	0 0 0							
26 - 26.99	0	0	0	0.15	0.48	0.3	0.0	7	0	0		0	0 0	0 0 0	0 0 0 0 0	0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0 0	0 0 0 0 0 0 0 0	0 0 0 0 0 0 0 0 0	0 0 0 0 0 0 0 0 0 0
27 - 27.99	0	0	0	0.22	0.17	0.35	0.1	3 0.0	· 99	. 4		0	0 0									
28 - 28.99 29 - 29.99	0 0		0.06	0.05 0	0.32	$0.11 \\ 0.06$	0.2		6 1 0.0	11 17 (22 0	$\begin{array}{c} 0 & 0.05 \\ 22 & 0 \end{array}$	0 0.05 0 22 0 0	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	22 0 0 0 0 0 0					
30 - 30.99	0	0	0	0	0.19	0.12	0.1	2 0.3	0.	12		0	0 0.12	0 0.12 0	$\begin{array}{cccccccccccccccccccccccccccccccccccc$	$\begin{array}{cccccccccccccccccccccccccccccccccccc$						
31 - 31.99	0	0	0	0.06	0.11	0.11	0.1	1	0 0.	22 ($\frac{1}{2}$	-1	7 0.17	7 0.17 0.06	0.17 0.06 0	7 0.17 0.06 0 0	7 0.17 0.06 0 0 0	7 0.17 0.06 0 0 0 0	7 0.17 0.06 0 0 0 0 0	7 0.17 0.06 0 0 0 0 0 0 0	7 0.17 0.06 0 0 0 0 0 0 0 0	7 0.17 0.06 0 0 0 0 0 0 0 0 0
32 - 32.99	0	0	0	0	0.11	0.11	0.0	6 0.2	0.1	22 (-1	0	7 0 0.06	7 0 0.06 0	7 0 0.06 0 0.06	7 0 0.06 0 0.06 0	7 0 0.06 0 0.06 0 0	7 0 0.06 0 0.06 0 0 0	7 0 0.06 0 0.06 0 0 0 0	7 0 0.06 0 0.06 0 0 0 0 0	7 0 0.06 0 0.06 0 0 0 0 0 0 0
33 - 33.99		0 0		0 0	0 0	0	0.0	- 5 - 0 - 0	- 12 - C		0	⊷ చ	0.05	0.05 0.05								
35 - 35.99	0	0	0	0	0	0	0.1	1 0.3		28	$\tilde{\mathbf{x}}$	12	22 0	22 0 0.06	22 0 0.06 0	22 0 0.06 0 0						
36 - 36.99	0	0	0	0	0	0	0.0	3 0.1	.7	43 (37	37 0	37 0 0	37 0 0 0	37 0 0 0 0 0	37 0 0 0 0 0	37 0 0 0 0 0 0	37 0 0 0 0 0 0 0 0	37 0 0 0 0 0 0 0 0 0	37 0 0 0 0 0 0 0 0 0 0	37 0 0 0 0 0 0 0 0 0 0 0
37 - 37.99	0	0	0	0	0	0	-	0 0.3	33 0.	47	0	.2	.2 0	.2 0 0	.2 0 0 0	.2 0 0 0 0	.2 0 0 0 0 0	.2 0 0 0 0 0 0	.2 0 0 0 0 0 0 0	.2 0 0 0 0 0 0 0 0 0 0 0 0 0	.2 0 0 0 0 0 0 0 0 0	.2 0 0 0 0 0 0 0 0 0 0
38 - 38.99	0	0	0	0	0	0	-	0 0.0	9 0.	27 (č.	ŭ	0	0 0.09	0 0.09 0	55 0 0.09 0 0	55 0 0.09 0 0 0	5 0 0.09 0 0 0 0	5 0 0.09 0 0 0 0 0	5 0 0.09 0 0 0 0 0 0	5 0 0.09 0 0 0 0 0 0 0 0	5 0 0.09 0 0 0 0 0 0 0 0 0
39 - 39.99	0	0	0	0	0	0	-	0	0 0.	67 (4	7 0	0 0	7 0 0 0.17	7 0 0 0.17 0	7 0 0 0.17 0 0	7 0 0 0.17 0 0 0	7 0 0 0.17 0 0 0 0	7 0 0 0.17 0 0 0 0 0	7 0 0 0.17 0 0 0 0 0 0	7 0 0 0.17 0 0 0 0 0 0 0 0
40 - 40.99	0	0	0	0	0	0	-	0	0	0		0	0 0	0 0 0.33	0 0 0.33 0	0 0 0.33 0 0	$0 \qquad 0 0.33 \qquad 0 \qquad 0 0$	$0 \qquad 0 \qquad 0.33 \qquad 0 \qquad 0 \qquad 0 \qquad 0.33$	$0 \qquad 0 \qquad 0.33 \qquad 0 \qquad 0 \qquad 0 \qquad 0.33 \qquad 0$	0 0 0.33 0 0 0 0.33 0 0	$0 \qquad 0 \qquad 0.33 \qquad 0 \qquad 0 \qquad 0 \qquad 0.33 \qquad 0 \qquad 0 \qquad 0.33$	0 0 0.33 0 0 0 0.33 0 0 0 0.33 0 0 0.33 0 0 0.33 0 0 0.33 0 0 0.33 0 0 0.33 0 0 0.33 0 0 0 0.33 0 0 0 0.33 0 0 0 0.33 0 0 0 0.33 0 0 0 0.33 0 0 0 0.33 0 0 0 0.33 0 0 0 0.33 0 0 0 0.33 0 0 0 0 0 0.33 0 0 0 0 0 0 0 0 0
41 - 41.99	0	0	0	0	0	0	-	0	0	0	$\tilde{\cdot}$	14	14 0	14 0 0.14	14 0 0.14 0.29	14 0 0.14 0.29 0.14	14 0 0.14 0.29 0.14 0.29	14 0 0.14 0.29 0.14 0.29 0	14 0 0.14 0.29 0.14 0.29 0 0	14 0 0.14 0.29 0.14 0.29 0 0 0 0	14 0 0.14 0.29 0.14 0.29 0 0 0 0 0	14 0 0.14 0.29 0.14 0.29 0 0 0 0 0 0 0 0 0
42 - 42.99	0	0	0	0	0	0	-	0	0	0	~	11	11 0	11 0 0.22	11 0 0.22 0.22	$11 \qquad 0 \qquad 0.22 \qquad 0.22 \qquad 0.22$	$11 \qquad 0 \qquad 0.22 \qquad 0.22 \qquad 0.22 \qquad 0.22$	$11 \qquad 0 \qquad 0.22 \qquad 0.22 \qquad 0.22 \qquad 0.22 \qquad 0$	11 0 0.22 0.22 0.22 0.22 0 0	11 0 0.22 0.22 0.22 0.22 0 0 0	11 0 0.22 0.22 0.22 0.22 0 0 0 0 0	11 0 0.22 0.22 0.22 0.22 0 0 0 0 0 0 0 0 0
43 - 43.99	0	0	0	0	0	0	_	0	0	0	·	0	0 0	$\begin{array}{cccccccccccccccccccccccccccccccccccc$								
44 - 44.99	0 0	0 0	0 0	0	0 0	0		0	0	0 0	\sim	.15	.15 0	15 0 0								
45 - 45.99 46 - 46.99			0 0	0 0	0 0			0 0	0 0													0 0 0 0.1 0.1 0 0.3 0 0.3 0.1 0.1
47 - 47.99	0)	0	0	0	0	-	0	0	0		0	0 0	0 0 0	0 0 0 0.33	0 0 0 0.33 0	0 0 0 0.33 0 0.33	0 0 0 0.33 0 0.33 0	0 0 0 0.33 0 0.33 0 0	0 0 0 0.33 0 0.33 0 0.33	0 0 0 0.33 0 0.33 0 0 0.33 0	0 0 0 0.33 0 0.33 0 0 0.33 0 0
48 - 48.99	0	0		>	0	0	_						,	0 0 0	0 0 0 0	0 0 0 0 0	0 0 0 0 0 0	0 0 0 0 0 0 0		0 0 0 0 0 0 0 0	0 0 0 0 0 0 0 0 0	
	,	0 0	0	0	0	0		C	C	0		0	0 0									

								Age								
nterval	9	2	×	6	10	11	12	13	14	15	16	17	18	19	20	21
- 28.99	0	0.5	0	0	0.5	0	0	0	0	0	0	0	0	0	0	0
- 29.99	0.4	0	0.4	0	0.2	0	0	0	0	0	0	0	0	0	0	0
- 30.99	0.15	0.69	0.08	0.08	0	0	0	0	0	0	0	0	0	0	0	0
- 31.99	0	0.14	0.14	0.29	0.14	0.14	0	0.14	0	0	0	0	0	0	0	0
32.99	0	0.27	0.07	0.33	0.27	0	0.07	0	0	0	0	0	0	0	0	0
. 33.99	0.03	0.07	0.14	0.28	0.38	0.1	0	0	0	0	0	0	0	0	0	0
. 34.99	0	0.03	0	0.24	0.45	0.17	0.07	0.03	0	0	0	0	0	0	0	0
. 35.99	0	0	0	0.3	0.33	0.18	0.15	0.03	0	0	0	0	0	0	0	0
- 36.99	0	0	0	0.19	0.38	0.22	0.09	0.06	0.06	0	0	0	0	0	0	0
. 37.99	0	0	0	0.03	0.39	0.3	0.12	0.09	0.03	0	0	0.03	0	0	0	0
- 38.99	0	0	0	0.09	0.05	0.32	0.27	0.09	0	0.14	0	0.05	0	0	0	0
. 39.99	0	0	0	0	0.07	0.21	0.36	0.29	0	0.07	0	0	0	0	0	0
40.99	0	0	0	0	0	0.3	0.1	0.2	0.1	0.3	0	0	0	0	0	0
- 41.99	0	0	0	0	0	0	0.29	0.29	0.14	0	0.07	0.21	0	0	0	0
42.99	0	0	0	0	0	0	0	0.6	0	0.2	0.2	0	0	0	0	0
- 43.99	0	0	0	0	0	0	0	0.12	0.12	0.25	0.38	0	0.12	0	0	0
- 44.99	0	0	0	0	0	0	0	0	0.25	0.25	0.25	0	0.12	0.12	0	0
- 45.99	0	0	0	0	0	0	0	0.2	0	0	0.2	0.4	0.2	0	0	0
- 46.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0.5	0.5	0
- 47.99	0	0	0	0	0	0	0	0.33	0	0	0	0	0.33	0.33	0	0
- 48.99	0	0	0	0	0	0	0	0	0	0	0	0	0.5	0	0	0.5
- 49.99	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
. 50 00		C	C		C	C			0	C		,	0	0	0	C

Table 11.7: Age-Length key, as proportion-at-age in each 1-inch length interval, based on scale ages for striped bass sampled in Virginia waters of the Atlantic Ocean during 2014.

Chapter 12

SUMMER FLOUNDER Paralichthys dentatus



12.1 INTRODUCTION

We aged a total of 819 summer flounder, Paralichthys dentatus, using their scales collected by the VMRC's Biological Sampling Program in 2014. Of 819 aged fish, 371 and 448 fish were collected in Chesapeake Bay (bay fish) and Virginia waters of the Atlantic Ocean (ocean fish), respectively. The average bay fish age was 3.5 years with a standard deviation of 1.6 and a standard error of 0.08. Nine age classes (0 to 8) were represented in the bay fish, comprising fish from the 2006 to 2014 year classes. The bay fish sample in 2014 was dominated by the year classes of 2012 and 2011 with 30% and 29%, respectively. The average ocean fish age was 4.9 years with a standard deviation of 2.1 and a standard error of 0.1. Fourteen age classes (1 to 14) were represented in the ocean fish, comprising fish from the 2000 to 2013 vear classes. The ocean fish sample in 2014 was dominated by the year classes of 2009 and 2010 with 24% and 23%, respectively. We also aged a total of 230 fish using their otoliths in addition to ageing their scales. The otolith ages were compared to the scale ages to examine how close both ages were to one another (see details in Results).

12.2 METHODS

12.2.1 Sample size for ageing

We estimated sample sizes for ageing summer flounder collected in both Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014, respectively, using a twostage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 C V^2 + B_a/L} \tag{12.1}$$

where A is the sample size for ageing summer flounder in 2014; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; Lwas the total number of summer flounder used by VMRC to estimate length distribution of the catches from 2008 to 2012. θ_a, V_a, B_a , and CV were calculated using pooled age-length data of summer flounder collected from 2008 to 2012 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (12.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval *l* from the length distribution of the 2008 to 2012 catch. A_l is number of fish to be aged for length interval lin 2014.

12.2.2 Handling of collection

Sagittal otoliths (hereafter, referred to as "otoliths") and scales were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted based on date of capture, their envelope labels were verified against VMRC's collection data, and each fish assigned a unique Age and Growth Laboratory identification number. All otoliths and scales were stored dry within their original labeled coin envelopes; otoliths were contained inside protective Axygen 2.0 ml microtubes.

12.2.3 Preparation

Scales

Summer flounder scales were prepared for age and growth analysis by making acetate impressions of the scale microstructure. Due to extreme variation in the size and shape of scales from individual fish, we selected only those scales that had even margins and which were of uniform size. We selected a range of four to six preferred scales (based on overall scale size) from each fish, making sure that only nonregenerated scales were used. Scale impressions were made on extruded clear acetate sheets (25 mm x 75 mm) with a Carver Laboratory Heated Press (model "C"). The scales were pressed with the following settings:

Pressure: 15000 psi Temperature: 77 °C (170 °F) Time: 5 to 10 min

summer flounder scales that were the size of a quarter (coin) or larger, were pressed individually for up to twenty minutes. After pressing, the impressions were viewed with a Bell and Howell microfiche reader and checked again for regeneration and incomplete margins. Impressions that were too light, or when all scales were regenerated a new impression was made using different scales from the same fish.

Click here to obtain the protocol at the CQFE website on how to prepare scale impression for ageing summer flounder.

Otoliths

We used our thin-section and bake technique to process spadefish sagittal otoliths (hereafter, referred to as "otoliths") for age determination. Otolith preparation began by randomly selecting either the right or left otolith. Each whole otolith was placed

in a ceramic "Coors" spot plate well and baked in a Thermolyne 1400 furnace at 400 °C. Baking time was dependent on the otolith's size and gauged by color, with a light caramel color desired. Once a suitable color was achieved the baked otolith was embedded in epoxy resin with its distal surface orientated downwards and allowed to harden overnight. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMetTM low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The otolith was positioned so the blades straddled each side of the otolith focus. It was crucial that this cut be perpendicular to the long axis of the otolith. Failure to do so resulted in broadening and distorted winter growth zones. A proper cut resulted in annuli that were clearly defined and delineated. Once cut, thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing summer flounder.

12.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of reading the information contained in its otolith, the date of its capture, and the species-specific period when it deposits its annulus. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called annuli. Technically, an otolith annulus is the combination of both the opaque and the translucent bands. In practice, only the opaque bands are counted as annuli. The number of these visible dark bands replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the otolith section is examined for translucent growth. If no translucent growth is visible beyond the last dark annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last dark annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the dark band of the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of dark bands in the otolith. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday", but before the dark band deposition period, is interpreted as being toward the next age class.

For example, summer flounder otolith deposition occurs between January and April (Bolz et al. 2000). A summer flounder captured between January 1 and April 30, before the end of the species' annulus formation period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of June 30, the period of annulus formation, which would be noted as 4+4.

Summer flounder scales are also considered to have a deposition between January and June (Bolz et al. 1999 and modified by CQFE), and age class assignment using these hard-parts is conducted in the same way as otoliths.

All summer flounder samples (scale pressings and sectioned otoliths) were aged by two different readers in chronological order based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish again without any knowledge of previously estimated ages or lengths, then assigned a final age to the fish. When the age readers were unable to agree on a final age, the fish was excluded from further analysis.

Scales

We determined fish age by viewing acetate impressions of scales (Figure 12.1) with a standard Bell and Howell R-735 microfiche reader equipped with 20 and 29 mm lenses. Annuli on summer flounder scales are identified based on two scale microstructure features, "crossing over" and circuli disruption. Primarily, "crossing over" in the lateral margins near the posterior/anterior interface of the scale is used to determine the origin of the annulus. Here compressed circuli (annulus) "cross-over" the previously deposited circuli of the previous



Figure 12.1: Scale impression of a 1 year-old summer flounder

year's growth. Typically annuli of the first three years can be observed transversing this interface as dark bands. These bands remain consistent throughout the posterior field and rejoin the posterior/anterior interface on the opposite side of the focus. Annuli can also be observed in the anterior lateral field of the scale. Here the annuli typically reveal a pattern of discontinuous and suddenly breaking segmented circuli. This event can also be distinguished by the presence of concentric white lines, which are typically associated with the disruption of circuli.

Annuli can also be observed bisecting the perpendicular plain of the radial striations in the anterior field of the scale. Radii emanate out from the focus of the scale towards the outer corner margins of the anterior field. These radial striations consist mainly of segmented concave circuli. The point of intersection between radii and annuli results in a "straightening out" of the concave circuli. This straightening of the circuli should be consistent throughout the

entire anterior field of the scale. This event is further amplified by the presence of concave circuli neighboring both directly above and below the annulus. The first year's annulus can be difficult to locate on some scales. It is typically best identified in the lateral field of the anterior portion of the scale. The distance from the focus to the first year's annulus is typically larger with respect to the following annuli. For the annuli two through six, summer growth generally decreases proportionally. For ages greater than six, a crowding effect of the annuli near the outer margins of the scale is observed. This crowding effect creates difficulties in edge interpretation. At this point it is best to focus on the straightening of the circuli at the anterior margins of the scale.

When ageing young summer flounder, zero through age two, extreme caution must be taken as not to over age the structure. In young fish there is no point of reference to aid in the determination of the first year; this invariably results in over examination of the scale and such events as hatching or saltwater incursion marks (checks) may be interpreted as the first year.

Otoliths

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 12.2). Each reader aged all of the otolith samples. By conven-



Figure 12.2: Otolith thin-section of a 4 yearold summer flounder with the last annulus on the edge of the thin-section

tion an annulus is identified as the narrow opaque zone, or winter growth. Typically the first year's annulus can be determined by first locating the focus of the otolith. The focus is generally located, depending on preparation, in the center of the otolith, and is visually well defined as a dark oblong region. The first year's annulus can be located directly below the focus, along the outer ridge of the sulcal groove on the ventral and dorsal sides of the otolith. This insertion point along the sulcal ridge resembles a check mark (not to be confused with a false annulus). Here the annulus can be followed outwards along the ventral and dorsal surfaces where it encircles the focus. Subsequent annuli also emanate from the sulcal ridge; however, they do not encircle the focus, but rather travel outwards to the distal surface of the otolith. To be considered a true annulus, each annulus must be rooted in the sulcus and travel without interruption to the distal surface of the otolith. The annuli in summer flounder have a tendency to split as they advance towards the distal surface. As a result, it is critical that reading path proceed in a direction down the sulcal ridge and outwards to the distal surface.

Click here to obtain the protocol at the CQFE website on how to age summer flounder using their otolith thinsections.

12.2.5 Comparison Tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for following comparisons: 1) between the two readers in the current year; 2) within each reader in the current year; 3) timeseries bias between the current and previous years within each reader; and 4) between scale and otoliths ages. The readings from the entire sample for the current year were used to examine the difference between two readers. A random sub-sample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2000 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

12.3 RESULTS

12.3.1 Sample size

We estimated a sample size of 383 bay summer flounder in 2014, ranging in length interval from 8 to 28 inches (Table 12.1). This sample size provided a range in CV for age composition approximately from the smallest CV of 6% for age 2 to the largest CV of 22% for age 6 of the bay fish. We randomly selected and aged 371 fish from 455 summer flounder collected by VMRC in Chesapeake Bay in 2014. We fell short in our over-all collections for this optimal length-class sampling estimate by 42 fish. However, we were short of no fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

We estimated a sample size of 446 ocean summer flounder in 2014, ranging in length interval from 11 to 32 inches (Table 12.2). This sample size provided a range in CV for age composition approximately from the smallest CV of 8% for age 3 to the largest CV of 21% for age 8 of the ocean fish. We randomly selected and aged 448 fish from 622 summer flounder collected by VMRC in Virginia waters of the Atlantic Ocean in 2014. We fell short in our over-all collections for this optimal length-class sampling estimate by 24 fish. However, we were short of no fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

12.3.2 Scales

Both readers had moderate self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 64% (1) year or less agreement of 98%) and a CVof 8.5% (test of symmetry: $\chi^2 = 10, df =$ 8, P = 0.265), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 62% (1 year or less agreement of 96%) and a CV of 8.6% (test of symmetry: $\chi^2 = 8, df$ =7, P=0.3326). There was an evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 59% (1) year or less agreement of 91%) and a CVof 9.2% (test of symmetry: $\chi^2 = 60.62$, df = 33, P = 0.0024) (Figure 12.3).



Figure 12.3: Between-reader comparison of scale age estimates for summer flounder collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

There was no time-series bias for both readers. Reader 1 had an agreement of 82% (1 year or less agreement of 100%) with ages of fish aged in 2000 with a CV of 3.7% (test of symmetry: $\chi^2 = 3.29$, df = 3, P = 0.3496). Reader 2 had an agreement of 74% (1 year or less agreement of 96%) with a CV of 6.2% (test of symmetry: $\chi^2 = 10.33$, df =5, P = 0.0663).

Of the 371 bay summer flounder aged with scales, 9 age classes (0 to 8) were represented (Table 12.3). The average age for the sample was 3.5 years. The standard deviation and standard error were 1.6 and 0.08, respectively. Year-class data (Figure 12.4) indicates that recruitment into the fishery in Chesapeake Bay begins at age 0, which corresponds to the 2014 year-class for summer flounder caught in 2014. Summer flounder in the sample in 2014 was dominated by the year classes of 2012 and 2011 with 30% and 29%, respectively. The sex ratio of male to female was 1:74 for the bay fish.



Figure 12.4: Year-class frequency distribution for summer flounder collected in Chesapeake Bay, Virginia for ageing in 2014. Distribution is broken down by sex and estimated using scale ages. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

Of the 448 ocean summer flounder aged with scales, 14 age classes (1 to 14) were represented (Table 12.4). The average age for the sample was 4.9 years. The standard deviation and standard error were 2.1 and 0.1, respectively. Year-class data (Figure 12.5) indicates that recruitment into the fishery in Virginia waters of Atlantic ocean begins at age 1, which corresponds to the 2013 year-class for summer flounder caught in 2014. Summer flounder in the sample in 2014 was dominated by the year classes of 2009 and 2010 with 24% and 23%, respectively. The sex ratio of male to female was 1:1.32 for the ocean fish.



Figure 12.5: Year-class frequency distribution for summer flounder collected in Virginia waters of the Atlantic Ocean for ageing in 2014. Distribution is broken down by sex and estimated using scale ages. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

12.3.3 Otoliths

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 92% and a CV of 1% (test of symmetry: $\chi^2 = 2$, df = 3, P = 0.5724), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 86% and a CV of 1.9% (test of symmetry: $\chi^2 = 5$, df = 6, P = 0.5438). There was an evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 85% (1 year or less agreement of 97%) and a CV of 2.4% (test of symmetry: $\chi^2 = 24.57, df = 13, P =$ 0.0263) (Figure 12.6).



Figure 12.6: Between-reader comparison of otolith age estimates for summer flounder collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

There was no time-series bias for both readers. Reader 1 had an agreement of 98% with ages of fish aged in 2003 with a CV of 0.3% (test of symmetry: $\chi^2 = 1$, df = 1, P = 0.3173). Reader 2 had an agreement of 92% with a CV of 1.6% (test of symmetry: $\chi^2 = 4$, df = 3, P = 0.2615).

Of the 230 summer flounder aged with otoliths, 14 age classes (0 to 11, 13 to 14) were represented (Table 12.5). The average age for the sample was 4.6 years. The standard deviation and standard error were 2.5 and 0.16, respectively.

12.3.4 Comparison of scale and otolith ages

We aged 230 summer flounder using scales and otoliths. There was no evidence of systematic disagreement between otolith and scale ages (test of symmetry: $\chi^2 = 33.24$, df = 24, P = 0.0991) with an average CV of 10.1%. There was an agreement of 52% between scale and otoliths ages whereas scales were assigned a lower and higher age than otoliths for 33% and 15% of the fish, respectively (Figure 12.7). There was also little evidence of bias between otolith and scale ages using an age bias plot (Figure 12.8), with no trend of either over-ageing younger or under-ageing older fish.



Figure 12.7: Comparison of scale and otolith age estimates for summer flounder collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.



Figure 12.8: Age-bias plot for summer flounder scale and otolith age estimates in 2014.

12.3.5 Age-Length-Key (ALK)

We developed an age-length-key for both bay (Table 12.6) and ocean fish (Table 12.7) using scale ages, separately. The ALK can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using scale ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

12.4 REFERENCES

- Bolz, G., R. Monaghan, K. Lang, R. Gregory, and J. Burnett. 2000. Proceedings of the summer flounder ageing workshop, 1-2 February 1999, Woods Hole, MA. Massachusetts. US Dep. Commerce, NOAA Tech. Memo. NMFS NE 156; 15 p.
- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analyzing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- Quinn, T. J. II, and R. B. Deriso. 1999. Quantitative Fish Dynamics. Oxford University Press. New York.
- R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.

Table 12.1: Number of bay summer flounder collected and aged in each 1-inch length interval in 2014.
'Target' represents the sample size for ageing estimated for 2014, and 'Need' represents number of
fish shorted in each length interval compared to the optimum sample size for ageing and number of
fish aged.

Interval	Target	Collected	Aged	Need
8 - 8.99	5	1	1	4
9 - 9.99	5	0	0	5
10 - 10.99	5	0	0	5
11 - 11.99	5	0	0	5
12 - 12.99	5	1	1	4
13 - 13.99	7	1	1	6
14 - 14.99	67	81	75	0
15 - 15.99	55	75	55	0
16 - 16.99	43	57	44	0
17 - 17.99	40	49	41	0
18 - 18.99	30	46	33	0
19 - 19.99	27	39	31	0
20 - 20.99	20	30	25	0
21 - 21.99	19	30	24	0
22 - 22.99	14	21	16	0
23 - 23.99	10	10	10	0
24 - 24.99	6	7	7	0
25 - 25.99	5	5	5	0
26 - 26.99	5	1	1	4
27 - 27.99	5	1	1	4
28 - 28.99	5	0	0	5
Totals	383	455	371	42

Interval	Target	Collected	Aged	Need
11 - 11.99	5	0	0	5
12 - 12.99	5	0	0	5
13 - 13.99	5	3	3	2
14 - 14.99	35	58	43	0
15 - 15.99	61	94	61	0
16 - 16.99	63	94	64	0
17 - 17.99	49	71	50	0
18 - 18.99	37	38	38	0
19 - 19.99	26	36	26	0
20 - 20.99	24	24	24	0
21 - 21.99	18	28	21	0
22 - 22.99	23	32	23	0
23 - 23.99	22	25	22	0
24 - 24.99	18	28	18	0
25 - 25.99	14	21	15	0
26 - 26.99	9	25	12	0
27 - 27.99	7	16	8	0
28 - 28.99	5	16	7	0
29 - 29.99	5	10	10	0
30 - 30.99	5	1	1	4
31 - 31.99	5	2	2	3
32 - 32.99	5	0	0	5
Totals	446	622	448	24

Table 12.2: Number of ocean summer flounder collected and aged in each 1-inch length interval in 2014. 'Target' represents the sample size for ageing estimated for 2014, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

					Age					
Interval	0	1	2	3	4	5	6	7	8	Totals
8 - 8.99	1	0	0	0	0	0	0	0	0	1
12 - 12.99	0	1	0	0	0	0	0	0	0	1
13 - 13.99	0	0	0	0	1	0	0	0	0	1
14 - 14.99	0	3	47	23	2	0	0	0	0	75
15 - 15.99	0	0	31	20	2	2	0	0	0	55
16 - 16.99	0	0	14	25	2	3	0	0	0	44
17 - 17.99	0	0	11	16	6	7	0	1	0	41
18 - 18.99	0	0	5	6	14	5	2	0	1	33
19 - 19.99	0	0	2	6	11	10	2	0	0	31
20 - 20.99	0	0	0	5	8	8	3	1	0	25
21 - 21.99	0	0	0	3	1	9	7	2	2	24
22 - 22.99	0	0	0	1	1	6	8	0	0	16
23 - 23.99	0	0	0	1	1	2	1	4	1	10
24 - 24.99	0	0	0	0	2	2	1	1	1	7
25 - 25.99	0	0	0	0	0	1	0	0	4	5
26 - 26.99	0	0	0	0	0	0	0	1	0	1
27 - 27.99	0	0	0	0	0	0	1	0	0	1
Totals	1	4	110	106	51	55	25	10	9	371

Table 12.3: The number of summer flounder assigned to each total length-at-age category for 371 fish sampled for scale age determination in Chesapeake Bay, Virginia during 2014.

							Age								
Interval	1	2	3	4	5	6	7	8	9	10	11	12	13	14	Totals
13 - 13.99	1	0	1	0	0	1	0	0	0	0	0	0	0	0	3
14 - 14.99	3	17	11	6	4	0	1	1	0	0	0	0	0	0	43
15 - 15.99	4	12	12	13	14	5	1	0	0	0	0	0	0	0	61
16 - 16.99	1	7	15	18	16	6	1	0	0	0	0	0	0	0	64
17 - 17.99	0	2	9	22	14	2	0	0	1	0	0	0	0	0	50
18 - 18.99	0	1	6	10	13	4	3	1	0	0	0	0	0	0	38
19 - 19.99	0	1	0	10	7	2	4	1	1	0	0	0	0	0	26
20 - 20.99	0	0	1	8	7	4	2	1	1	0	0	0	0	0	24
21 - 21.99	0	0	3	4	9	5	0	0	0	0	0	0	0	0	21
22 - 22.99	0	0	2	5	9	5	1	1	0	0	0	0	0	0	23
23 - 23.99	0	0	1	3	6	6	5	1	0	0	0	0	0	0	22
24 - 24.99	0	0	0	3	6	4	1	3	0	0	1	0	0	0	18
25 - 25.99	0	0	0	0	1	6	2	2	3	0	0	1	0	0	15
26 - 26.99	0	0	0	0	0	4	1	2	0	1	4	0	0	0	12
27 - 27.99	0	0	0	0	0	1	2	2	1	1	1	0	0	0	8
28 - 28.99	0	0	0	0	0	1	0	1	2	1	1	1	0	0	7
29 - 29.99	0	0	0	0	0	1	3	1	2	0	0	1	1	1	10
30 - 30.99	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1
31 - 31.99	0	0	0	0	0	0	0	0	1	1	0	0	0	0	2
Totals	9	40	61	102	106	57	27	18	12	4	7	3	1	1	448

Table 12.4: The number of summer flounder assigned to each total length-at-age category for 448 fish sampled for scale age determination in Virginia waters of Atlantic ocean during 2014.

							Age								
Interval	0	1	2	3	4	5	6	7	8	9	10	11	13	14	Totals
8 - 8.99	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1
12 - 12.99	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1
13 - 13.99	0	1	0	0	0	0	0	0	0	0	0	0	0	0	1
14 - 14.99	0	2	12	1	1	2	0	0	0	0	0	0	0	0	18
15 - 15.99	0	1	11	4	1	1	1	0	0	0	0	0	0	0	19
16 - 16.99	0	0	11	9	8	4	1	1	0	0	0	0	0	0	34
17 - 17.99	0	0	7	6	6	5	1	1	2	0	0	0	0	0	28
18 - 18.99	0	0	1	7	0	3	0	0	1	0	0	0	0	0	12
19 - 19.99	0	0	1	4	2	3	0	1	1	0	0	1	0	0	13
20 - 20.99	0	0	0	3	5	3	0	0	1	0	0	0	0	0	12
21 - 21.99	0	0	0	3	4	6	2	2	1	0	0	0	0	0	18
22 - 22.99	0	0	0	1	3	6	4	1	0	0	0	0	0	0	15
23 - 23.99	0	0	0	1	1	3	2	3	2	0	1	0	0	1	14
24 - 24.99	0	0	0	0	3	3	4	3	1	0	0	0	0	0	14
25 - 25.99	0	0	0	0	0	4	0	1	1	0	0	0	0	0	6
26 - 26.99	0	0	0	0	0	1	0	4	0	0	0	0	0	0	5
27 - 27.99	0	0	0	0	0	0	0	1	3	0	0	1	0	0	5
28 - 28.99	0	0	0	0	0	1	0	0	0	1	1	1	0	1	5
29 - 29.99	0	0	0	0	0	0	0	2	1	0	2	1	1	0	7
30 - 30.99	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1
31 - 31.99	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1
Totals	1	5	43	39	34	45	15	20	14	1	5	4	2	2	230

Table 12.5: The number of summer flounder assigned to each total length-at-age category for 230 fish sampled for otolith age determination in Chesapeake Bay and Virginia waters of Atlantic Ocean during 2014.

				Age					
Interval	0	1	2	3	4	5	6	7	8
8 - 8.99	1	0	0	0	0	0	0	0	0
12 - 12.99	0	1	0	0	0	0	0	0	0
13 - 13.99	0	0	0	0	1	0	0	0	0
14 - 14.99	0	0.04	0.63	0.31	0.03	0	0	0	0
15 - 15.99	0	0	0.56	0.36	0.04	0.04	0	0	0
16 - 16.99	0	0	0.32	0.57	0.05	0.07	0	0	0
17 - 17.99	0	0	0.27	0.39	0.15	0.17	0	0.02	0
18 - 18.99	0	0	0.15	0.18	0.42	0.15	0.06	0	0.03
19 - 19.99	0	0	0.06	0.19	0.35	0.32	0.06	0	0
20 - 20.99	0	0	0	0.2	0.32	0.32	0.12	0.04	0
21 - 21.99	0	0	0	0.12	0.04	0.38	0.29	0.08	0.08
22 - 22.99	0	0	0	0.06	0.06	0.38	0.5	0	0
23 - 23.99	0	0	0	0.1	0.1	0.2	0.1	0.4	0.1
24 - 24.99	0	0	0	0	0.29	0.29	0.14	0.14	0.14
25 - 25.99	0	0	0	0	0	0.2	0	0	0.8
26 - 26.99	0	0	0	0	0	0	0	1	0
27 - 27.99	0	0	0	0	0	0	1	0	0

Table 12.6: Age-Length key, as proportion-at-age in each 1-inch length interval, based on scale ages for summer flounder sampled in Chesapeake Bay, Virginia during 2014.

	<u> </u>		5	<u> </u>	τ	>	Age	þ		5			5
Interval		2	ω	4	თ	9	7	8	9	10	11	, I	12
13 - 13.99	0.33	0	0.33	0	0	0.33	0	0	0	0	0		0
14 - 14.99	0.07	0.4	0.26	0.14	0.09	0	0.02	0.02	0	0	0		0
15 - 15.99	0.07	0.2	0.2	0.21	0.23	0.08	0.02	0	0	0	0		0
16 - 16.99	0.02	0.11	0.23	0.28	0.25	0.09	0.02	0	0	0	0		0
17 - 17.99	0	0.04	0.18	0.44	0.28	0.04	0	0	0.02	0	0		0
18 - 18.99	0	0.03	0.16	0.26	0.34	0.11	0.08	0.03	0	0	0		0
19 - 19.99	0	0.04	0	0.38	0.27	0.08	0.15	0.04	0.04	0	0		0
20 - 20.99	0	0	0.04	0.33	0.29	0.17	0.08	0.04	0.04	0	0		0
21 - 21.99	0	0	0.14	0.19	0.43	0.24	0	0	0	0	0		0
22 - 22.99	0	0	0.09	0.22	0.39	0.22	0.04	0.04	0	0	0		0
23 - 23.99	0	0	0.05	0.14	0.27	0.27	0.23	0.05	0	0	0		0
24 - 24.99	0	0	0	0.17	0.33	0.22	0.06	0.17	0	0	0.06		0
25 - 25.99	0	0	0	0	0.07	0.4	0.13	0.13	0.2	0	0	0	.07
26 - 26.99	0	0	0	0	0	0.33	0.08	0.17	0	0.08	0.33		0
27 - 27.99	0	0	0	0	0	0.12	0.25	0.25	0.12	0.12	0.12		0
28 - 28.99	0	0	0	0	0	0.14	0	0.14	0.29	0.14	0.14	0	.14
29 - 29.99	0	0	0	0	0	0.1	0.3	0.1	0.2	0	0		0.1
30 - 30.99	0	0	0	0	0	0	0	<u> </u>	0	0	0		0
31 - 31.99	0	0	0	0	0	0	0	0	0.5	0.5	0		0

Table 12.7: Age-Length key, as proportion-at-age in each 1-inch length interval, based on scale ages for summer flounder sampled in Virginia waters

Chapter 13

TAUTOG Tautoga onitis



13.1 INTRODUCTION

We aged a total of 308 tautog, Tautoga onitis, using their opercula collected by the VMRC's Biological Sampling Program in 2014. Of 308 aged fish, 264 and 44 fish were collected in Chesapeake Bay (bay fish) and Virginia waters of the Atlantic Ocean (ocean fish), respectively. The average age for the bay fish was 5.2 years with a standard deviation of 1.4 and a standard error of 0.09. Ten age classes (3 to 12) were represented in the bay fish, comprising fish from the 2002 to 2011 year classes. The bay fish sample in 2014 was dominated by the year class of 2009 with 46%. The average age for the ocean fish was 9.1 years with a standard deviation of 5.1 and a standard error of 0.77. Fifteen age classes (3,5 to 17, 30) were represented in the ocean fish, comprising fish from the 1984, 1997 to 2009, 2011 year classes. The ocean fish sample in 2014 was dominated by the year classes of 2009 and 2008 with 23% and 20%, respectively. We also aged a total of 301 fish using their otoliths in addition to ageing their opercula. The otolith ages were compared to the operculum ages to examine how close both ages were to one another (see details in Results).

13.2 METHODS

13.2.1 Sample size for ageing

We estimated sample sizes for ageing tautog collected in both Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014, respectively, using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 C V^2 + B_a/L} \tag{13.1}$$

where A is the sample size for ageing tautog in 2014; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CVis the coefficient of variation; L was the total number of tautog used by VMRC to estimate length distribution of the catches from 2008 to 2012. θ_a , V_a , B_a , and CV were calculated using pooled age-length data of tautog collected from 2008 to 2012 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (13.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval *l* from the length distribution of the 2008 to 2012 catch. A_l is number of fish to be aged for length interval lin 2014.

13.2.2 Handling of collection

Sagittal otoliths (hereafter, referred to as "otoliths") and opercula were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted based on date of capture. Their envelope labels were verified against VMRC's collection data, and each fish assigned a unique Age and Growth Laboratory identification number. All otoliths and opercula were stored dry within their original labeled coin envelopes; otoliths were contained inside protective Axygen 2.0 ml microtubes.

13.2.3 Preparation

Opercula

Tautog opercula were boiled for several minutes to remove any attached skin and connective tissue. After boiling, opercula were inspected for damage. If there were no obvious flaws, the opercula was dried and then stored in a new, labeled envelope.

Click here to obtain the protocol at the CQFE website on how to prepare operculum for ageing tautog.

Otoliths

We used our thin-section and bake technique to process spadefish sagittal otoliths (hereafter, referred to as "otoliths") for age determination. Otolith preparation began by randomly selecting either the right or left otolith. Each whole otolith was placed in a ceramic "Coors" spot plate well and baked in a Thermolyne 1400 furnace at 400 °C. Baking time was dependent on the otolith's size and gauged by color, with a light caramel color desired. Once a suitable color was achieved the baked otolith was embedded in epoxy resin with its distal surface orientated downwards and allowed to harden overnight. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMetTM low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The otolith was positioned so the blades straddled each side of the otolith focus. It was crucial that this cut be perpendicular to the long axis of the otolith. Failure to do so resulted in broadening and distorted winter growth zones. A proper cut resulted in annuli that were clearly defined and delineated. Once cut, thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing tautog.

13.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of reading the information contained in its otolith, the date of its capture, and the species-specific period when it deposits its annulus. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called annuli. Technically, an otolith annulus is the combination of both the opaque and the translucent bands. In practice, only the opaque bands are counted as annuli. The number of these visible dark bands replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the otolith section is examined for translucent growth. If no translucent growth is visible beyond the last dark annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last dark annulus, a "+" is added to the notation. By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the dark band of the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of dark bands in the otolith. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday", but before the dark band deposition period, is interpreted as being toward the next age class.

For example, tautog otolith deposition occurs between May and July (Hostetter and Munroe 1993). A tautog captured between January 1 and July 31, before the end of the species' annulus formation period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of June 30, the period of annulus formation, which would be noted as 4+4.

Tautog opercula are also considered to have a deposition period of May through July (Hostetter and Munroe 1993), and age class assignment using these hard-parts is conducted in the same way as otoliths.

All tautog samples (opercula and sectioned otoliths) were aged by two different readers in chronological order based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish again without any knowledge of previously estimated ages or lengths, then assigned a final age to the fish. When the age readers were unable to agree on a final age, the fish was excluded from further analysis.

Opercula

All prepared opercula were aged by two different readers in chronological order based on collection date, without knowledge of previously estimated ages or the specimen lengths. Opercula were aged on a light table with no magnification (Figure 13.1).



Figure 13.1: Operculum of a 7 year-old tautog

Otoliths

All otolith thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 13.2). Each reader aged all of the otolith samples.

Click here to obtain the protocol at the CQFE website on how to age tautog using their otolith thin-sections.



Figure 13.2: Otolith thin-section of 6 year-old tautog

13.2.5 Comparison Tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for following comparisons: 1) between the two readers in the current year; 2) within each reader in the current year; 3) timeseries bias between the current and previous years within each reader; and 4) between operculum and otoliths ages. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2000 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

13.3 RESULTS

13.3.1 Sample size

We estimated a sample size of 422 bay tautog in 2014, ranging in length interval from 12 to 26 inches (Table 13.1). This sample size provided a range in CV for age composition approximately from the smallest CVof 9% for age 4 and 5 to the largest CV of 21% for age 8 of the bay fish. We aged 264 of 265 tautog (The rest of fish were either without opercula or over-collected for certain length interval(s)) collected by VMRC in Chesapeake Bay in 2014. We fell short in our over-all collections for this optimal length-class sampling estimate by 163 fish. However, we were short of many fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would definitely be influenced significantly. Therefore, precaution should be used when developing ALK using these age data.

We estimated a sample size of 409 ocean tautog in 2014, ranging in length interval from 11 to 30 inches (Table 13.2). This sample size provided a range in CV for age composition approximately from the smallest CV of 10% for age 4 to the largest CV of 25% for age 14 of the ocean fish. We aged all 44 tautog collected by VMRC in Virginia waters of the Atlantic Ocean in 2014. We fell short in our over-all collections for this optimal length-class sampling estimate by 365 fish. However, we were short of many fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would definitely be influenced significantly. Therefore, precaution should be used when developing ALK using these age data.

13.3.2 Opercula

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 72% (1 year or less agreement of 96%) and a CV of 4.5% (test of symmetry: $\chi^2 = 11.33$, df = 7, P = 0.1247), and there was no significant difference between the first and second readings for Reader 2 with an agreement of

76% (1 year or less agreement of 98%) and a CV of 3.1% (test of symmetry: $\chi^2 = 6$, df = 6, P = 0.4232). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 67% (1 year or less agreement of 94%) and a CVof 4.8% (test of symmetry: $\chi^2 = 30.18$, df = 26, P = 0.2601) (Figure 13.3).



Figure 13.3: Between-reader comparison of operculum age estimates for tautog collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

There was no time-series bias for both readers. Reader 1 had an agreement of 68% (1 year or less agreement of 96%) with ages of fish aged in 2000 with a CV of 4.5% (test of symmetry: $\chi^2 = 7.33$, df = 10, P = 0.6936). Reader 2 had an agreement of 62% (1 year or less agreement of 98%) with a CV of 5.4% (test of symmetry: $\chi^2 = 8.8$, df = 8, P = 0.3594).

Of the 264 bay tautog aged with opercula, 10 age classes (3 to 12) were represented (Table 13.3). The average age for the sample was 5.2 years. The standard deviation and standard error were 1.4 and 0.09, respectively. Year-class data (Figure 13.4) indicates that recruitment into the fishery in Chesapeake Bay begins at age 3, which corresponds to the 2011 year-class for tautog caught in 2014. Tautog in the sample in 2014 was dominated by the year class of 2009 with 46%. The sex ratio of male to female was 1:0.69 for the bay fish.



Figure 13.4: Year-class frequency distribution for tautog collected in Chesapeake Bay, Virginia for ageing in 2014. Distribution is broken down by sex and estimated using operculum ages. 'Unknown' represents the fish gonads that were not available for examination or were not examined for sex during sampling.

Of the 44 ocean tautog aged with opercula, 15 age classes (3, 5 to 17, 30) were represented (Table 13.4). The average age for the sample was 9.1 years. The standard deviation and standard error were 5.1 and 0.77, respectively. Year-class data (Figure 13.5) indicates that recruitment into the fishery in Virginia waters of Atlantic ocean begins at age 3, which corresponds to the 2011 year-class for tautog caught in 2014. Tautog in the sample in 2014 was dominated by the year classes of 2009 and 2008 with 23% and 20%, respectively. The sex ratio of male to female was 1:1.47 for the ocean fish.

13.3.3 Otoliths

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 96% and a CV of 0.5% (test of symmetry: $\chi^2 = 2$, df = 2, P = 0.3679), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 92% and a CV of 1.6% (test of symmetry: $\chi^2 = 2$, df = 3, P = 0.5724).



Figure 13.5: Year-class frequency distribution for tautog collected in Virginia waters of the Atlantic Ocean for ageing in 2014. Distribution is broken down by sex and estimated using operculum ages. 'Unknown' represents the fish gonads that were not available for examination or were not examined for sex during sampling.

There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 86% (1 year or less agreement of 98%) and a CV of 1.8% (test of symmetry: $\chi^2 = 21.94$, df = 18, P =0.2345) (Figure 13.6). There was no time-



Figure 13.6: Between-reader comparison of otolith age estimates for tautog collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

series bias for both readers. Reader 1 had an agreement of 100% with ages of fish aged in 2003. Reader 2 had an agreement of 98% with a CV of 0.2% (test of symmetry: χ^2 = 1, df = 1, P = 0.3173).

Of the 301 tautog aged with otoliths, 17 age classes (2 to 17, 28) were represented (Table 13.5). The average age for the sample was 5.8 years. The standard deviation and standard error were 2.8 and 0.16, respectively.

13.3.4 Comparison of operculum and otolith ages

We aged 301 tautog using opercula and otoliths. There was no evidence of systematic disagreement between otolith and operculum ages (test of symmetry: $\chi^2 =$ 35.6, df = 24, P = 0.0599) with an average CV of 5.2%. There was an agreement of 66% between operculum and otoliths ages whereas opercula were assigned a lower and higher age than otoliths for 17% and 18% of the fish, respectively (Figure 13.7). There was also little evidence of bias between otolith and operculum ages using an age bias plot(Figure 13.8), with no trend of either over-ageing younger or under-ageing older fish.



Figure 13.7: Comparison of operculum and otolith age estimates for tautog collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.



Figure 13.8: Age-bias plot for tautog operculum and otolith age estimates in 2014.

13.3.5 Age-Length-Key (ALK)

We developed an age-length-key for both bay (Table 13.6) and ocean fish (Table 13.7) using operculum ages, separately. The ALK can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using operculum ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

13.4 REFERENCES

- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analyzing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- Hostetter, E. B., and T. A. Munroe. 1993.Age, growth, and reproduction of tautog Tautoga onitis (Labridae: Perciformes) from coastal waters of Virginia. Fishery Bulletin 91: 45-64.

- Quinn, T. J. II, and R. B. Deriso. 1999. Quantitative Fish Dynamics. Oxford University Press. New York.
- R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.

Interval	Target	Collected	Aged	Need
12 - 12.99	5	0	0	5
13 - 13.99	30	0	0	30
14 - 14.99	78	18	18	60
15 - 15.99	80	60	60	20
16 - 16.99	68	74	73	0
17 - 17.99	57	51	51	6
18 - 18.99	41	41	41	0
19 - 19.99	27	13	13	14
20 - 20.99	11	8	8	3
21 - 21.99	5	0	0	5
22 - 22.99	5	0	0	5
23 - 23.99	5	0	0	5
24 - 24.99	5	0	0	5
26 - 26.99	5	0	0	5
Totals	422	265	264	163

Table 13.1: Number of bay tautog collected and aged in each 1-inch length interval in 2014. 'Target' represents the sample size for ageing estimated for 2014, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Table 13.2: Number of ocean tautog collected and aged in each 1-inch length interval in 2014	4.
'Target' represents the sample size for ageing estimated for 2014, and 'Need' represents number of	of
fish shorted in each length interval compared to the optimum sample size for ageing and number of	of
fish aged.	

Interval	Target	Collected	Aged	Need
11 - 11.99	5	0	0	5
13 - 13.99	16	0	0	16
14 - 14.99	47	1	1	46
15 - 15.99	63	6	6	57
16 - 16.99	68	5	5	63
17 - 17.99	37	4	4	33
18 - 18.99	39	3	3	36
19 - 19.99	26	4	4	22
20 - 20.99	21	3	3	18
21 - 21.99	8	2	2	6
22 - 22.99	13	1	1	12
23 - 23.99	10	5	5	5
24 - 24.99	5	4	4	1
25 - 25.99	5	2	2	3
26 - 26.99	26	0	0	26
27 - 27.99	5	2	2	3
28 - 28.99	5	1	1	4
29 - 29.99	5	1	1	4
30 - 30.99	5	0	0	5
Totals	409	44	44	365

					Age						
Interval	3	4	5	6	7	8	9	10	11	12	Totals
14 - 14.99	10	2	6	0	0	0	0	0	0	0	18
15 - 15.99	9	13	34	4	0	0	0	0	0	0	60
16 - 16.99	9	11	39	12	1	1	0	0	0	0	73
17 - 17.99	1	3	24	16	2	5	0	0	0	0	51
18 - 18.99	0	1	14	20	2	4	0	0	0	0	41
19 - 19.99	0	0	4	3	0	4	0	2	0	0	13
20 - 20.99	0	0	0	0	1	2	2	1	1	1	8
Totals	29	30	121	55	6	16	2	3	1	1	264

Table 13.3: The number of tautog assigned to each total length-at-age category for 264 fish sampled for operculum age determination in Chesapeake Bay, Virginia during 2014.

(Go back to $\overline{\text{text}}$)

								Age								
Interval	3	5	6	7	8	9	10	11	12	13	14	15	16	17	30	Totals
14 - 14.99	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
15 - 15.99	0	4	1	1	0	0	0	0	0	0	0	0	0	0	0	6
16 - 16.99	0	3	1	1	0	0	0	0	0	0	0	0	0	0	0	5
17 - 17.99	0	3	1	0	0	0	0	0	0	0	0	0	0	0	0	4
18 - 18.99	0	0	3	0	0	0	0	0	0	0	0	0	0	0	0	3
19 - 19.99	0	0	1	2	1	0	0	0	0	0	0	0	0	0	0	4
20 - 20.99	0	0	2	1	0	0	0	0	0	0	0	0	0	0	0	3
21 - 21.99	0	0	0	0	1	0	1	0	0	0	0	0	0	0	0	2
22 - 22.99	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	1
23 - 23.99	0	0	0	0	0	1	0	0	1	1	1	0	1	0	0	5
24 - 24.99	0	0	0	0	0	0	0	1	0	2	0	1	0	0	0	4
25 - 25.99	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	2
27 - 27.99	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	2
28 - 28.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1
29 - 29.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
Totals	1	10	9	5	2	1	1	2	2	3	2	2	1	2	1	44

Table 13.4: The number of tautog assigned to each total length-at-age category for 44 fish sampled for operculum age determination in Virginia waters of Atlantic ocean during 2014.
									Age									
Interval	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	28	Totals
14 - 14.99	1	9	2	6	0	0	0	0	0	0	0	0	0	0	0	0	0	18
15 - 15.99	0	15	6	37	6	0	0	0	0	0	0	0	0	0	0	0	0	64
16 - 16.99	0	11	9	44	5	4	3	0	0	0	0	0	0	0	0	0	0	76
17 - 17.99	0	2	3	30	12	4	3	1	0	0	0	0	0	0	0	0	0	55
18 - 18.99	0	0	1	19	15	5	3	1	0	0	0	0	0	0	0	0	0	44
19 - 19.99	0	0	0	4	3	2	4	1	2	0	0	0	0	0	0	0	0	16
20 - 20.99	0	0	0	1	1	1	5	0	0	2	1	0	0	0	0	0	0	11
21 - 21.99	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0	2
22 - 22.99	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	1
23 - 23.99	0	0	0	0	0	0	0	0	0	1	0	0	1	2	1	0	0	5
24 - 24.99	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	1	0	3
25 - 25.99	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	2
27 - 27.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	2
28 - 28.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1
29 - 29.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
Totals	1	37	21	141	42	16	18	3	3	4	2	3	1	4	1	3	1	301

Table 13.5: The number of tautog assigned to each total length-at-age category for 301 fish sampled for otolith age determination in Chesapeake Bay and Virginia waters of Atlantic Ocean during 2014.

					Age					
Interval	3	4	5	6	7	8	9	10	11	12
14 - 14.99	0.56	0.11	0.33	0	0	0	0	0	0	0
15 - 15.99	0.15	0.22	0.57	0.07	0	0	0	0	0	0
16 - 16.99	0.12	0.15	0.53	0.16	0.01	0.01	0	0	0	0
17 - 17.99	0.02	0.06	0.47	0.31	0.04	0.1	0	0	0	0
18 - 18.99	0	0.02	0.34	0.49	0.05	0.1	0	0	0	0
19 - 19.99	0	0	0.31	0.23	0	0.31	0	0.15	0	0
20 - 20.99	0	0	0	0	0.12	0.25	0.25	0.12	0.12	0.12

Table 13.6: Age-Length key, as proportion-at-age in each 1-inch length interval, based on operculum ages for tautog sampled in Chesapeake Bay, Virginia during 2014.

							Age								
terval	3	ъ	9	2	∞	6	10	11	12	13	14	15	16	17	30
14.99		0	0	0	0	0	0	0	0	0	0	0	0	0	0
15.99	0	0.67	0.17	0.17	0	0	0	0	0	0	0	0	0	0	0
16.99	0	0.6	0.2	0.2	0	0	0	0	0	0	0	0	0	0	0
17.99	0	0.75	0.25	0	0	0	0	0	0	0	0	0	0	0	0
18.99	0	0	Η	0	0	0	0	0	0	0	0	0	0	0	0
19.99	0	0	0.25	0.5	0.25	0	0	0	0	0	0	0	0	0	0
20.99	0	0	0.67	0.33	0	0	0	0	0	0	0	0	0	0	0
21.99	0	0	0	0	0.5	0	0.5	0	0	0	0	0	0	0	0
22.99	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
23.99	0	0	0	0	0	0.2	0	0	0.2	0.2	0.2	0	0.2	0	0
24.99	0	0	0	0	0	0	0	0.25	0	0.5	0	0.25	0	0	0
25.99	0	0	0	0	0	0	0	0	0.5	0	0	0.5	0	0	0
27.99	0	0	0	0	0	0	0	0	0	0	0.5	0	0	0.5	0
28.99	0	0	0	0	0	0	0	0	0	0	0	0	0	-	0
29.99	С	0	0	0	0	0	0	0	0	С	0	C	C	C	-

Table 13.7: Age-Length key, as proportion-at-age in each 1-inch length interval, based on operculum ages for tautog sampled in Virginia waters of the Atlantic Ocean during 2014.

(Go back to text)

135

Chapter 14

WEAKFISH Cynoscion regalis



14.1 INTRODUCTION

We aged a total of 295 weakfish, *Cynoscion* regalis, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2014. The weakfish ages ranged from 1 to 6 years old with an average age of 2.7, a standard deviation of 1, and a standard error of 0.06. Six age classes (1 to 6) were represented, comprising fish of the 2008 to 2013 year-classes. The sample was dominated by fish from the year-classes of 2011 and 2012 with 38.3% and 32.9%, respectively.

14.2 METHODS

14.2.1 Sample size for ageing

We estimated sample size for ageing weakfish in 2014 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 C V^2 + B_a/L} \tag{14.1}$$

where A is the sample size for ageing weakfish in 2014; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of weakfish used by VMRC to estimate length distribution of the catches from 2008 to 2012. θ_a , V_a , B_a , and CV were calculated using pooled agelength data of weakfish collected from 2008 to 2012 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (14.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A

138

should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2008 to 2012 catch. A_l is number of fish to be aged for length interval l in 2014.

14.2.2 Handling of collections

Otoliths were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

14.2.3 Preparation

Sagittal otoliths, hereafter, referred to as "otoliths", were processed for age determination following the methods described in Lowerre-Barbieri et al. (1994) with a few modifications. The left or right otolith was randomly selected and attached, distal side down, to a glass slide with clear CrystalbondTM 509 adhesive. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a pencil across the otolith surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMetTM low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). Thinsections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-sections.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing weakfish.

14.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the speciesspecific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, weakfish annulus formation occurs between April and July (Lowerre-Barbieri et al. 1994 and modified by CQFE). A weakfish captured between January 1 and July 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of July 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 14.1). Each reader aged all of the otolith samples. All sam-



Figure 14.1: Otolith thin-section of 4 year-old weakfish

ples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age weakfish using their otolith thin-sections.

14.2.5Comparison tests

A symmetry test (Hoenig et al. 1995)and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 2.15.2 (R Core Team 2012).

14.3RESULTS

14.3.1Sample size

We estimated a sample size of 333 for ageing weakfish in 2014, ranging in length interval from 6 to 35 inches (Table 14.1). This sample size provided a range in (CV)for age composition approximately from the smallest (CV) of 6% for age 2 to the

largest (CV) of 23% for age 4. In 2014, we randomly selected and aged 295 fish from 377 weakfish collected by VMRC. We fell short in our over-all collections for this optimal length-class sampling estimate by 53 fish. However, we were short of no fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

14.3.2**Reading** precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 100%, and there was no significant difference between the first and second readings for Reader 2 with an agreement of 100%. There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 100% (Figure 14.2).



Figure 14.2: Between-reader comparison of otolith age estimates for weakfish collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2014.

There was no time-series bias for both readers. Reader 1 had an agreement of 98%with ages of fish aged in 2003 with a CV of 0.2% (test of symmetry: $\chi^2 = 1, df = 1, P$ = 0.3173). Reader 2 had an agreement of

98% with a CV of 0.2% (test of symmetry: $\chi^2 = 1, df = 1, P = 0.3173$).

14.3.3 Year class

Of the 295 fish aged with otoliths, 6 age classes (1 to 6) were represented (Table 14.2). The average age was 2.7 years, and the standard deviation and standard error were 1 and 0.06, respectively. Year-class data show that the fishery was comprised of 6 year-classes: fish from the 2008 to 2013 year-classes, with fish primarily from the year-classes of 2011 and 2012 with 38.3% and 32.9%, respectively. The ratio of males to females was 1:1.81 in the sample collected (Figure 14.3).



Figure 14.3: Year-class frequency distribution for weakfish collected for ageing in 2014. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

14.3.4 Age-length key

We developed an age-length-key (Table 14.3) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

14.4 REFERENCES

- Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.
- Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analyzing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.
- Lowerre-Barbieri, S.K., M.E. Chittenden Jr., and C.M. Jones. 1994. A comparison of a validated otolith method to age weakfish, Cynoscion regalis, with the traditional scale method. Fish Bull. 92:555-568.
- Quinn, T. J. II, and R. B. Deriso. 1999. Quantitative Fish Dynamics. Oxford University Press. New York.
- R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.

Interval	Target	Collected	Aged	Need
6 - 6.99	5	1	1	4
7 - 7.99	5	24	6	0
8 - 8.99	5	18	6	0
9 - 9.99	29	45	30	0
10 - 10.99	58	71	58	0
11 - 11.99	41	47	42	0
12 - 12.99	27	33	28	0
13 - 13.99	19	22	20	0
14 - 14.99	15	19	16	0
15 - 15.99	16	20	16	0
16 - 16.99	12	13	12	0
17 - 17.99	8	11	8	0
18 - 18.99	8	9	8	0
19 - 19.99	5	6	6	0
20 - 20.99	5	3	3	2
21 - 21.99	5	1	1	4
22 - 22.99	5	7	7	0
23 - 23.99	5	4	4	1
24 - 24.99	5	1	1	4
25 - 25.99	5	5	5	0
26 - 26.99	5	8	8	0
27 - 27.99	5	7	7	0
28 - 28.99	5	1	1	4
29 - 29.99	5	1	1	4
30 - 30.99	5	0	0	5
31 - 31.99	5	0	0	5
32 - 32.99	5	0	0	5
33 - 33.99	5	0	0	5
34 - 34.99	5	0	0	5
35 - 35.99	5	0	0	5
Totals	333	377	295	53

Table 14.1: Number of weakfish collected and aged in each 1-inch length interval in 2014. 'Target' represents the sample size for ageing estimated for 2014, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

			Age				
Interval	1	2	3	4	5	6	Totals
6 - 6.99	1	0	0	0	0	0	1
7 - 7.99	6	0	0	0	0	0	6
8 - 8.99	3	3	0	0	0	0	6
9 - 9.99	5	23	2	0	0	0	30
10 - 10.99	7	29	21	1	0	0	58
11 - 11.99	1	15	23	3	0	0	42
12 - 12.99	1	12	12	3	0	0	28
13 - 13.99	1	7	9	3	0	0	20
14 - 14.99	0	2	8	6	0	0	16
15 - 15.99	0	3	7	6	0	0	16
16 - 16.99	0	1	10	1	0	0	12
17 - 17.99	0	0	3	5	0	0	8
18 - 18.99	0	1	5	2	0	0	8
19 - 19.99	0	1	2	3	0	0	6
20 - 20.99	0	0	3	0	0	0	3
21 - 21.99	0	0	1	0	0	0	1
22 - 22.99	0	0	4	3	0	0	7
23 - 23.99	0	0	2	2	0	0	4
24 - 24.99	0	0	0	1	0	0	1
25 - 25.99	0	0	1	2	2	0	5
26 - 26.99	0	0	0	6	2	0	8
27 - 27.99	0	0	0	2	5	0	7
28 - 28.99	0	0	0	0	0	1	1
29 - 29.99	0	0	0	0	1	0	1
Totals	25	97	113	49	10	1	295

Table 14.2: The number of weakfish assigned to each total length-at-age category for 295 fish sampled for otolith age determination in Virginia during 2014.

			Age			
Interval	1	2	3	4	5	6
6 - 6.99	1	0	0	0	0	0
7 - 7.99	1	0	0	0	0	0
8 - 8.99	0.5	0.5	0	0	0	0
9 - 9.99	0.17	0.77	0.07	0	0	0
10 - 10.99	0.12	0.5	0.36	0.02	0	0
11 - 11.99	0.02	0.36	0.55	0.07	0	0
12 - 12.99	0.04	0.43	0.43	0.11	0	0
13 - 13.99	0.05	0.35	0.45	0.15	0	0
14 - 14.99	0	0.12	0.5	0.38	0	0
15 - 15.99	0	0.19	0.44	0.38	0	0
16 - 16.99	0	0.08	0.83	0.08	0	0
17 - 17.99	0	0	0.38	0.62	0	0
18 - 18.99	0	0.12	0.62	0.25	0	0
19 - 19.99	0	0.17	0.33	0.5	0	0
20 - 20.99	0	0	1	0	0	0
21 - 21.99	0	0	1	0	0	0
22 - 22.99	0	0	0.57	0.43	0	0
23 - 23.99	0	0	0.5	0.5	0	0
24 - 24.99	0	0	0	1	0	0
25 - 25.99	0	0	0.2	0.4	0.4	0
26 - 26.99	0	0	0	0.75	0.25	0
27 - 27.99	0	0	0	0.29	0.71	0
28 - 28.99	0	0	0	0	0	1
29 - 29.99	0	0	0	0	1	0

Table 14.3: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for weakfish sampled for age determination in Virginia during 2014.