2015 Final Report Virginia ~ Chesapeake Bay Finfish Ageing and Population Analysis

Hongsheng Liao, Cynthia M. Jones,
& Jessica L. Gilmore

September 28, 2016

VMRC ~ ODU Age and Growth Laboratory
Center for Quantitative Fisheries Ecology
Old Dominion University
Norfolk, Virginia 23508

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Virginia and Chesapeake Bay Finfish Ageing and Population Analysis

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Contents

EXECUTIVE SUMMARY

In this report we present the ageing results of 14 finfish species collected from commercial and recreational catches made in the Chesapeake Bay and Virginia waters of the Atlantic Ocean, U.S.A. in 2015. All fish were collected by the Virginia Marine Resources Commission's (VMRC) Stock Assessment Program and the Center for Quantitative Fisheries Ecology (CQFE) at Old Dominion University in 2015 and aged in 2016 at the Age and Growth Laboratory of CQFE. This report is broken into chapters, one for each of the 14 species. We present measures of ageing precision, graphs of year-class distributions, and age-length keys for each species.

Three calcified structures (hard-parts) are used in age determination. Specifically, two calcified structures were used for determining fish ages of the following three species: striped bass, *Morone saxatilis*, (n = 885); summer flounder, *Paralichthys dentatus*, (n = 884); and tautog, *Tautoga onitis*, (n = 279). Scales and otoliths were used to age striped bass and summer flounder, opercula and otoliths were used to age tautog. Comparing alternative hard-parts allowed us to assess their usefulness in determining fish age as well as the relative precision of each structure. Ages were determined from otoliths only for the following species: Atlantic croaker, *Micropogonias undulatus*, (n = 357); black drum, *Pogonias cromis*, (n = 69); bluefish, *Pomatomus saltatrix*, (n = 442); cobia, *Rachycentron canadum*, (n = 342); red drum, *Sciaenops ocellatus*, (n = 31); sheepshead, *Archosargus probatocephalus*, (n = 119); Atlantic spadefish, *Chaetodipterus faber*, (n = 135); Spanish mackerel, *Scomberomorous maculates*, (n = 231); spot, *Leiostomus xanthurus*, (n = 201); spotted seatrout, *Cynoscion nebulosus*, (n = 308); and weakfish, *Cynoscion regalis*, (n = 243). In total, we made 10,850 age readings from scales, otoliths and opercula collected during 2015. A summary of the age ranges for all species aged is presented in Table 1.

In this report, we also present sample sizes and coefficient of variation (CV) for estimates of age composition for the following species: Atlantic croaker, bluefish, spadefish, Spanish mackerel, spot, spotted seatrout, striped bass, summer flounder, tautog, and weakfish. The sample sizes and the CVs enabled us to determine how many fish we needed to age in each length interval and to measure the precision for estimates of major age classes in each species, respectively, enhancing our efficiency and effectiveness on ageing those species.

To support environmental and wildlife agencies, and charities, we donated more than 4800 pounds of dissected fish to the Salvation Army to feed the homeless, and the Wildlife Response, Inc., a local wildlife rescue agency which is responsible for saving injured animals found by the public.

In 2015, we continued to upgrade our Age and Growth Laboratory website, which can be accessed at http://odu.edu/sci/research/cqfe/research/ageing-lab. The website includes an electronic version of this document and our previous VMRC final reports from 2001 to 2014. The site also provides more detailed explanations of the methods and structures we use in age determination.

In order to share the VMRC/ODU data and findings with the stakeholders and other fisheries biologists, in 2015, we developed two website applications (apps) and posted them at the CQFE website. The first one is called "Fish Age Estimator" and designed to estimate the probabilities of ages given a length of a fish (Click here to open the app). The second one is called "Sample Size Estimator for Ageing" and designed to help fisheries biologists and educators to estimate necessary sample sizes with certain coefficients of variation (CV) (Click here to open the app).

Table 1: The minimum and maximum ages, number of fish and their hard-parts collected, number of fish aged, and age readings for the 14 finfish species in 2015. The hard-parts and age readings include both scales and otoliths for striped bass and summer flounder, and both opercula and otoliths for tautog.

Species	Number of fish collected	Number of hard-parts	Number of fish aged	Number of readings	Minimum age	Maximum age
Atlantic Croaker	464	464	357	714	1	10
Black Drum	69	69	69	138	3	53
Bluefish	682	678	442	884	0	13
Cobia	350	343	342	684	2	11
Red Drum	31	31	31	62	1	6
Sheepshead	121	119	119	238	2	32
Spadefish	137	135	135	270	1	8
Spanish Mackerel	330	327	231	462	0	8
Spotted Seatrout	328	328	308	616	0	10
Spot	263	263	201	402	0	4
Striped Bass	1,081	1,387	885	2,432	3	22
Summer Flounder	1,095	1,386	884	2,356	1	15
Tautog	279	548	279	1,106	3	31
Weakfish	283	283	243	486	1	5
Totals	5,513	6,361	4,526	10,850		

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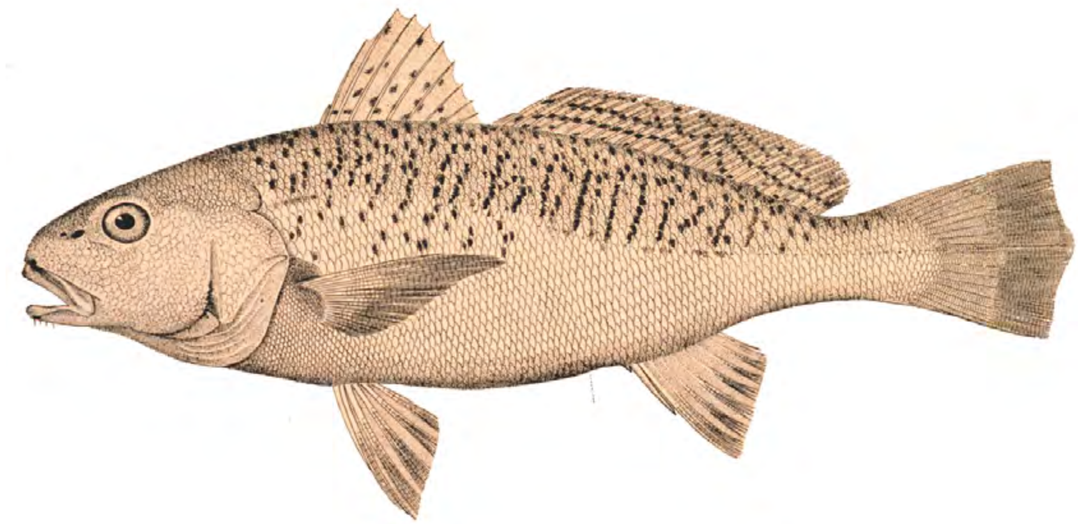
ACKNOWLEDGEMENTS

We thank Alicia Brown, Aris Horsey, Allison Roberts, Haxley Gomez Vigil, and James Black for their technical expertise in preparing otoliths, scales, and opercula for age determination. They all put in long hours processing "tons" of fish in our lab. A special note of appreciation is extended to Joe Grist, Joe Cimino, Adam Kenyon, and their technicians at the VMRC, including Richard Hancock, Myra Thompson, and Chris Williams for their many efforts in this cooperative project. We would like also to thank our graduate students Kathleen Kirch and Mike Schmidtke for their help in processing fish whenever we were short of hands.

Chapter 1

ATLANTIC CROAKER
Micropogonias undulatus



1.1 INTRODUCTION

We aged a total of 357 Atlantic croaker, *Micropogonias undulatus*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2015. Croaker ages ranged from 1 to 10 years old with an average age of 4.1, a standard deviation of 1.7, and a standard error of 0.09. Ten age classes (1 to 10) were represented, comprising fish of the 2005 to 2014 year-classes. The sample was dominated by fish from the year-class of 2012 with 52.4%.

1.2 METHODS

1.2.1 Sample size for ageing

We estimated sample size for ageing croaker in 2015 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 CV^2 + B_a/L} \tag{1.1}$$

where A is the sample size for ageing croaker in 2015; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of croaker used by VMRC to estimate length distribution of the catches from 2009 to 2013. θ_a, V_a, B_a, and CV were calculated using pooled age-length data of croaker collected from 2009 to 2013 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (1.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by ageing an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2009 to 2013 catch. A_l is number of fish to be aged for length interval l in 2015.

1.2.2 Handling of collections

Otoliths were received by the Age & and Growth Laboratory in labeled coin envelopes, were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

1.2.3 Preparation

Sagittal otoliths, hereafter, referred to as "otoliths", were processed for age determination following the methods described in Barbieri et al. (1994) with a few modifications. The left or right otolith was randomly selected and attached, distal side down, to a glass slide with clear Crystalbond^{TM} 509 adhesive or imbedded in epoxy. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a pencil across the otolith surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet^{TM} low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). Thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-sections.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing Atlantic croaker.

1.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, Atlantic croaker otolith annulus formation occurs between April and May (Barbieri et al. 1994a and b). A croaker captured between January 1 and May 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification. Each reader aged all of the otolith samples.

Due to discrepancy on identification of the first annulus of Atlantic croaker among Atlantic states, Atlantic States Marine Fisheries Commission (ASMFC) has decided not to count the smallest annulus at the center of the thin-section as the first annulus. Following ASMFC's instruction, we didn't count the smallest annulus at the center as the first annulus in 2015 (Figure 1.1) .

All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the

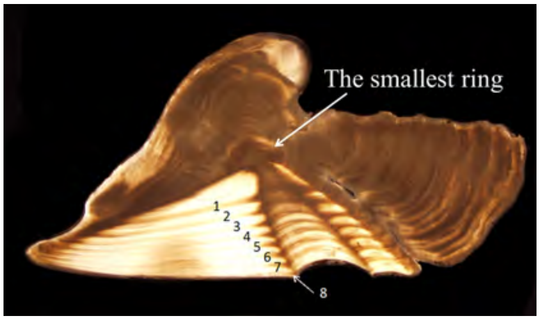


Figure 1.1: Otolith thin-sections of a 8 year-old croaker without counting the smallest ring and with the last annulus on the edge of the thin-section

fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age Atlantic croaker using their otolith thin-sections.

1.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

1.3 RESULTS

1.3.1 Sample size

We estimated a sample size of 418 Atlantic croaker in 2015, ranging in length interval from 5 to 20 inches (Table 1.1). This sample size provided a range in (CV) for age composition approximately from the smallest (CV) of 9% for age 4 and 5 to the largest (CV) of 25% for age 1. In 2015, we randomly selected and aged 357 fish from 464 croaker collected by VMRC. We fell short in our over-all collections for this optimal length-class sampling estimate by 74 fish. However, we were short of some fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would possibly be influenced significantly.

1.3.2 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 96% and a CV of 1% (test of symmetry: $\chi^2 = 2$, $df = 2$, $P = 0.3679$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 96% and a CV of 0.7% (test of symmetry: $\chi^2 = 2$, $df = 2$, $P = 0.3679$). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 94.96% and a CV of 0.9% (test of symmetry: $\chi^2 = 5.29$, $df = 6$, $P = 0.5077$) (Figure 1.2).

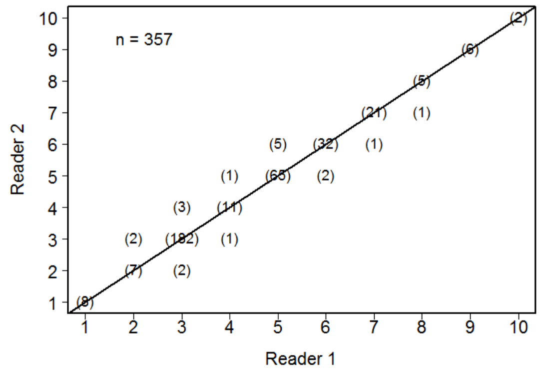


Figure 1.2: Between-reader comparison of otolith age estimates for Atlantic croaker collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

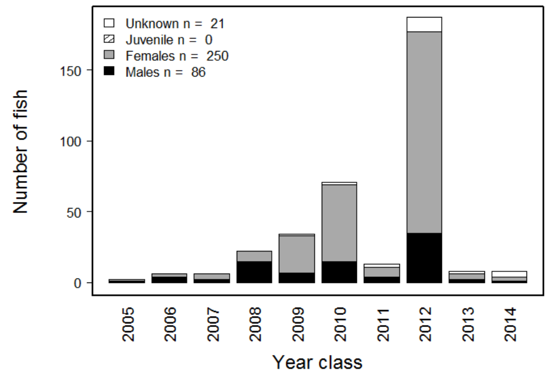


Figure 1.3: Year-class frequency distribution for Atlantic croaker collected for ageing in 2015. Distribution is broken down by sex. 'Unknown' is for gonads that were not available for examination or were not examined for sex during sampling.

There was no time-series bias for both readers. Reader 1 had an agreement of 100% with ages of fish aged in 2003. Reader 2 had an agreement of 100% .

1.3.3 Year class

Of the 357 fish aged with otoliths, 10 age classes (1 to 10) were represented (Table 1.2). The average age was 4.1 years, and the standard deviation and standard error were 1.7 and 0.09, respectively. Year-class data show that the fishery was comprised of 10 year-classes: fish from the 2005 to 2014 year-classes, with fish primarily from the year class of 2012 with 52.4%. The ratio of males to females was 1:2.91 in the sample collected (Figure 1.3).

1.3.4 Age-length key

We developed an age-length-key (Table 1.3) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

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Table 1.1: Number of Atlantic croaker collected and aged in each 1-inch length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
5 - 5.99	5	1	1	4
6 - 6.99	5	12	6	0
7 - 7.99	6	15	6	0
8 - 8.99	6	27	6	0
9 - 9.99	17	38	18	0
10 - 10.99	28	52	28	0
11 - 11.99	51	63	53	0
12 - 12.99	90	105	90	0
13 - 13.99	66	77	75	0
14 - 14.99	54	44	44	10
15 - 15.99	37	25	25	12
16 - 16.99	23	5	5	18
17 - 17.99	15	0	0	15
18 - 18.99	5	0	0	5
19 - 19.99	5	0	0	5
20 - 20.99	5	0	0	5
Totals	418	464	357	74

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Table 1.2: The number of Atlantic croaker assigned to each total length-at-age category for 357 fish sampled for otolith age determination in Virginia during 2015.

	Age										
Interval	1	2	3	4	5	6	7	8	9	10	Totals
5 - 5.99	1	0	0	0	0	0	0	0	0	0	1
6 - 6.99	6	0	0	0	0	0	0	0	0	0	6
7 - 7.99	1	1	4	0	0	0	0	0	0	0	6
8 - 8.99	0	1	3	1	1	0	0	0	0	0	6
9 - 9.99	0	2	15	0	0	1	0	0	0	0	18
10 - 10.99	0	1	15	2	8	2	0	0	0	0	28
11 - 11.99	0	1	37	3	5	3	2	1	0	1	53
12 - 12.99	0	1	47	4	22	6	8	0	1	1	90
13 - 13.99	0	0	29	0	19	15	6	3	3	0	75
14 - 14.99	0	0	23	1	9	5	4	1	1	0	44
15 - 15.99	0	1	12	2	4	2	2	1	1	0	25
16 - 16.99	0	0	2	0	3	0	0	0	0	0	5
Totals	8	8	187	13	71	34	22	6	6	2	357

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Table 1.3: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for Atlantic croaker sampled for age determination in Virginia during 2015.

	Age									
Interval	1	2	3	4	5	6	7	8	9	10
5 - 5.99	1	0	0	0	0	0	0	0	0	0
6 - 6.99	1	0	0	0	0	0	0	0	0	0
7 - 7.99	0.17	0.17	0.67	0	0	0	0	0	0	0
8 - 8.99	0	0.17	0.5	0.17	0.17	0	0	0	0	0
9 - 9.99	0	0.11	0.83	0	0	0.06	0	0	0	0
10 - 10.99	0	0.04	0.54	0.07	0.29	0.07	0	0	0	0
11 - 11.99	0	0.02	0.7	0.06	0.09	0.06	0.04	0.02	0	0.02
12 - 12.99	0	0.01	0.52	0.04	0.24	0.07	0.09	0	0.01	0.01
13 - 13.99	0	0	0.39	0	0.25	0.2	0.08	0.04	0.04	0
14 - 14.99	0	0	0.52	0.02	0.2	0.11	0.09	0.02	0.02	0
15 - 15.99	0	0.04	0.48	0.08	0.16	0.08	0.08	0.04	0.04	0
16 - 16.99	0	0	0.4	0	0.6	0	0	0	0	0

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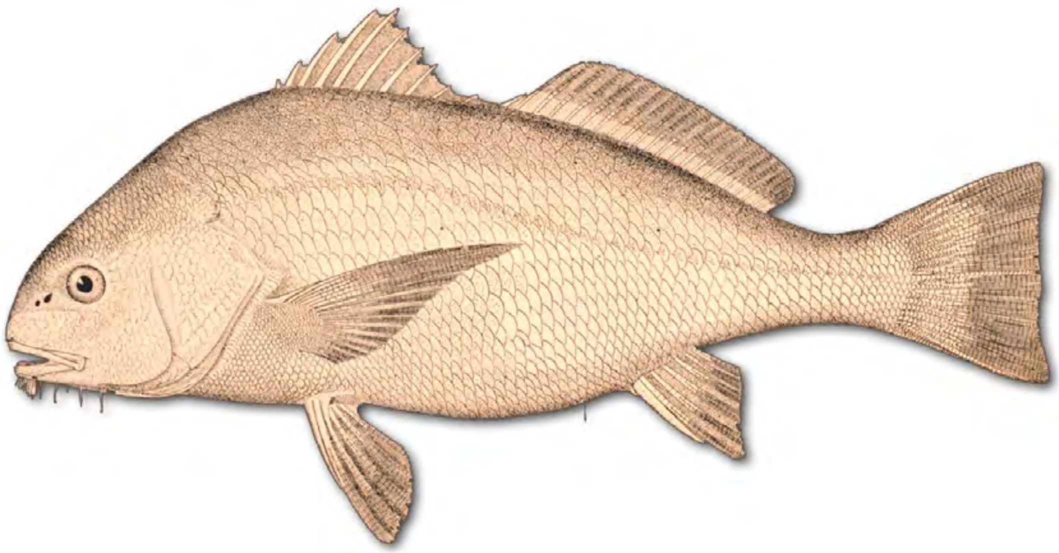
Chapter 2

BLACK DRUM *Pogonias cromis*



2.1 INTRODUCTION

We aged a total of 69 black drum, *Pogonias cromis*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2015. Black drum ages ranged from 3 to 53 years old with an average age of 14.9, a standard deviation of 12.5, and a standard error of 1.5. Twenty-five age classes (3 to 19, 21, 25, 38, 40, 42, 44 to 45, and 53) were represented, comprising fish of the 1962, 1970 to 1971, 1973, 1975, 1977, 1990, 1994, and 1996 to 2012 year-classes. The sample was dominated by fish from the year-classes of 2011, 1999, and 2007 with 14.5%, 14.5%, and 11.6%, respectively.

2.2 METHODS

2.2.1 Handling of collections

Sagittal otoliths, hereafter, referred to as "otoliths", were received by the Age and Growth Laboratory in labeled coin envelopes. In the lab they were sorted by date of capture, their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

2.2.2 Preparation

Otoliths were processed for age determination following the methods described in Bobko (1991) and Jones and Wells (1998). The left or right sagittal otolith was randomly selected and attached, distal side down, to a glass slide with Crystalbond™ 509 adhesive or embedded in epoxy. The otoliths were viewed by eye, and when necessary, under a stereo microscope to identify the location of the core, and the position of the core marked using a pencil across the otolith surface. At least one transverse cross-section (hereafter "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet™ low-speed saw equipped with two, three inch diameter, Norton Diamond Grinding Wheels, separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The position of the marked core fell within the 0.5 mm space between the blades, such that the core was included in the removed thin-section. Otolith thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the sections.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing black drum.

2.2.3 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, black drum otolith annulus formation occurs between May and June (Beckman et al. 1990; Bobko 1991; Jones and Wells 1998). A black drum captured between January 1 and June 30, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 2.1). Each reader aged all of the otolith samples.

All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or

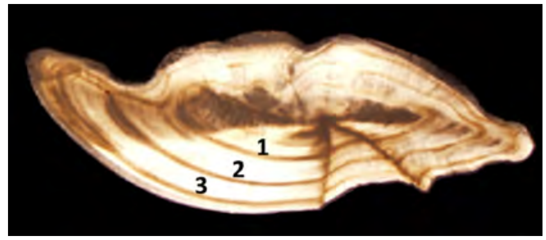


Figure 2.1: Otolith thin-section of a 3 year-old black drum

the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age black drum using their otolith thin-sections.

2.2.4 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

2.3 RESULTS

2.3.1 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 70% and a *CV* of 3.8% (test of symmetry: $\chi^2 = 13$, $df = 14$, $P = 0.5265$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 96% and a *CV* of 0.2% (test of symmetry: $\chi^2 = 2$, $df = 2$, $P = 0.3679$). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 89.86% and a *CV* of 0.3% (test of symmetry: $\chi^2 = 7$, $df = 7$, $P = 0.4289$) (Figure 2.2).

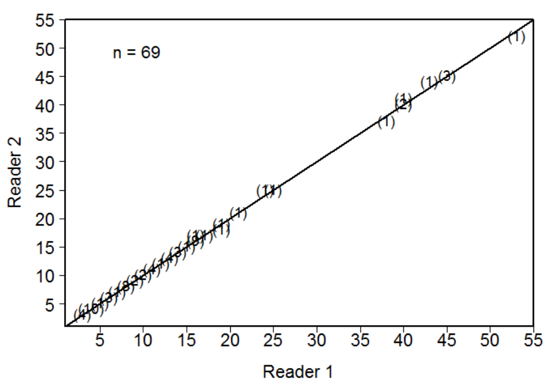


Figure 2.2: Between-reader comparison of otolith age estimates for black drum collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

There was no time-series bias for both readers. Reader 1 had an agreement of 70% with ages of fish aged in 2000 with a *CV* of 2% (test of symmetry: $\chi^2 = 15$, $df = 15$, $P = 0.4514$). Reader 2 had an agreement of 94%with a *CV* of 0.1% (test of symmetry: $\chi^2 = 3$, $df = 3$, $P = 0.3916$).

2.3.2 Year class

Of the 69 fish aged with otoliths, 25 age classes (3 to 19, 21, 25, 38, 40, 42, 44 to 45, and 53) were represented (Table 2.1). The average age was 14.9 years, and the standard deviation and standard error were 12.5 and 1.5, respectively. Year-class data show that the fishery was comprised of 25 year-classes: fish from the 1962, 1970 to 1971, 1973, 1975, 1977, 1990, 1994, and 1996 to 2012 year-classes, with fish primarily from the year classes of 2011, 1999, and 2007 with 14.5%, 14.5%, and 11.6%, respectively. The ratio of males to females was 1:0.64 in the sample collected (Figure 2.3).

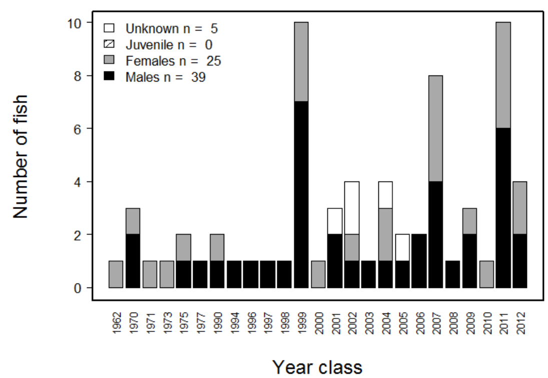


Figure 2.3: Year-class frequency distribution for black drum collected for ageing in 2015. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

2.3.3 Age-length key

We developed an age-length-key (Table 2.2) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

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Table 2.1: The number of black drum assigned to each total length (inch)-at-age category for 69 fish sampled for otolith age determination in Virginia during 2015.

	Age																								
Interval	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	21	25	38	40	42	44	45	53	Totals
21 - 21.99	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
22 - 22.99	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
24 - 24.99	0	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	2
25 - 25.99	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
26 - 26.99	2	3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	5
27 - 27.99	0	4	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	5
29 - 29.99	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
31 - 31.99	0	0	0	2	0	2	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	5
32 - 32.99	0	0	0	0	1	4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	5
33 - 33.99	0	0	0	0	0	1	0	1	0	0	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	4
34 - 34.99	0	0	0	0	0	1	2	0	2	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	6
35 - 35.99	0	0	0	0	0	0	0	0	0	1	0	0	0	2	0	0	0	0	0	0	0	0	0	0	0	3
36 - 36.99	0	0	0	0	0	0	0	0	1	0	1	1	0	1	0	0	1	0	0	0	0	0	0	0	0	5
37 - 37.99	0	0	0	0	0	0	0	0	1	0	1	2	0	1	1	0	0	0	0	0	0	0	0	0	0	6
38 - 38.99	0	0	0	0	0	0	0	0	0	0	1	0	0	2	0	0	0	0	0	0	0	0	0	0	0	3
39 - 39.99	0	0	0	0	0	0	0	0	0	0	0	0	0	2	0	0	0	0	0	0	0	0	0	0	0	2
40 - 40.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	0	0	0	0	0	0	0	3
41 - 41.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1
42 - 42.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1
44 - 44.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	0	2
45 - 45.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	2	1	5
49 - 49.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	2
Totals	4	10	1	3	1	8	2	2	4	1	4	3	1	10	1	1	1	1	2	1	2	1	1	3	1	69

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Table 2.2: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for black drum sampled for age determination in Virginia during 2015.

	Age																								
Interval	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	21	25	38	40	42	44	45	53
21 - 21.99	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
22 - 22.99	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
24 - 24.99	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
25 - 25.99	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
26 - 26.99	0.4	0.6	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
27 - 27.99	0	0.8	0.2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
29 - 29.99	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
31 - 31.99	0	0	0	0.4	0	0.4	0	0.2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
32 - 32.99	0	0	0	0	0.2	0.8	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
33 - 33.99	0	0	0	0	0	0.25	0	0.25	0	0	0	0	0.25	0.25	0	0	0	0	0	0	0	0	0	0	0
34 - 34.99	0	0	0	0	0	0.17	0.33	0	0.33	0	0.17	0	0	0	0	0	0	0	0	0	0	0	0	0	0
35 - 35.99	0	0	0	0	0	0	0	0	0	0.33	0	0	0	0.67	0	0	0	0	0	0	0	0	0	0	0
36 - 36.99	0	0	0	0	0	0	0	0	0.2	0	0.2	0.2	0	0.2	0	0	0.2	0	0	0	0	0	0	0	0
37 - 37.99	0	0	0	0	0	0	0	0	0.17	0	0.17	0.33	0	0.17	0.17	0	0	0	0	0	0	0	0	0	0
38 - 38.99	0	0	0	0	0	0	0	0	0	0	0.33	0	0	0.67	0	0	0	0	0	0	0	0	0	0	0
39 - 39.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0
40 - 40.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0.33	0	0.33	0	0.33	0	0	0	0	0	0	0
41 - 41.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
42 - 42.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
44 - 44.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0.5	0	0	0	0	0.5	0
45 - 45.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0.2	0.2	0	0	0.4	0.2
49 - 49.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0.5	0	0.5	0	0

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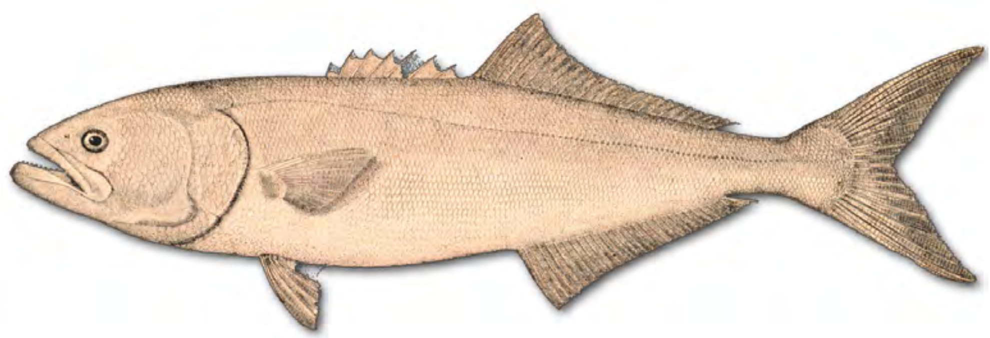
Chapter 3

BLUEFISH *Pomatomus saltatrix*



3.1 INTRODUCTION

We aged a total of 442 bluefish, *Pomatomus saltatrix*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2015. Bluefish ages ranged from 0 to 13 years old with an average age of 3.1, a standard deviation of 2.6, and a standard error of 0.12. Fourteen age classes (0 to 13) were represented, comprising fish of the 2002 to 2015 year-classes. The sample was dominated by fish from the year-classes of 2013, 2014, and 2012 with 25.3%, 22.4%, and 14.9%, respectively.

3.2 METHODS

3.2.1 Sample size for ageing

We estimated sample size for ageing bluefish in 2015 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 CV^2 + B_a/L} \tag{3.1}$$

where A is the sample size for ageing bluefish in 2015; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of bluefish used by VMRC to estimate length distribution of the catches from 2009 to 2013. θ_a, V_a, B_a, and CV were calculated using pooled age-length data of bluefish collected from 2009 to 2013 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (3.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2009 to 2013 catch. A_l is number of fish to be aged for length interval l in 2015. Based on VMRC's request in 2010, we used 1-cm length interval for bluefish, which differed from other species (1-inch).

3.2.2 Handling of collections

Otoliths were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

3.2.3 Preparation

We used our thin-section and bake technique to process bluefish sagittal otoliths (hereafter, referred to as "otoliths") for age determination (Robillard et al. 2009). Otolith preparation began by randomly selecting either the right or left otolith. Each whole otolith was placed in a ceramic "Coors" spot plate well and baked in a Thermolyne 1400 furnace at 400 °C. Baking time was dependent on the otolith's size and gauged by color, with a light caramel color desired. Once a suitable color was achieved the baked otolith was embedded in epoxy resin with its distal surface orientated downwards. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core. Then, the position of the core was marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet^{TM} low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The otolith was positioned so the blades straddled each side of the otolith focus. It was crucial that this cut be perpendicular to the long axis of the otolith. Failure to do so resulted in broad and distorted winter growth zones. A proper cut resulted in annuli that were clearly defined and delineated. Once cut, thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing bluefish.

3.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, bluefish otolith deposition occurs between March and June (Robillard et al. 2009). A bluefish captured between January 1 and May 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 3.1). Each reader aged all of the otolith samples.

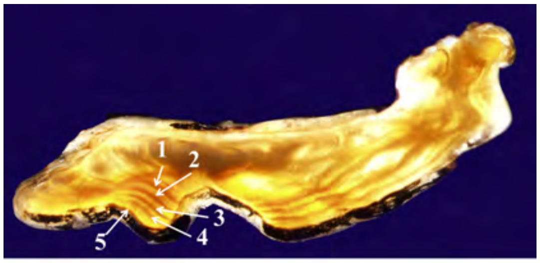


Figure 3.1: Otolith thin-section of a 5 year-old bluefish with the last annulus on the edge of the thin-section

If an otolith was properly sectioned, the sulcal groove came to a sharp point within the middle of the focus. Typically the first year's annulus was found by locating the focus of the otolith, which was characterized as a visually distinct dark, oblong region found in the center of the otolith. The first year's annulus had the highest visibility proximal to the focus along the edge of the sulcal groove. Once located, the first year's annulus was followed outward from the sulcal groove towards the dorsal perimeter of the otolith. Often, but not always, the first year was associated with a very distinct crenellation on the dorsal surface and a prominent protrusion on the ventral surface. Both of these landmarks had a tendency to become less prominent in older fish.

Even with the bake and thin-section technique, interpretation of the growth zones from the otoliths of young bluefish was difficult. Rapid growth within the first year of life prevents a sharp delineation between opaque and translucent zones. When the exact location of the first year was not clearly evident, and the otolith had been sectioned accurately, a combination of surface landscape (1st year crenellation) and the position of the second annuli were used to help determine the position of the first annulus.

What appeared to be "double annuli" were occasionally observed in bluefish 4-7 years of age and older. This double-annulus formation was typically characterized by distinct and separate annuli in extremely close proximity to each other. We do not know if the formation of these double annuli were two separate annuli, or in fact only one, but they seemed to occur during times of reduced growth after maturation. "Double annuli" were considered to be one annulus when both marks joined to form a central origin (the origin being the sulcal groove and the outer peripheral edge of the otolith). If these annuli did not meet to form a central origin they were considered two distinct annuli, and were counted as such.

All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age bluefish using their otolith thin-sections.

3.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

3.3 RESULTS

3.3.1 Sample size

We estimated a sample size of 459 bluefish in 2015, ranging in length interval from 14 to 99 centimeters (Table 3.1). This sample size provided a range in (CV) for age composition approximately from the smallest (CV) of 6% for age 1 and 2 to the largest (CV) of 25% for age 8. In 2015, we randomly selected and aged 442 fish from 678 bluefish collected by VMRC. We fell short in our over-all collections for this optimal length-class sampling estimate by 71 fish. However, we were short of no fish from the major length intervals (The interval requires more than 5 fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

3.3.2 Reading precision

Reader 1 had high self-precision and Read 2 had moderate self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 96% and a CV of 3% (test of symmetry: $\chi^2 = 2$, $df = 2$, $P = 0.3679$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 92% and a CV of 9.4% (test of symmetry: $\chi^2 = 4$, $df = 2$, $P = 0.1353$). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 86.2% and a CV of 3% (test of symmetry: $\chi^2 = 19.09$, $df = 18$, $P = 0.3864$) (Figure 3.2).

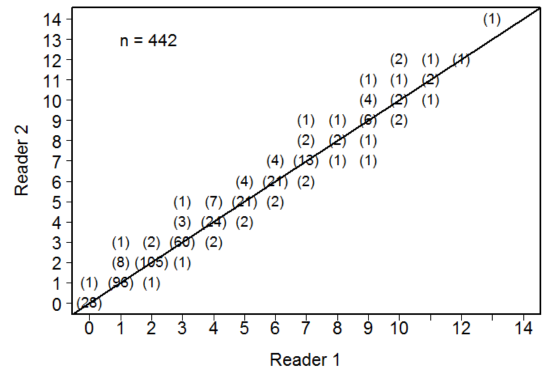


Figure 3.2: Between-reader comparison of otolith age estimates for bluefish collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

There was no time-series bias for both readers. Reader 1 had an agreement of 94% with ages of fish aged in 2000 with a CV of 2.5% (test of symmetry: $\chi^2 = 3$, $df = 3$, $P = 0.3916$). Reader 2 had an agreement of 94% with a CV of 4.2% (test of symmetry: $\chi^2 = 3$, $df = 3$, $P = 0.3916$).

3.3.3 Year class

Of the 442 fish aged with otoliths, 14 age classes (0 to 13) were represented (Table 3.2). The average age was 3.1 years, and the standard deviation and standard error were 2.6 and 0.12, respectively. Year-class data show that the fishery was comprised of 14 year-classes: fish from the 2002 to 2015 year-classes, with fish primarily from the year classes of 2013, 2014, and 2012 with 25.3%, 22.4%, and 14.9%, respectively. The ratio of males to females was 1:1.66 in the sample collected (Figure 3.3).

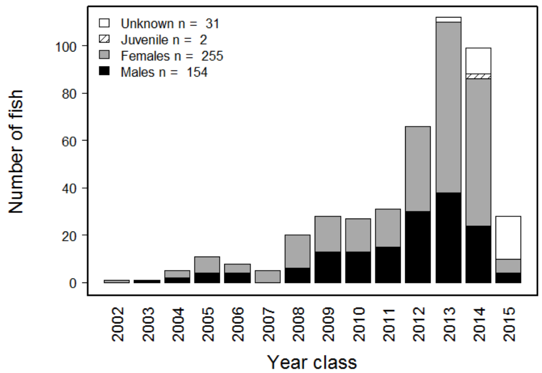


Figure 3.3: Year-class frequency distribution for bluefish collected for ageing in 2015. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

3.3.4 Age-length key

We developed an age-length-key (Table 3.3) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length cm intervals.

3.4 REFERENCES

Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.

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Table 3.1: Number of bluefish collected and aged in each 1-cm length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

(Go back to text)

Interval	Target	Collected	Aged	Need
14 - 14.99	5	0	0	5
15 - 15.99	5	0	0	5
16 - 16.99	5	0	0	5
17 - 17.99	5	3	3	2
18 - 18.99	5	3	3	2
19 - 19.99	5	3	3	2
20 - 20.99	5	16	6	0
21 - 21.99	5	11	6	0
22 - 22.99	5	17	6	0
23 - 23.99	5	12	6	0
24 - 24.99	5	12	6	0
25 - 25.99	5	13	6	0
26 - 26.99	5	15	6	0
27 - 27.99	5	12	6	0
28 - 28.99	5	11	6	0
29 - 29.99	5	14	6	0
30 - 30.99	5	11	6	0
31 - 31.99	5	10	6	0
32 - 32.99	5	11	6	0
33 - 33.99	5	13	6	0
34 - 34.99	5	7	6	0
35 - 35.99	6	13	6	0
36 - 36.99	7	11	8	0
37 - 37.99	7	9	8	0
38 - 38.99	7	16	8	0
39 - 39.99	8	11	8	0
40 - 40.99	8	17	8	0
41 - 41.99	7	11	8	0
42 - 42.99	8	18	8	0
43 - 43.99	7	10	8	0
44 - 44.99	6	16	6	0
45 - 45.99	8	13	8	0
46 - 46.99	7	18	8	0
47 - 47.99	7	12	8	0
48 - 48.99	6	11	6	0
49 - 49.99	5	10	6	0
50 - 50.99	5	7	6	0
51 - 51.99	5	6	6	0
52 - 52.99	5	5	5	0
53 - 53.99	5	9	6	0

(To continue)

Table 3.1 (Continued)

Interval	Target	Collected	Aged	Need
54 - 54.99	5	4	4	1
55 - 55.99	5	8	6	0
56 - 56.99	5	6	6	0
57 - 57.99	5	8	6	0
58 - 58.99	5	7	6	0
59 - 59.99	5	10	6	0
60 - 60.99	5	11	6	0
61 - 61.99	5	10	6	0
62 - 62.99	5	7	7	0
63 - 63.99	5	4	4	1
64 - 64.99	5	4	4	1
65 - 65.99	5	6	6	0
66 - 66.99	5	3	3	2
67 - 67.99	5	2	2	3
68 - 68.99	5	12	6	0
69 - 69.99	5	7	6	0
70 - 70.99	5	13	6	0
71 - 71.99	5	9	6	0
72 - 72.99	5	5	5	0
73 - 73.99	5	7	6	0
74 - 74.99	5	10	6	0
75 - 75.99	5	6	6	0
76 - 76.99	5	8	6	0
77 - 77.99	5	8	6	0
78 - 78.99	5	7	6	0
79 - 79.99	5	11	6	0
80 - 80.99	5	2	2	3
81 - 81.99	5	6	6	0
82 - 82.99	5	7	5	0
83 - 83.99	5	8	7	0
84 - 84.99	5	4	4	1
85 - 85.99	5	4	4	1
86 - 86.99	5	5	5	0
87 - 87.99	5	5	5	0
88 - 88.99	5	9	9	0
89 - 89.99	5	1	1	4
90 - 90.99	5	4	4	1
91 - 91.99	5	1	1	4
92 - 92.99	5	5	5	0
93 - 93.99	5	1	1	4
94 - 94.99	5	2	2	3
95 - 95.99	5	1	1	4
96 - 96.99	5	1	1	4
97 - 97.99	5	1	1	4
98 - 98.99	5	1	1	4

(To continue)

Table 3.1 (Continued)

Interval	Target	Collected	Aged	Need
99 - 99.99	5	0	0	5
Totals	459	678	442	71

(Go back to text)

Table 3.2: The number of bluefish assigned to each total length (cm)-at-age category for 442 fish sampled for otolith age determination in Virginia during 2015.

	Age														
Interval	0	1	2	3	4	5	6	7	8	9	10	11	12	13	Totals
17 - 17.99	3	0	0	0	0	0	0	0	0	0	0	0	0	0	3
18 - 18.99	2	1	0	0	0	0	0	0	0	0	0	0	0	0	3
19 - 19.99	2	1	0	0	0	0	0	0	0	0	0	0	0	0	3
20 - 20.99	3	3	0	0	0	0	0	0	0	0	0	0	0	0	6
21 - 21.99	3	3	0	0	0	0	0	0	0	0	0	0	0	0	6
22 - 22.99	3	3	0	0	0	0	0	0	0	0	0	0	0	0	6
23 - 23.99	3	3	0	0	0	0	0	0	0	0	0	0	0	0	6
24 - 24.99	3	3	0	0	0	0	0	0	0	0	0	0	0	0	6
25 - 25.99	1	5	0	0	0	0	0	0	0	0	0	0	0	0	6
26 - 26.99	2	3	1	0	0	0	0	0	0	0	0	0	0	0	6
27 - 27.99	2	4	0	0	0	0	0	0	0	0	0	0	0	0	6
28 - 28.99	1	4	1	0	0	0	0	0	0	0	0	0	0	0	6
29 - 29.99	0	5	1	0	0	0	0	0	0	0	0	0	0	0	6
30 - 30.99	0	6	0	0	0	0	0	0	0	0	0	0	0	0	6
31 - 31.99	0	6	0	0	0	0	0	0	0	0	0	0	0	0	6
32 - 32.99	0	5	1	0	0	0	0	0	0	0	0	0	0	0	6
33 - 33.99	0	4	2	0	0	0	0	0	0	0	0	0	0	0	6
34 - 34.99	0	4	2	0	0	0	0	0	0	0	0	0	0	0	6
35 - 35.99	0	3	2	1	0	0	0	0	0	0	0	0	0	0	6
36 - 36.99	0	4	4	0	0	0	0	0	0	0	0	0	0	0	8
37 - 37.99	0	4	4	0	0	0	0	0	0	0	0	0	0	0	8
38 - 38.99	0	2	5	1	0	0	0	0	0	0	0	0	0	0	8
39 - 39.99	0	4	3	1	0	0	0	0	0	0	0	0	0	0	8
40 - 40.99	0	3	5	0	0	0	0	0	0	0	0	0	0	0	8
41 - 41.99	0	4	4	0	0	0	0	0	0	0	0	0	0	0	8
42 - 42.99	0	2	6	0	0	0	0	0	0	0	0	0	0	0	8
43 - 43.99	0	2	5	1	0	0	0	0	0	0	0	0	0	0	8
44 - 44.99	0	2	4	0	0	0	0	0	0	0	0	0	0	0	6
45 - 45.99	0	1	7	0	0	0	0	0	0	0	0	0	0	0	8
46 - 46.99	0	0	8	0	0	0	0	0	0	0	0	0	0	0	8
47 - 47.99	0	0	8	0	0	0	0	0	0	0	0	0	0	0	8
48 - 48.99	0	2	4	0	0	0	0	0	0	0	0	0	0	0	6
49 - 49.99	0	0	6	0	0	0	0	0	0	0	0	0	0	0	6
50 - 50.99	0	0	4	2	0	0	0	0	0	0	0	0	0	0	6
51 - 51.99	0	0	4	2	0	0	0	0	0	0	0	0	0	0	6
52 - 52.99	0	1	0	4	0	0	0	0	0	0	0	0	0	0	5
53 - 53.99	0	0	1	5	0	0	0	0	0	0	0	0	0	0	6
54 - 54.99	0	1	1	2	0	0	0	0	0	0	0	0	0	0	4
55 - 55.99	0	0	1	5	0	0	0	0	0	0	0	0	0	0	6
56 - 56.99	0	0	1	4	1	0	0	0	0	0	0	0	0	0	6
57 - 57.99	0	0	3	2	1	0	0	0	0	0	0	0	0	0	6

(To continue)

Table 3.2 (Continued)

	Age														
Interval	0	1	2	3	4	5	6	7	8	9	10	11	12	13	Totals
58 - 58.99	0	0	2	4	0	0	0	0	0	0	0	0	0	0	6
59 - 59.99	0	1	2	1	2	0	0	0	0	0	0	0	0	0	6
60 - 60.99	0	0	3	3	0	0	0	0	0	0	0	0	0	0	6
61 - 61.99	0	0	2	3	1	0	0	0	0	0	0	0	0	0	6
62 - 62.99	0	0	3	2	2	0	0	0	0	0	0	0	0	0	7
63 - 63.99	0	0	0	1	2	1	0	0	0	0	0	0	0	0	4
64 - 64.99	0	0	0	3	1	0	0	0	0	0	0	0	0	0	4
65 - 65.99	0	0	1	5	0	0	0	0	0	0	0	0	0	0	6
66 - 66.99	0	0	1	1	1	0	0	0	0	0	0	0	0	0	3
67 - 67.99	0	0	0	0	2	0	0	0	0	0	0	0	0	0	2
68 - 68.99	0	0	0	3	1	2	0	0	0	0	0	0	0	0	6
69 - 69.99	0	0	0	2	3	1	0	0	0	0	0	0	0	0	6
70 - 70.99	0	0	0	3	2	1	0	0	0	0	0	0	0	0	6
71 - 71.99	0	0	0	1	4	0	1	0	0	0	0	0	0	0	6
72 - 72.99	0	0	0	3	1	1	0	0	0	0	0	0	0	0	5
73 - 73.99	0	0	0	0	1	3	2	0	0	0	0	0	0	0	6
74 - 74.99	0	0	0	1	1	2	1	1	0	0	0	0	0	0	6
75 - 75.99	0	0	0	0	2	2	2	0	0	0	0	0	0	0	6
76 - 76.99	0	0	0	0	0	5	1	0	0	0	0	0	0	0	6
77 - 77.99	0	0	0	0	1	3	2	0	0	0	0	0	0	0	6
78 - 78.99	0	0	0	0	1	3	2	0	0	0	0	0	0	0	6
79 - 79.99	0	0	0	0	1	0	5	0	0	0	0	0	0	0	6
80 - 80.99	0	0	0	0	0	1	1	0	0	0	0	0	0	0	2
81 - 81.99	0	0	0	0	0	0	2	3	1	0	0	0	0	0	6
82 - 82.99	0	0	0	0	0	1	0	3	1	0	0	0	0	0	5
83 - 83.99	0	0	0	0	0	1	3	3	0	0	0	0	0	0	7
84 - 84.99	0	0	0	0	0	0	2	1	1	0	0	0	0	0	4
85 - 85.99	0	0	0	0	0	0	1	1	1	1	0	0	0	0	4
86 - 86.99	0	0	0	0	0	0	1	2	0	1	0	1	0	0	5
87 - 87.99	0	0	0	0	0	0	0	4	0	0	1	0	0	0	5
88 - 88.99	0	0	0	0	0	0	2	0	0	3	4	0	0	0	9
89 - 89.99	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1
90 - 90.99	0	0	0	0	0	0	0	2	1	1	0	0	0	0	4
91 - 91.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
92 - 92.99	0	0	0	0	0	0	0	0	0	1	2	1	1	0	5
93 - 93.99	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1
94 - 94.99	0	0	0	0	0	0	0	0	0	0	1	1	0	0	2
95 - 95.99	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1
96 - 96.99	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1
97 - 97.99	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1
98 - 98.99	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1
Totals	28	99	112	66	31	27	28	20	5	8	11	5	1	1	442

(Go back to text)

Table 3.3: Age-Length key, as proportion-at-age in each 1-cm length interval, based on otolith ages for bluefish sampled for age determination in Virginia during 2015.

	Age													
Interval	0	1	2	3	4	5	6	7	8	9	10	11	12	13
17 - 17.99	1	0	0	0	0	0	0	0	0	0	0	0	0	0
18 - 18.99	0.67	0.33	0	0	0	0	0	0	0	0	0	0	0	0
19 - 19.99	0.67	0.33	0	0	0	0	0	0	0	0	0	0	0	0
20 - 20.99	0.5	0.5	0	0	0	0	0	0	0	0	0	0	0	0
21 - 21.99	0.5	0.5	0	0	0	0	0	0	0	0	0	0	0	0
22 - 22.99	0.5	0.5	0	0	0	0	0	0	0	0	0	0	0	0
23 - 23.99	0.5	0.5	0	0	0	0	0	0	0	0	0	0	0	0
24 - 24.99	0.5	0.5	0	0	0	0	0	0	0	0	0	0	0	0
25 - 25.99	0.17	0.83	0	0	0	0	0	0	0	0	0	0	0	0
26 - 26.99	0.33	0.5	0.17	0	0	0	0	0	0	0	0	0	0	0
27 - 27.99	0.33	0.67	0	0	0	0	0	0	0	0	0	0	0	0
28 - 28.99	0.17	0.67	0.17	0	0	0	0	0	0	0	0	0	0	0
29 - 29.99	0	0.83	0.17	0	0	0	0	0	0	0	0	0	0	0
30 - 30.99	0	1	0	0	0	0	0	0	0	0	0	0	0	0
31 - 31.99	0	1	0	0	0	0	0	0	0	0	0	0	0	0
32 - 32.99	0	0.83	0.17	0	0	0	0	0	0	0	0	0	0	0
33 - 33.99	0	0.67	0.33	0	0	0	0	0	0	0	0	0	0	0
34 - 34.99	0	0.67	0.33	0	0	0	0	0	0	0	0	0	0	0
35 - 35.99	0	0.5	0.33	0.17	0	0	0	0	0	0	0	0	0	0
36 - 36.99	0	0.5	0.5	0	0	0	0	0	0	0	0	0	0	0
37 - 37.99	0	0.5	0.5	0	0	0	0	0	0	0	0	0	0	0
38 - 38.99	0	0.25	0.62	0.12	0	0	0	0	0	0	0	0	0	0
39 - 39.99	0	0.5	0.38	0.12	0	0	0	0	0	0	0	0	0	0
40 - 40.99	0	0.38	0.62	0	0	0	0	0	0	0	0	0	0	0
41 - 41.99	0	0.5	0.5	0	0	0	0	0	0	0	0	0	0	0
42 - 42.99	0	0.25	0.75	0	0	0	0	0	0	0	0	0	0	0
43 - 43.99	0	0.25	0.62	0.12	0	0	0	0	0	0	0	0	0	0
44 - 44.99	0	0.33	0.67	0	0	0	0	0	0	0	0	0	0	0
45 - 45.99	0	0.12	0.88	0	0	0	0	0	0	0	0	0	0	0
46 - 46.99	0	0	1	0	0	0	0	0	0	0	0	0	0	0
47 - 47.99	0	0	1	0	0	0	0	0	0	0	0	0	0	0
48 - 48.99	0	0.33	0.67	0	0	0	0	0	0	0	0	0	0	0
49 - 49.99	0	0	1	0	0	0	0	0	0	0	0	0	0	0
50 - 50.99	0	0	0.67	0.33	0	0	0	0	0	0	0	0	0	0
51 - 51.99	0	0	0.67	0.33	0	0	0	0	0	0	0	0	0	0
52 - 52.99	0	0.2	0	0.8	0	0	0	0	0	0	0	0	0	0
53 - 53.99	0	0	0.17	0.83	0	0	0	0	0	0	0	0	0	0
54 - 54.99	0	0.25	0.25	0.5	0	0	0	0	0	0	0	0	0	0
55 - 55.99	0	0	0.17	0.83	0	0	0	0	0	0	0	0	0	0
56 - 56.99	0	0	0.17	0.67	0.17	0	0	0	0	0	0	0	0	0
57 - 57.99	0	0	0.5	0.33	0.17	0	0	0	0	0	0	0	0	0

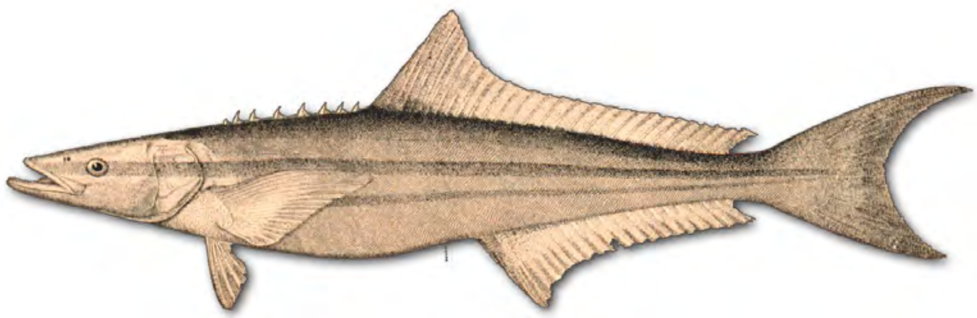
(To continue)

Table 3.3 (Continued)

	Age													
Interval	0	1	2	3	4	5	6	7	8	9	10	11	12	13
58 - 58.99	0	0	0.33	0.67	0	0	0	0	0	0	0	0	0	0
59 - 59.99	0	0.17	0.33	0.17	0.33	0	0	0	0	0	0	0	0	0
60 - 60.99	0	0	0.5	0.5	0	0	0	0	0	0	0	0	0	0
61 - 61.99	0	0	0.33	0.5	0.17	0	0	0	0	0	0	0	0	0
62 - 62.99	0	0	0.43	0.29	0.29	0	0	0	0	0	0	0	0	0
63 - 63.99	0	0	0	0.25	0.5	0.25	0	0	0	0	0	0	0	0
64 - 64.99	0	0	0	0.75	0.25	0	0	0	0	0	0	0	0	0
65 - 65.99	0	0	0.17	0.83	0	0	0	0	0	0	0	0	0	0
66 - 66.99	0	0	0.33	0.33	0.33	0	0	0	0	0	0	0	0	0
67 - 67.99	0	0	0	0	1	0	0	0	0	0	0	0	0	0
68 - 68.99	0	0	0	0.5	0.17	0.33	0	0	0	0	0	0	0	0
69 - 69.99	0	0	0	0.33	0.5	0.17	0	0	0	0	0	0	0	0
70 - 70.99	0	0	0	0.5	0.33	0.17	0	0	0	0	0	0	0	0
71 - 71.99	0	0	0	0.17	0.67	0	0.17	0	0	0	0	0	0	0
72 - 72.99	0	0	0	0.6	0.2	0.2	0	0	0	0	0	0	0	0
73 - 73.99	0	0	0	0	0.17	0.5	0.33	0	0	0	0	0	0	0
74 - 74.99	0	0	0	0.17	0.17	0.33	0.17	0.17	0	0	0	0	0	0
75 - 75.99	0	0	0	0	0.33	0.33	0.33	0	0	0	0	0	0	0
76 - 76.99	0	0	0	0	0	0.83	0.17	0	0	0	0	0	0	0
77 - 77.99	0	0	0	0	0.17	0.5	0.33	0	0	0	0	0	0	0
78 - 78.99	0	0	0	0	0.17	0.5	0.33	0	0	0	0	0	0	0
79 - 79.99	0	0	0	0	0.17	0	0.83	0	0	0	0	0	0	0
80 - 80.99	0	0	0	0	0	0.5	0.5	0	0	0	0	0	0	0
81 - 81.99	0	0	0	0	0	0	0.33	0.5	0.17	0	0	0	0	0
82 - 82.99	0	0	0	0	0	0.2	0	0.6	0.2	0	0	0	0	0
83 - 83.99	0	0	0	0	0	0.14	0.43	0.43	0	0	0	0	0	0
84 - 84.99	0	0	0	0	0	0	0.5	0.25	0.25	0	0	0	0	0
85 - 85.99	0	0	0	0	0	0	0.25	0.25	0.25	0.25	0	0	0	0
86 - 86.99	0	0	0	0	0	0	0.2	0.4	0	0.2	0	0.2	0	0
87 - 87.99	0	0	0	0	0	0	0	0.8	0	0	0.2	0	0	0
88 - 88.99	0	0	0	0	0	0	0.22	0	0	0.33	0.44	0	0	0
89 - 89.99	0	0	0	0	0	0	0	0	0	0	1	0	0	0
90 - 90.99	0	0	0	0	0	0	0	0.5	0.25	0.25	0	0	0	0
91 - 91.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1
92 - 92.99	0	0	0	0	0	0	0	0	0	0.2	0.4	0.2	0.2	0
93 - 93.99	0	0	0	0	0	0	0	0	0	0	0	1	0	0
94 - 94.99	0	0	0	0	0	0	0	0	0	0	0.5	0.5	0	0
95 - 95.99	0	0	0	0	0	0	0	0	0	0	0	1	0	0
96 - 96.99	0	0	0	0	0	0	0	0	0	0	1	0	0	0
97 - 97.99	0	0	0	0	0	0	0	0	0	0	1	0	0	0
98 - 98.99	0	0	0	0	0	0	0	0	0	1	0	0	0	0

Chapter 4

COBIA *Rachycentron canadum*



4.1 INTRODUCTION

We aged a total of 342 cobia, *Rachycentron canadum*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2015. Cobia ages ranged from 2 to 11 years old with an average age of 4.7, a standard deviation of 1.6, and a standard error of 0.09. Ten age classes (2 to 11) were represented, comprising fish of the 2004 to 2013 year-classes. The sample was dominated by fish from the year-classes of 2010, 2012, and 2011 with 35.7%, 23.7%, and 23.7%, respectively.

4.2 METHODS

4.2.1 Handling of collections

Sagittal otoliths, hereafter, referred to as "otoliths", were received by the Age and Growth Laboratory in labeled coin envelopes and were sorted by date of capture, their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored inside of protective Axygen 2 ml micro-tubes within their original labeled coin envelopes.

4.2.2 Preparation

Otoliths were processed for age determination. The left or right otolith was randomly selected and embedded, distal side down, in epoxy resin and allowed to harden overnight. The otoliths were viewed by eye, and when necessary, under a stereo microscope to identify the location of the core, and the position of the core marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet™ low-speed saw equipped with two, three inch diameter, Norton Diamond Grinding Wheels, separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The position of the marked core fell within the 0.5 mm space between the blades, such that the core was included in the removed thin section. Otolith thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing cobia.

4.2.3 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, cobia otolith annulus deposition occurs between June and July (Richards 1967 and modified by CQFE). A cobia captured between January 1 and July 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of July 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 4.1). Each reader aged all of the otolith samples. All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

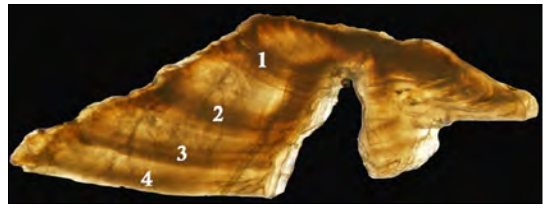


Figure 4.1: Otolith thin-section of a 4 year-old cobia.

Click here to obtain the protocol at the CQFE website on how to age cobia using their otolith thin-sections.

4.2.4 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

4.3 RESULTS

4.3.1 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 98% and a *CV* of 0.4% (test of symmetry: $\chi^2 = 1$, $df = 1$, $P = 0.3173$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 98% and a *CV* of 0.4% (test of symmetry: $\chi^2 = 1$, $df = 1$, $P = 0.3173$). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 94.74% and a *CV* of 0.8% (test of symmetry: $\chi^2 = 2.83$, $df = 5$, $P = 0.7257$) (Figure 4.2).

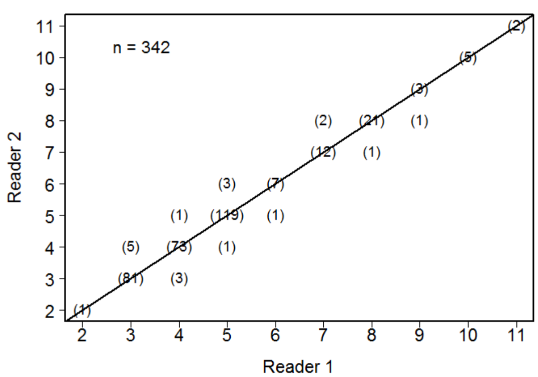


Figure 4.2: Between-reader comparison of otolith age estimates for cobia collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

There was no time-series bias for both readers. Reader 1 had an agreement of 82% with ages of fish aged in 2000 with a *CV* of 1.8% (test of symmetry: $\chi^2 = 9$, $df = 8$, $P = 0.3423$). Reader 2 had an agreement of 88% with a *CV* of 1.5% (test of symmetry: $\chi^2 = 6$, $df = 5$, $P = 0.3062$).

4.3.2 Year class

Of the 342 fish aged with otoliths, 10 age classes (2 to 11) were represented (Table 4.1). The average age was 4.7 years, and the standard deviation and standard error were 1.6 and 0.09, respectively. Year-class data show that the fishery was comprised of 10 year-classes: fish from the 2004 to 2013 year-classes, with fish primarily from the year classes of 2010, 2012, and 2011 with 35.7%, 23.7%, and 23.7%, respectively. The ratio of males to females was 1:0.99 in the sample collected (Figure 4.3).

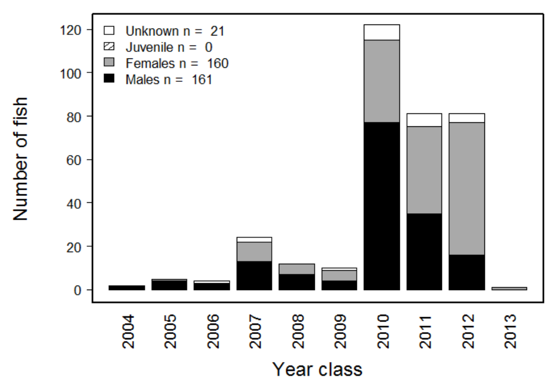


Figure 4.3: Year-class frequency distribution for cobia collected for ageing in 2015. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

4.3.3 Age-length key

We developed an age-length-key (Table 4.2) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

4.4 REFERENCES

Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.

Hoenig, J.M., M.J. Morgan, and C.A. Brown. 1995. Analysing differences between two age determination methods by tests of symmetry. Can. J. Fish. Aquat. Sci. 52:364-368.

R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.

Richards, C.E. 1967. Age, Growth, and Fecundity of the Cobia, Rachycentron canadum, from Chesapeake Bay and Adjacent Waters. Contribution No. 252, Virginia Institute of Marine Science, Gloucester Point, Virginia, 343-350.

Table 4.1: The number of cobia assigned to each total length (inch)-at-age category for 342 fish sampled for otolith age determination in Virginia during 2015.

	Age										
Interval	2	3	4	5	6	7	8	9	10	11	Totals
34 - 34.99	0	0	1	0	0	0	0	0	0	0	1
35 - 35.99	1	4	2	0	0	0	0	0	0	0	7
36 - 36.99	0	12	7	2	0	0	0	0	0	0	21
37 - 37.99	0	15	3	1	1	0	0	0	0	0	20
38 - 38.99	0	18	11	4	1	0	0	0	0	0	34
39 - 39.99	0	12	10	6	0	1	0	0	0	0	29
40 - 40.99	0	11	5	25	0	0	1	0	0	0	42
41 - 41.99	0	1	5	19	1	3	1	1	0	0	31
42 - 42.99	0	4	7	12	0	1	1	0	0	0	25
43 - 43.99	0	2	2	6	1	1	1	1	0	0	14
44 - 44.99	0	0	8	6	0	1	2	1	0	0	18
45 - 45.99	0	2	6	6	1	0	1	0	2	0	18
46 - 46.99	0	0	5	4	0	0	0	0	0	1	10
47 - 47.99	0	0	4	5	1	0	3	0	0	0	13
48 - 48.99	0	0	2	4	0	0	3	0	0	1	10
49 - 49.99	0	0	1	4	0	0	2	0	1	0	8
50 - 50.99	0	0	1	8	0	0	1	0	0	0	10
51 - 51.99	0	0	0	7	1	1	1	0	1	0	11
52 - 52.99	0	0	1	1	2	0	0	0	0	0	4
53 - 53.99	0	0	0	1	1	1	1	0	0	0	4
54 - 54.99	0	0	0	1	0	0	0	1	0	0	2
55 - 55.99	0	0	0	0	0	1	3	0	0	0	4
56 - 56.99	0	0	0	0	0	2	0	0	0	0	2
58 - 58.99	0	0	0	0	0	0	1	0	0	0	1
61 - 61.99	0	0	0	0	0	0	2	0	0	0	2
64 - 64.99	0	0	0	0	0	0	0	0	1	0	1
Totals	1	81	81	122	10	12	24	4	5	2	342

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Table 4.2: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for cobia sampled for age determination in Virginia during 2015.

	Age									
Interval	2	3	4	5	6	7	8	9	10	11
34 - 34.99	0	0	1	0	0	0	0	0	0	0
35 - 35.99	0.14	0.57	0.29	0	0	0	0	0	0	0
36 - 36.99	0	0.57	0.33	0.1	0	0	0	0	0	0
37 - 37.99	0	0.75	0.15	0.05	0.05	0	0	0	0	0
38 - 38.99	0	0.53	0.32	0.12	0.03	0	0	0	0	0
39 - 39.99	0	0.41	0.34	0.21	0	0.03	0	0	0	0
40 - 40.99	0	0.26	0.12	0.6	0	0	0.02	0	0	0
41 - 41.99	0	0.03	0.16	0.61	0.03	0.1	0.03	0.03	0	0
42 - 42.99	0	0.16	0.28	0.48	0	0.04	0.04	0	0	0
43 - 43.99	0	0.14	0.14	0.43	0.07	0.07	0.07	0.07	0	0
44 - 44.99	0	0	0.44	0.33	0	0.06	0.11	0.06	0	0
45 - 45.99	0	0.11	0.33	0.33	0.06	0	0.06	0	0.11	0
46 - 46.99	0	0	0.5	0.4	0	0	0	0	0	0.1
47 - 47.99	0	0	0.31	0.38	0.08	0	0.23	0	0	0
48 - 48.99	0	0	0.2	0.4	0	0	0.3	0	0	0.1
49 - 49.99	0	0	0.12	0.5	0	0	0.25	0	0.12	0
50 - 50.99	0	0	0.1	0.8	0	0	0.1	0	0	0
51 - 51.99	0	0	0	0.64	0.09	0.09	0.09	0	0.09	0
52 - 52.99	0	0	0.25	0.25	0.5	0	0	0	0	0
53 - 53.99	0	0	0	0.25	0.25	0.25	0.25	0	0	0
54 - 54.99	0	0	0	0.5	0	0	0	0.5	0	0
55 - 55.99	0	0	0	0	0	0.25	0.75	0	0	0
56 - 56.99	0	0	0	0	0	1	0	0	0	0
58 - 58.99	0	0	0	0	0	0	1	0	0	0
61 - 61.99	0	0	0	0	0	0	1	0	0	0
64 - 64.99	0	0	0	0	0	0	0	0	1	0

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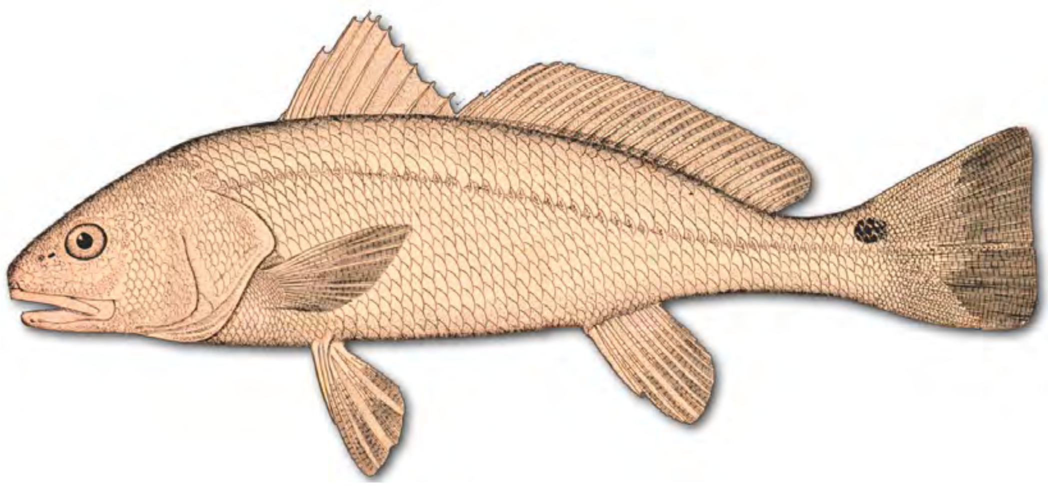
Chapter 5

RED DRUM *Sciaenops ocellatus*



5.1 INTRODUCTION

We aged a total of 31 red drum, *Sciaenops ocellatus*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2015. Red drum ages ranged from 1 to 6 years old with an average age of 3.4, a standard deviation of 1, and a standard error of 0.18. Four age classes (1, 3 to 4, and 6) were represented, comprising fish of the 2009, 2011 to 2012, and 2014 year-classes. The sample was dominated by fish from the year-classes of 2011 and 2012 with 48.4% and 38.7%, respectively.

5.2 METHODS

5.2.1 Handling of collections

Sagittal otoliths, hereafter, referred to as "otoliths", were received by the Age and Growth Laboratory in labeled coin envelopes, and were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

5.2.2 Preparation

Otoliths were processed for age determination following the methods described in Ross et al. (1993) and Jones and Wells (1998) for red drum. The left or right sagittal otolith was randomly selected and attached, distal side down, to a glass slide with Crystalbond™ 509 adhesive. The otoliths were viewed by eye, and when necessary, under a stereo microscope to identify the location of the core, and the position of the core marked using a pencil across the otolith surface. At least one transverse cross-section (hereafter "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet™ low-speed saw equipped with two, three inch diameter, Norton Diamond Grinding Wheels, separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The position of the marked core fell within the 0.5 mm space between the blades, such that the core was included in the removed thin-section. Otolith thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the sections.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing red drum.

5.2.3 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, red drum annulus formation occurs between March and June (Ross et al. 1995 and modified by CQFE). A red drum captured between January 1 and June 30, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of June 30, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 5.1). Each reader aged all of the otolith samples.

All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the

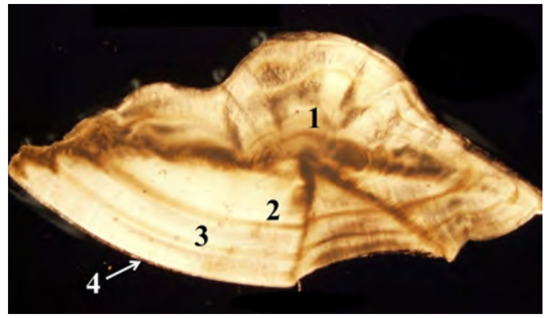


Figure 5.1: Otolith thin-section of a 4 year-old red drum with the last annulus on the edge of the thin-section

fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age red drum using their otolith thin-sections.

5.2.4 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (*CV*) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. When the sample size for the current year was smaller than 50, the entire sample was read by each reader for the second time to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

5.3 RESULTS

5.3.1 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 100% , and there was no significant difference between the first and second readings for Reader 2 with an agreement of 100%. There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 100% (Figure 5.2).

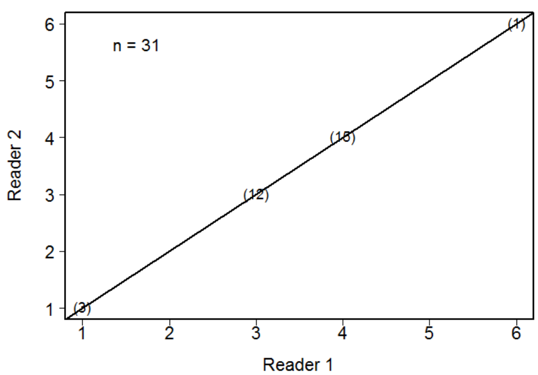


Figure 5.2: Between-reader comparison of otolith age estimates for red drum collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

There was no time-series bias for both readers. Reader 1 had an agreement of 98% with ages of fish aged in 2000 with a *CV* of 0.4% (test of symmetry: $\chi^2 = 1$, $df = 1$, $P = 0.3173$). Reader 2 had an agreement of 98% with a *CV* of 0.1% (test of symmetry: $\chi^2 = 1$, $df = 1$, $P = 0.3173$).

5.3.2 Year class

Of the 31 fish aged with otoliths, 4 age classes (1, 3 to 4, and 6) were represented (Table 5.1). The average age was 3.4 years, and the standard deviation and standard error were 1 and 0.18, respectively. Year-class data show that the fishery was comprised of 4 year-classes: fish from the 2009, 2011 to 2012, and 2014 year-classes, with fish primarily from the year classes of 2011 and 2012 with 48.4% and 38.7%, respectively. The ratio of males to females was 1:1.07 in the sample collected (Figure 5.3).

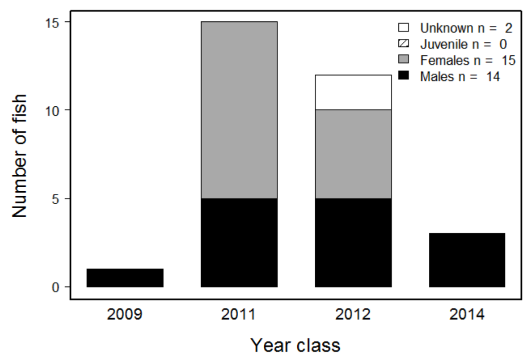


Figure 5.3: Year-class frequency distribution for red drum collected for ageing in 2015. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

5.3.3 Age-length key

We developed an age-length-key (Table 5.2) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

5.4 REFERENCES

Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.

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Ross, J., Stevens, T., Vaughan, D. 1995. Age, Growth, Mortality, and Reproductive Biology of Red Drums in North Carolina Waters. Trans. Amer. Fish. Soc. 124:37-54.

Table 5.1: The number of red drum assigned to each total length (inch)-at-age category for 31 fish sampled for otolith age determination in Virginia during 2015.

	Age				
Interval	1	3	4	6	Totals
18 - 18.99	1	0	0	0	1
19 - 19.99	2	0	0	0	2
21 - 21.99	0	2	0	0	2
22 - 22.99	0	2	0	0	2
23 - 23.99	0	3	1	0	4
24 - 24.99	0	3	2	0	5
25 - 25.99	0	2	1	0	3
26 - 26.99	0	0	4	0	4
27 - 27.99	0	0	3	0	3
28 - 28.99	0	0	3	0	3
30 - 30.99	0	0	1	0	1
36 - 36.99	0	0	0	1	1
Totals	3	12	15	1	31

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Table 5.2: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for red drum sampled for age determination in Virginia during 2015.

	Age			
Interval	1	3	4	6
18 - 18.99	1	0	0	0
19 - 19.99	1	0	0	0
21 - 21.99	0	1	0	0
22 - 22.99	0	1	0	0
23 - 23.99	0	0.75	0.25	0
24 - 24.99	0	0.6	0.4	0
25 - 25.99	0	0.67	0.33	0
26 - 26.99	0	0	1	0
27 - 27.99	0	0	1	0
28 - 28.99	0	0	1	0
30 - 30.99	0	0	1	0
36 - 36.99	0	0	0	1

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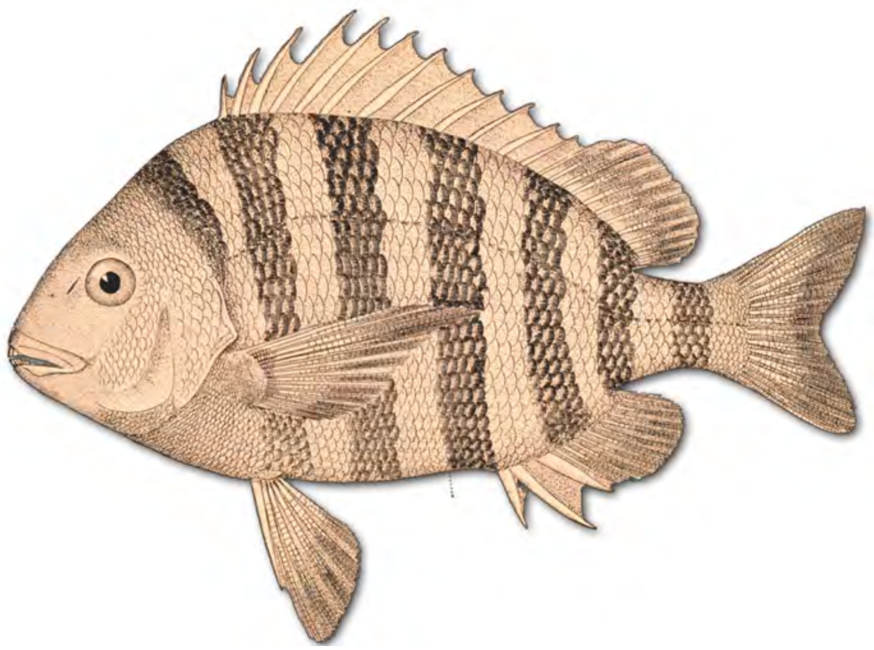
Chapter 6

SHEEPSHEAD *Archosargus probatocephalus*



6.1 INTRODUCTION

We aged a total of 119 sheepshead, *Archosargus probatocephalus*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2015. Sheepshead ages ranged from 2 to 32 years old with an average age of 9.1, a standard deviation of 6.6, and a standard error of 0.61. Twenty age classes (2 to 4, 6 to 10, 12 to 14, 17 to 19, 21, 23 to 25, 27, and 32) were represented, comprising fish of the 1983, 1988, 1990 to 1992, 1994, 1996 to 1998, 2001 to 2003, 2005 to 2009, and 2011 to 2013 year-classes. The sample was dominated by fish from the year-class of 2011 with 48.7%.

6.2 METHODS

6.2.1 Handling of collections

Sagittal otoliths, hereafter, referred to as "otoliths", were received by the Age and Growth Laboratory in labeled coin envelopes,and were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

6.2.2 Preparation

Otoliths were processed for age determination following the methods described in Ballenger et al. (2011). The left or right otolith was randomly selected and embedded, distal side down, in epoxy resin and allowed to harden overnight. The otoliths were viewed by eye, and when necessary, under a stereo microscope to identify the location of the core, and the position of the core marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet^{TM} low-speed saw equipped with two, three inch diameter, Norton Diamond Grinding Wheels, separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The position of the marked core fell within the 0.5 mm space between the blades, such that the core was included in the removed thin section. Otolith thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing sheepshead.

6.2.3 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, sheepshead otolith annulus formation occurs between May and June (Ballenger 2011). A sheepshead captured between January 1 and July 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 6.1). Each reader aged all of the otolith samples. All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When

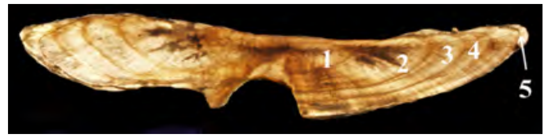


Figure 6.1: Otolith thin-section of a 5 year-old sheepshead

the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age sheepshead using their otolith thin-sections.

6.2.4 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

6.3 RESULTS

6.3.1 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 94% and a *CV* of 0.5% (test of symmetry: $\chi^2 = 3$, $df = 3$, $P = 0.3916$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 92% and a *CV* of 0.5% (test of symmetry: $\chi^2 = 4$, $df = 3$, $P = 0.2615$). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 84.03% and a *CV* of 1.1% (test of symmetry: $\chi^2 = 13.67$, $df = 11$, $P = 0.252$) (Figure 6.2).

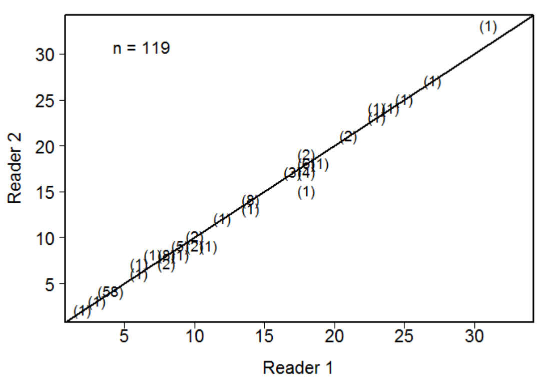


Figure 6.2: Between-reader comparison of otolith age estimates for sheepshead collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

There was no time-series bias for both readers. Reader 1 had an agreement of 92% with ages of fish aged in 2008 with a *CV* of 0.3% (test of symmetry: $\chi^2 = 2$, $df = 3$, $P = 0.5724$). Reader 2 had an agreement of 100% .

6.3.2 Year class

Of the 119 fish aged with otoliths, 20 age classes (2 to 4, 6 to 10, 12 to 14, 17 to 19, 21, 23 to 25, 27, and 32) were represented (Table 6.1). The average age was 9.1 years, and the standard deviation and standard error were 6.6 and 0.61, respectively. Year-class data show that the fishery was comprised of 20 year-classes: fish from the 1983, 1988, 1990 to 1992, 1994, 1996 to 1998, 2001 to 2003, 2005 to 2009, and 2011 to 2013 year-classes, with fish primarily from the year class of 2011 with 48.7%. The ratio of males to females was 1:1.39 in the sample collected (Figure 6.3).

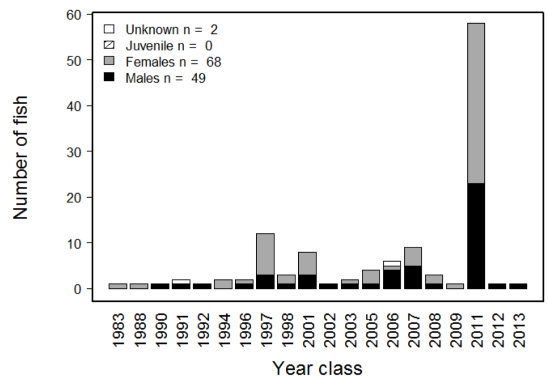


Figure 6.3: Year-class frequency distribution for sheepshead collected for ageing in 2015. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

6.3.3 Age-length key

We developed an age-length-key (Table 6.2) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

6.4 REFERENCES

Campana, S.E., M.C. Annand, and J.I. McMillan. 1995. Graphical and statistical methods for determining the consistency of age determinations. Trans. Am. Fish. Soc. 124:131-138.

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Table 6.1: The number of sheepshead assigned to each total length (inch)-at-age category for 119 fish sampled for otolith age determination in Virginia during 2015.

	Age																			
Interval	2	3	4	6	7	8	9	10	12	13	14	17	18	19	21	23	24	25	27	32	Totals
14 - 14.99	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
15 - 15.99	0	0	4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	4
16 - 16.99	0	1	8	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	10
17 - 17.99	0	0	21	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	21
18 - 18.99	0	0	20	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	21
19 - 19.99	0	0	2	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	3
20 - 20.99	0	0	3	0	1	3	1	0	0	0	0	0	0	0	0	0	0	0	0	0	8
21 - 21.99	0	0	0	0	2	3	2	2	0	0	1	3	1	0	0	0	1	0	0	0	15
22 - 22.99	0	0	0	0	0	1	2	1	2	0	1	0	4	0	1	0	1	0	0	0	13
23 - 23.99	0	0	0	0	0	0	1	1	0	0	4	0	4	1	0	0	0	1	0	1	13
24 - 24.99	0	0	0	0	0	0	0	0	0	1	1	0	2	0	0	1	0	0	0	0	5
25 - 25.99	0	0	0	0	0	0	0	0	0	0	1	0	1	1	1	0	0	0	1	0	5
Totals	1	1	58	1	3	9	6	4	2	1	8	3	12	2	2	1	2	1	1	1	119

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Table 6.2: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for sheepshead sampled for age determination in Virginia during 2015.

	Age																		
Interval	2	3	4	6	7	8	9	10	12	13	14	17	18	19	21	23	24	25	27	32
14 - 14.99	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
15 - 15.99	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
16 - 16.99	0	0.1	0.8	0.1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
17 - 17.99	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
18 - 18.99	0	0	0.95	0	0	0.05	0	0	0	0	0	0	0	0	0	0	0	0	0	0
19 - 19.99	0	0	0.67	0	0	0.33	0	0	0	0	0	0	0	0	0	0	0	0	0	0
20 - 20.99	0	0	0.38	0	0.12	0.38	0.12	0	0	0	0	0	0	0	0	0	0	0	0	0
21 - 21.99	0	0	0	0	0.13	0.2	0.13	0.13	0	0	0.07	0.2	0.07	0	0	0	0.07	0	0	0
22 - 22.99	0	0	0	0	0	0.08	0.15	0.08	0.15	0	0.08	0	0.31	0	0.08	0	0.08	0	0	0
23 - 23.99	0	0	0	0	0	0	0.08	0.08	0	0	0.31	0	0.31	0.08	0	0	0	0.08	0	0.08
24 - 24.99	0	0	0	0	0	0	0	0	0	0.2	0.2	0	0.4	0	0	0.2	0	0	0	0
25 - 25.99	0	0	0	0	0	0	0	0	0	0	0.2	0	0.2	0.2	0.2	0	0	0	0.2	0

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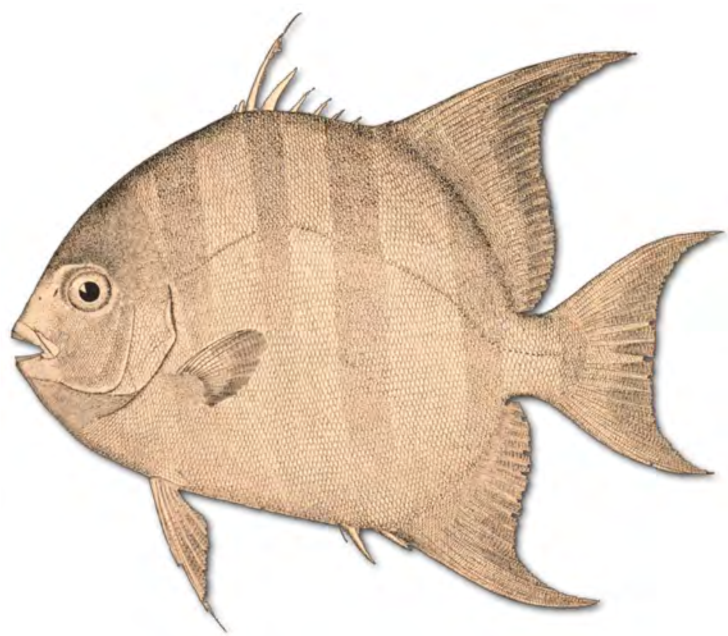
Chapter 7

ATLANTIC SPADEFISH
Chaetodipterus faber



7.1 INTRODUCTION

We aged a total of 135 spadefish, *Chaetodipterus faber*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2015. Spadefish ages ranged from 1 to 8 years old with an average age of 3.6, a standard deviation of 1.6, and a standard error of 0.14. Eight age classes (1 to 8) were represented, comprising fish of the 2007 to 2014 year-classes. The sample was dominated by fish from the year-classes of 2012 and 2010 with 34.1% and 27.4%, respectively.

7.2 METHODS

7.2.1 Sample size for ageing

We estimated sample size for ageing spadefish in 2015 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 CV^2 + B_a/L} \qquad (7.1)$$

where A is the sample size for ageing spadefish in 2015; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of spadefish used by VMRC to estimate length distribution of the catches from 2009 to 2013. θ_a, V_a, B_a, and CV were calculated using pooled age-length data of spadefish collected from 2009 to 2013 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (7.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2009 to 2013 catch. A_l is number of fish to be aged for length interval l in 2015.

7.2.2 Handling of collections

Otoliths were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

7.2.3 Preparation

We used our thin-section and bake technique to process spadefish sagittal otoliths (hereafter, referred to as "otoliths") for age determination. Otolith preparation began by randomly selecting either the right or left otolith. Each whole otolith was placed in a ceramic "Coors" spot plate well and baked in a Thermolyne 1400 furnace at 400 °C. Baking time was dependent on the otolith's size and gauged by color, with a light caramel color desired. Once a suitable color was achieved the baked otolith was embedded in epoxy resin with its distal surface orientated downwards and allowed to harden overnight. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet^{TM} low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The otolith was positioned so the blades straddled each side of the otolith focus. It was crucial that this cut be perpendicular to the long axis of the otolith. Failure to do so resulted in broadening and distored winter growth zones. A proper cut resulted in annuli that were clearly defined and delineated. Once cut, thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing Atlantic spadefish.

7.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, spadefish otolith annulus formation occurs between December and July (Hayse 1989 and modified by CQFE). A spadefish captured between January 1 and July 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of July 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 7.1). Each reader aged all of the otolith samples. All samples were aged in chronological order, based on collection date, without knowledge of

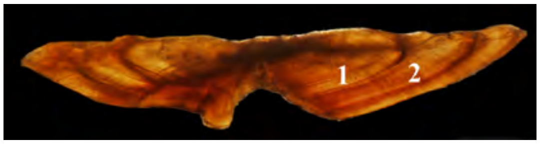


Figure 7.1: Otolith thin-section of a 2 year-old spadefish

previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age Atlantic spadefish using their otolith thin-sections.

7.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

7.3 RESULTS

7.3.1 Sample size

We estimated a sample size of 279 spadefish in 2015, ranging in length interval from 4 to 25 inches (Table 7.1). This sample size provided a range in (CV) for age composition approximately from the smallest (CV) of 6% for age 2 to the largest (CV) of 20% for age 1. In 2015, we spadefishaged all 135 collected by VMRC. We fell short in our over-all collections for this optimal length-class sampling estimate by 150 fish. However, we were short of many fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would definitely be influenced significantly. Therefore, precaution should be used when developing ALK using these age data.

7.3.2 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 94% and a CV of 0.9% (test of symmetry: $\chi^2 = 1$, $df = 2$, $P = 0.6065$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 86% and a CV of 2.2% (test of symmetry: $\chi^2 = 3$, $df = 3$, $P = 0.3916$). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 86.67% and a CV of 2.1% (test of symmetry: $\chi^2 = 8.4$, $df = 6$, $P = 0.2102$) (Figure 7.2).

There was no time-series bias for both readers. Reader 1 had an agreement of 86% with ages of fish aged in 2003 with a CV of 1.9% (test of symmetry: $\chi^2 = 5$, $df = 6$, $P = 0.5438$). Reader 2 had an agreement of

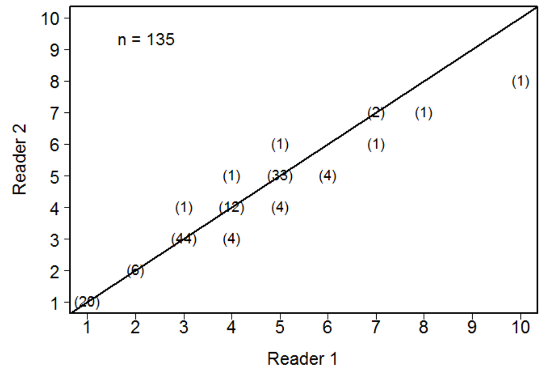


Figure 7.2: Between-reader comparison of otolith age estimates for spadefish collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

94% with a *CV* of 1.2% (test of symmetry: $\chi^2 = 3$, $df = 3$, $P = 0.3916$).

7.3.3 Year class

Of the 135 fish aged with otoliths, 8 age classes (1 to 8) were represented (Table 7.2). The average age was 3.6 years, and the standard deviation and standard error were 1.6 and 0.14, respectively. Year-class data show that the fishery was comprised of 8 year-classes: fish from the 2007 to 2014 year-classes, with fish primarily from the year classes of 2012 and 2010 with 34.1% and 27.4%, respectively. The ratio of males to females was 1:1.29 in the sample collected (Figure 7.3).

7.3.4 Age-length key

We developed an age-length-key (Table 7.3) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

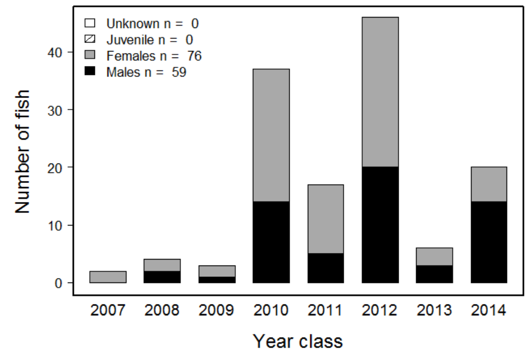


Figure 7.3: Year-class frequency distribution for spadefish collected for ageing in 2015. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

7.4 REFERENCES

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Table 7.1: Number of Atlantic spadefish collected and aged in each 1-inch length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
4 - 4.99	5	0	0	5
5 - 5.99	7	5	5	2
6 - 6.99	37	18	18	19
7 - 7.99	43	7	7	36
8 - 8.99	30	10	10	20
9 - 9.99	22	9	9	13
10 - 10.99	15	7	7	8
11 - 11.99	11	9	9	2
12 - 12.99	15	7	7	8
13 - 13.99	16	8	8	8
14 - 14.99	12	6	6	6
15 - 15.99	12	10	10	2
16 - 16.99	10	14	14	0
17 - 17.99	13	12	12	1
18 - 18.99	9	11	11	0
19 - 19.99	7	2	2	5
20 - 20.99	5	0	0	5
21 - 21.99	5	0	0	5
25 - 25.99	5	0	0	5
Totals	279	135	135	150

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Table 7.2: The number of Atlantic spadefish assigned to each total length-at-age category for 135 fish sampled for otolith age determination in Virginia during 2015.

	Age								
Interval	1	2	3	4	5	6	7	8	Totals
5 - 5.99	5	0	0	0	0	0	0	0	5
6 - 6.99	12	0	6	0	0	0	0	0	18
7 - 7.99	3	1	3	0	0	0	0	0	7
8 - 8.99	0	2	8	0	0	0	0	0	10
9 - 9.99	0	2	7	0	0	0	0	0	9
10 - 10.99	0	1	6	0	0	0	0	0	7
11 - 11.99	0	0	8	0	1	0	0	0	9
12 - 12.99	0	0	4	2	1	0	0	0	7
13 - 13.99	0	0	4	2	2	0	0	0	8
14 - 14.99	0	0	0	3	3	0	0	0	6
15 - 15.99	0	0	0	3	7	0	0	0	10
16 - 16.99	0	0	0	2	10	1	1	0	14
17 - 17.99	0	0	0	4	7	0	0	1	12
18 - 18.99	0	0	0	1	6	2	1	1	11
19 - 19.99	0	0	0	0	0	0	2	0	2
Totals	20	6	46	17	37	3	4	2	135

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Table 7.3: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for Atlantic spadefish sampled for age determination in Virginia during 2015.

	Age							
Interval	1	2	3	4	5	6	7	8
5 - 5.99	1	0	0	0	0	0	0	0
6 - 6.99	0.67	0	0.33	0	0	0	0	0
7 - 7.99	0.43	0.14	0.43	0	0	0	0	0
8 - 8.99	0	0.2	0.8	0	0	0	0	0
9 - 9.99	0	0.22	0.78	0	0	0	0	0
10 - 10.99	0	0.14	0.86	0	0	0	0	0
11 - 11.99	0	0	0.89	0	0.11	0	0	0
12 - 12.99	0	0	0.57	0.29	0.14	0	0	0
13 - 13.99	0	0	0.5	0.25	0.25	0	0	0
14 - 14.99	0	0	0	0.5	0.5	0	0	0
15 - 15.99	0	0	0	0.3	0.7	0	0	0
16 - 16.99	0	0	0	0.14	0.71	0.07	0.07	0
17 - 17.99	0	0	0	0.33	0.58	0	0	0.08
18 - 18.99	0	0	0	0.09	0.55	0.18	0.09	0.09
19 - 19.99	0	0	0	0	0	0	1	0

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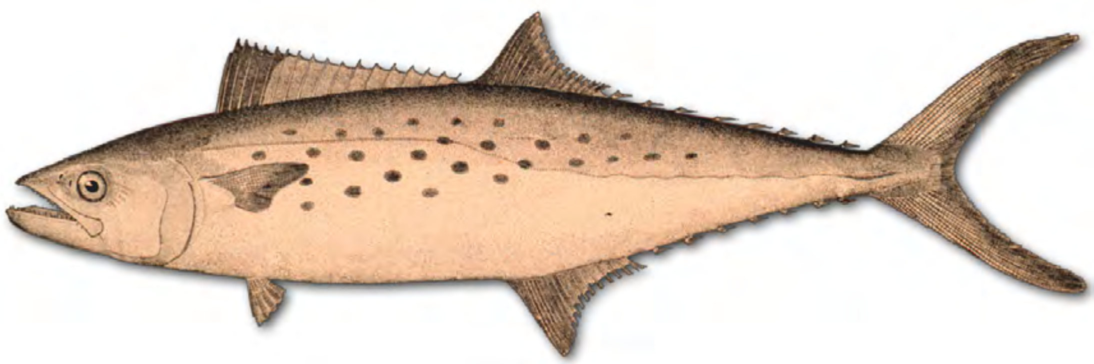
Chapter 8

SPANISH MACKEREL
Scomberomorous maculatus



8.1 INTRODUCTION

We aged a total of 231 Spanish mackerel, *Scomberomorous maculatus*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2015. Spanish mackerel ages ranged from 0 to 8 years old with an average age of 1.4, a standard deviation of 1, and a standard error of 0.07. Seven age classes (0 to 5, and 8) were represented, comprising fish of the 2007, and 2010 to 2015 year-classes. The sample was dominated by fish from the year-class of 2014 with 60.2%.

8.2 METHODS

8.2.1 Sample size for ageing

We estimated sample size for ageing Spanish mackerel in 2015 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 CV^2 + B_a/L} \qquad (8.1)$$

where A is the sample size for ageing Spanish mackerel in 2015; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of Spanish mackerel used by VMRC to estimate length distribution of the catches from 2009 to 2013. θ_a, V_a, B_a, and CV were calculated using pooled age-length data of Spanish mackerel collected from 2009 to 2013 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (8.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2009 to 2013 catch. A_l is number of fish to be aged for length interval l in 2015.

8.2.2 Handling of collections

Otoliths were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

8.2.3 Preparation

Sagittal otoliths, hereafter, referred to as "otolith", were processed for age determination. The left or right otolith was randomly selected and embedded, distal side down, in epoxy resin and allowed to harden overnight. The otoliths were viewed by eye, and when necessary, under a stereo microscope to identify the location of the core, and the position of the core marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet^{TM} low-speed saw equipped with two, three inch diameter, Norton Diamond Grinding Wheels, separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The position of the marked core fell within the 0.5 mm space between the blades, such that the core was included in the removed thin section. Otolith thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing Spanish mackerel.

8.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, Spanish mackerel annulus formation occurs between May and June (Schmidt et al. 1993). A Spanish mackerel captured between January 1 and June 30, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 8.1). Each reader aged all of the otolith samples. All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When

Figure 8.1: Otolith thin-section of a 3 year-old Spanish mackerel with the last annulus on the edge of the thin-section

the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age Spanish mackerel using their otolith thin-sections.

8.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

8.3 RESULTS

8.3.1 Sample size

We estimated a sample size of 266 Spanish mackerel in 2015, ranging in length interval from 10 to 30 inches (Table 8.1). This sample size provided a range in (CV) for age composition approximately from the smallest (CV) of 4% for age 1 to the largest (CV) of 18% for age 3. In 2015, we randomly selected and aged 231 fish from 327 Spanish mackerel collected by VMRC. We fell short in our over-all collections for this optimal length-class sampling estimate by 44 fish. However, we were short of no fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

8.3.2 Reading precision

Reader 1 had moderate self-precision and Reader 2 had high self-precision. Specifically, there was a difference between the first and second readings for Reader 1 with an agreement of 80% and a CV of 8.1% (test of symmetry: $\chi^2 = 10$, $df = 3$, $P = 0.0186$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 100% . There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 95.24% and a CV of 1.7% (test of symmetry: $\chi^2 = 2.33$, $df = 4$, $P = 0.6747$) (Figure 8.2).

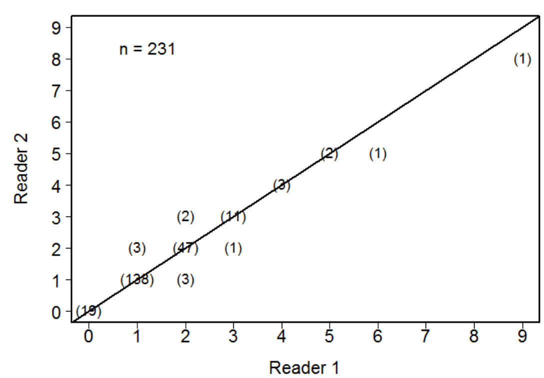


Figure 8.2: Between-reader comparison of otolith age estimates for Spanish mackerel collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

There was no time-series bias for both readers. Reader 1 had an agreement of 96% with fish aged in 2003 with a *CV* of 1.3% (test of symmetry: $\chi^2 = 2$, $df = 2$, $P = 0.3679$). Reader 2 had an agreement of 90% with a *CV* of 3.4% (test of symmetry: $\chi^2 = 5$, $df = 3$, $P = 0.1718$).

8.3.3 Year class

Of the 231 fish aged with otoliths, 7 age classes (0 to 5, and 8) were represented (Table 8.2). The average age was 1.4 years, and the standard deviation and standard error were 1 and 0.07, respectively. Year-class data show that the fishery was comprised of 7 year-classes: fish from the 2007, and 2010 to 2015 year-classes, with fish primarily from the year class of 2014 with 60.2%. The ratio of males to females was 1:1.86 in the sample collected (Figure 8.3).

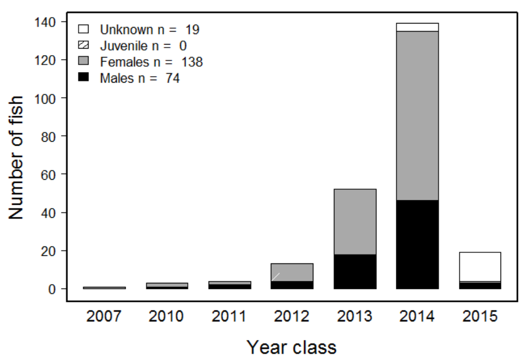


Figure 8.3: Year-class frequency distribution for Spanish mackerel collected for ageing in 2015. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

8.3.4 Age-length key

We developed an age-length-key (Table 8.3) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

8.4 REFERENCES

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Schmidt, D., Collins, M., Wyanski, D. 1993. Age, growth, maturity, and spawning of Spanish mackerel from the Atlantic coast of the southeastern United States. Fishery Bulletin 91s.

Table 8.1: Number of Spanish mackerel collected and aged in each 1-inch length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
10 - 10.99	5	0	0	5
11 - 11.99	5	0	0	5
12 - 12.99	5	0	0	5
13 - 13.99	5	1	1	4
14 - 14.99	19	30	23	0
15 - 15.99	38	61	38	0
16 - 16.99	41	64	41	0
17 - 17.99	36	50	36	0
18 - 18.99	21	35	22	0
19 - 19.99	17	27	18	0
20 - 20.99	13	19	14	0
21 - 21.99	13	15	14	0
22 - 22.99	8	9	8	0
23 - 23.99	5	6	6	0
24 - 24.99	5	5	5	0
25 - 25.99	5	1	1	4
26 - 26.99	5	2	2	3
27 - 27.99	5	1	1	4
28 - 28.99	5	0	0	5
29 - 29.99	5	1	1	4
30 - 30.99	5	0	0	5
Totals	266	327	231	44

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Table 8.2: The number of Spanish mackerel assigned to each total length-at-age category for 231 fish sampled for otolith age determination in Virginia during 2015.

	Age							
Interval	0	1	2	3	4	5	8	Totals
13 - 13.99	0	1	0	0	0	0	0	1
14 - 14.99	6	17	0	0	0	0	0	23
15 - 15.99	6	31	1	0	0	0	0	38
16 - 16.99	7	34	0	0	0	0	0	41
17 - 17.99	0	32	4	0	0	0	0	36
18 - 18.99	0	15	7	0	0	0	0	22
19 - 19.99	0	5	11	2	0	0	0	18
20 - 20.99	0	3	10	1	0	0	0	14
21 - 21.99	0	1	9	1	2	1	0	14
22 - 22.99	0	0	4	4	0	0	0	8
23 - 23.99	0	0	3	1	1	1	0	6
24 - 24.99	0	0	2	3	0	0	0	5
25 - 25.99	0	0	1	0	0	0	0	1
26 - 26.99	0	0	0	1	1	0	0	2
27 - 27.99	0	0	0	0	0	1	0	1
29 - 29.99	0	0	0	0	0	0	1	1
Totals	19	139	52	13	4	3	1	231

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Table 8.3: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for Spanish mackerel sampled for age determination in Virginia during 2015.

	Age						
Interval	0	1	2	3	4	5	8
13 - 13.99	0	1	0	0	0	0	0
14 - 14.99	0.26	0.74	0	0	0	0	0
15 - 15.99	0.16	0.82	0.03	0	0	0	0
16 - 16.99	0.17	0.83	0	0	0	0	0
17 - 17.99	0	0.89	0.11	0	0	0	0
18 - 18.99	0	0.68	0.32	0	0	0	0
19 - 19.99	0	0.28	0.61	0.11	0	0	0
20 - 20.99	0	0.21	0.71	0.07	0	0	0
21 - 21.99	0	0.07	0.64	0.07	0.14	0.07	0
22 - 22.99	0	0	0.5	0.5	0	0	0
23 - 23.99	0	0	0.5	0.17	0.17	0.17	0
24 - 24.99	0	0	0.4	0.6	0	0	0
25 - 25.99	0	0	1	0	0	0	0
26 - 26.99	0	0	0	0.5	0.5	0	0
27 - 27.99	0	0	0	0	0	1	0
29 - 29.99	0	0	0	0	0	0	1

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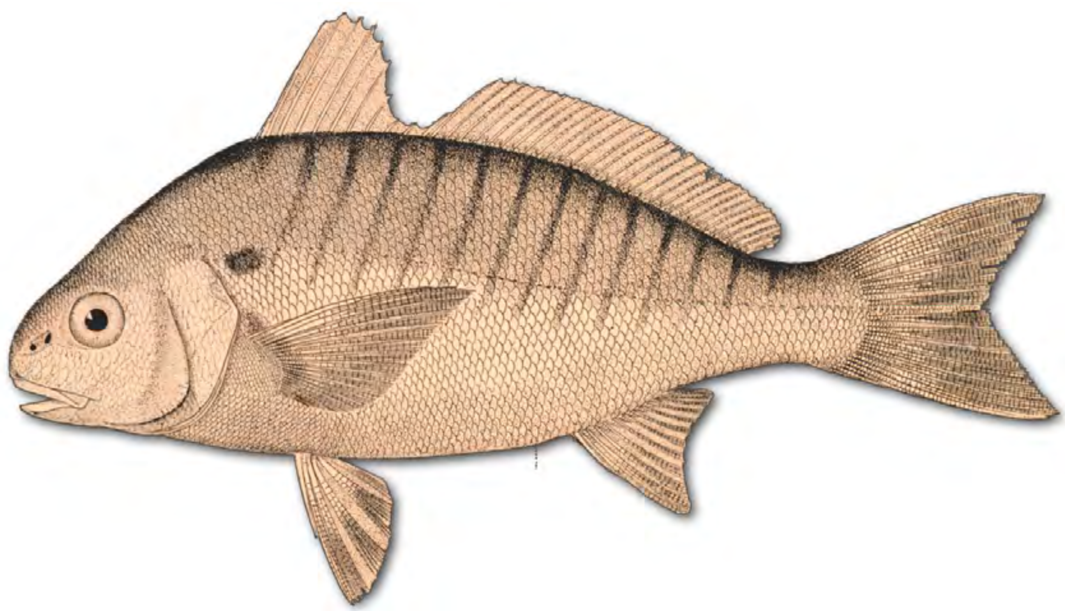
Chapter 9

SPOT *Leiostomus xanthurus*



9.1 INTRODUCTION

We aged a total of 201 spot, *Leiostomus xanthurus*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2015. spot ages ranged from 0 to 4 years old with an average age of 1.3, a standard deviation of 0.7, and a standard error of 0.05. Five age classes (0 to 4) were represented, comprising fish of the 2011 to 2015 year-classes. The sample was dominated by fish from the year-class of 2014 with 71.1%.

9.2 METHODS

9.2.1 Sample size for ageing

We estimated sample size for ageing spot in 2015 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 CV^2 + B_a/L} \tag{9.1}$$

where A is the sample size for ageing spot in 2015; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of spot used by VMRC to estimate length distribution of the catches from 2009 to 2013. θ_a, V_a, B_a, and CV were calculated using pooled age-length data of spot collected from 2009 to 2013 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (1.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by ageing an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2009 to 2013 catch. A_l is number of fish to be aged for length interval l in 2015.

9.2.2 Handling of collections

Otoliths were received by the Age & and Growth Laboratory in labeled coin envelopes, were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

9.2.3 Preparation

Sagittal otoliths, hereafter, referred to as "otoliths", were processed for age determination following the methods described in Barbieri et al. (1994) with a few modifications. The left or right otolith was randomly selected and embedded (distal side down) in epoxy resin and allowed to harden overnight. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet^{TM} low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). Thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-sections.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing spot.

9.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, spot otolith annulus formation occurs between May and July (Piner and Jones 2004). A spot captured between January 1 and May 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 9.1). Each reader aged all of the otolith samples. Due to discrepancy on identification of the first annulus of spot among Atlantic states, Atlantic States Marine Fisheries Commission (ASMFC) has decided not to count the smallest annulus at the center of the thin-section as the first annulus. Following ASMFC's instruction, we didn't count the smallest annulus at the center as the first annulus in 2015.

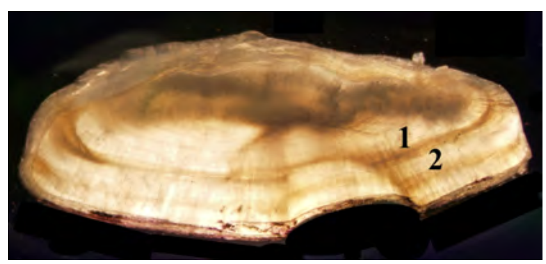


Figure 9.1: Otolith thin-section of a 2 year-old spot

All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age spot using their otolith thin-sections.

9.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

9.3 RESULTS

9.3.1 Sample size

We estimated a sample size of 203 spot in 2015, ranging in length interval from 4 to 12 inches (Table 9.1). This sample size provided a range in (CV) for age composition approximately from the smallest (CV) of 5% for age 1 to the largest (CV) of 11% for age 2. In 2015, we randomly selected and aged 201 fish from 263 spot collected by VMRC. We fell short in our over-all collections for this optimal length-class sampling estimate by 11 fish. However, we were short of no fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

9.3.2 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 94% and a CV of 4.3% (test of symmetry: $\chi^2 = 3$, $df = 3$, $P = 0.3916$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 98% and a CV of 0.9% (test of symmetry: $\chi^2 = 1$, $df = 1$, $P = 0.3173$). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 94.53% and a CV of 2.1% (test of symmetry: $\chi^2 = 7.57$, $df = 3$, $P = 0.0558$) (Figure 9.2).

There was no time-series bias for both readers. Reader 1 had an agreement of 98% with ages of fish aged in 2003 with a CV of 0.9% (test of symmetry: $\chi^2 = 1$, $df = 1$, $P = 0.3173$). Reader 2 had an agreement of 98% with a CV of 0.9% (test of symmetry: $\chi^2 = 1$, $df = 1$, $P = 0.3173$).

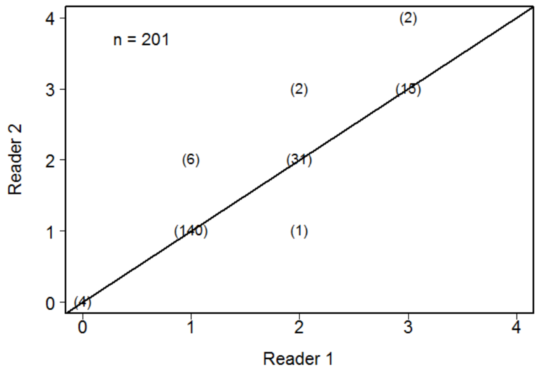


Figure 9.2: Between-reader comparison of otolith age estimates for spot collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

9.3.3 Year class

Of the 201 fish aged with otoliths, 5 age classes (0 to 4) were represented (Table 9.2). The average age was 1.3 years, and the standard deviation and standard error were 0.7 and 0.05, respectively. Year-class data show that the fishery was comprised of 5 year-classes: fish from the 2011 to 2015 year-classes, with fish primarily from the year class of 2014 with 71.1%. The ratio of males to females was 1:4.74 in the sample collected (Figure 9.3).

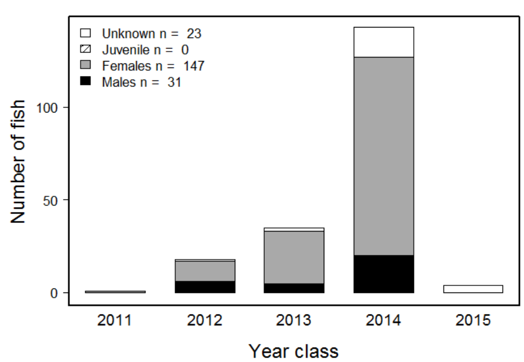


Figure 9.3: Year-class frequency distribution for spot collected for ageing in 2015. Distribution is broken down by sex. 'Unknown' is for gonads that were not available for examination or were not examined for sex during sampling.

9.3.4 Age-length key

We developed an age-length-key (Table 9.3) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

9.4 REFERENCES

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Table 9.1: Number of spot collected and aged in each 1-inch length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
4 - 4.99	5	2	2	3
5 - 5.99	5	12	6	0
6 - 6.99	5	15	6	0
7 - 7.99	23	40	24	0
8 - 8.99	44	50	44	0
9 - 9.99	61	78	62	0
10 - 10.99	46	60	51	0
11 - 11.99	9	6	6	3
12 - 12.99	5	0	0	5
Totals	203	263	201	11

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Table 9.2: The number of spot assigned to each total length-at-age category for 201 fish sampled for otolith age determination in Virginia during 2015.

	Age					
Interval	0	1	2	3	4	Totals
4 - 4.99	2	0	0	0	0	2
5 - 5.99	2	3	1	0	0	6
6 - 6.99	0	6	0	0	0	6
7 - 7.99	0	22	2	0	0	24
8 - 8.99	0	31	12	1	0	44
9 - 9.99	0	36	14	11	1	62
10 - 10.99	0	40	5	6	0	51
11 - 11.99	0	5	1	0	0	6
Totals	4	143	35	18	1	201

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Table 9.3: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for spot sampled for age determination in Virginia during 2015.

	Age				
Interval	0	1	2	3	4
4 - 4.99	1	0	0	0	0
5 - 5.99	0.33	0.5	0.17	0	0
6 - 6.99	0	1	0	0	0
7 - 7.99	0	0.92	0.08	0	0
8 - 8.99	0	0.7	0.27	0.02	0
9 - 9.99	0	0.58	0.23	0.18	0.02
10 - 10.99	0	0.78	0.1	0.12	0
11 - 11.99	0	0.83	0.17	0	0

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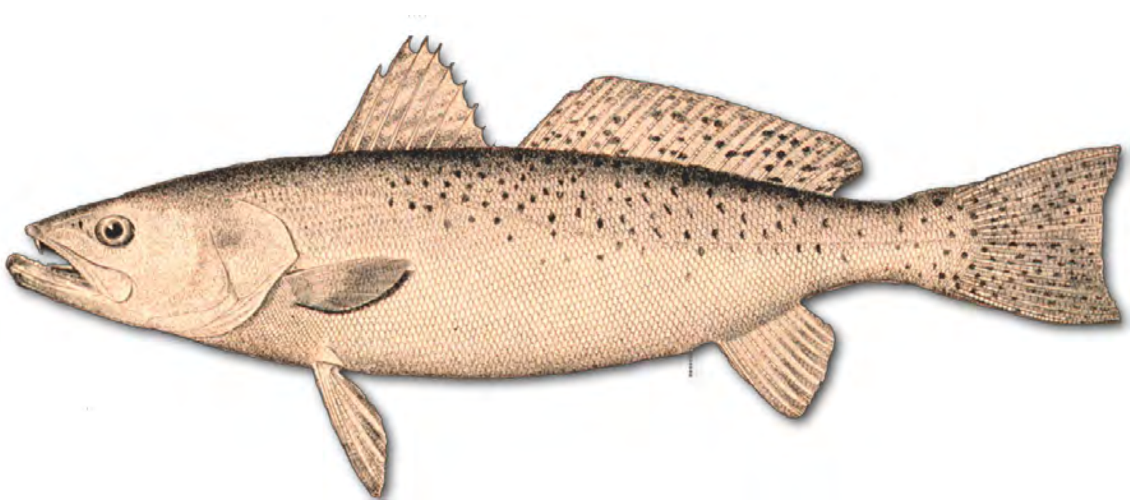
Chapter 10

SPOTTED SEATROUT *Cynoscion nebulosus*



10.1 INTRODUCTION

We aged a total of 308 spotted seatrout, *Cynoscion nebulosus*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2015. Spotted seatrout ages ranged from 0 to 10 years old with an average age of 2.5, a standard deviation of 2.1, and a standard error of 0.12. Ten age classes (0 to 8, and 10) were represented, comprising fish of the 2005, and 2007 to 2015 year-classes. The sample was dominated by fish from the year-classes of 2011, 2014, and 2015 with 27.6%, 26.9%, and 16.9%, respectively.

10.2 METHODS

10.2.1 Sample size for ageing

We estimated sample size for ageing spotted seatrout in 2015 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 CV^2 + B_a/L} \qquad (10.1)$$

where A is the sample size for ageing spotted seatrout in 2015; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of spotted seatrout used by VMRC to estimate length distribution of the catches from 2009 to 2013. θ_a, V_a, B_a, and CV were calculated using pooled age-length data of spotted seatrout collected from 2009 to 2013 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (10.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2009 to 2013 catch. A_l is number of fish to be aged for length interval l in 2015.

10.2.2 Handling of collections

Otoliths were received by the Age & Growth Laboratory in labeled coin envelopes. In the lab they were sorted by date of capture, their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

10.2.3 Preparation

Sagittal otoliths, hereafter, referred to as "otoliths", were processed for age determination. The left or right otolith was randomly selected and attached, distal side down, to a glass slide with clear Crystalbond™ 509 adhesive. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a pencil across the otolith surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet™ low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). Thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-sections.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing spotted seatrout.

10.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, spotted seatrout otolith annulus formation occurs between March and May (Ihde and Chittenden 2003). A spotted seatrout captured between January 1 and May 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of May 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 10.1). Each reader aged all of the otolith samples. All samples were aged in chronological order, based

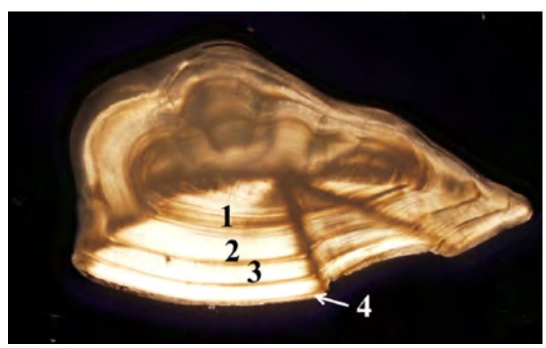


Figure 10.1: Otolith thin-section of a 4 year-old spotted seatrout with the last annulus on the edge of the thin-section

on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age spotted seatrout using their otolith thin-sections.

10.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

10.3 RESULTS

10.3.1 Sample size

We estimated a sample size of 342 spotted seatrout in 2015, ranging in length interval from 8 to 35 inches (Table 10.1). This sample size provided a range in (CV) for age composition approximately from the smallest (CV) of 5% for age 1 to the largest (CV) of 24% for age 4. In 2015, we aged 308 of 328 spotted seatrout (The rest of fish were either without otoliths or overcollected for certain length interval(s)) collected by VMRC. We fell short in our overall collections for this optimal length-class sampling estimate by 57 fish. However, we were short of some fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would possibly be influenced significantly.

10.3.2 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 98% and a CV of 0.6% (test of symmetry: $\chi^2 = 1$, $df = 1$, $P = 0.3173$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 100%. There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 100% (Figure 10.2).

There was no time-series bias for both readers. Reader 1 had an agreement of 100% with ages of fish aged in 2003. Reader 2 had an agreement of 100%.

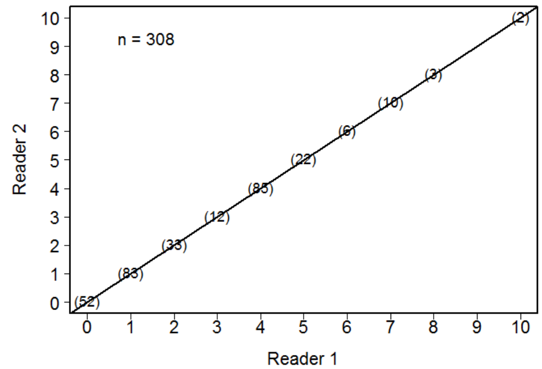


Figure 10.2: Between-reader comparison of otolith age estimates for spotted seatrout collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

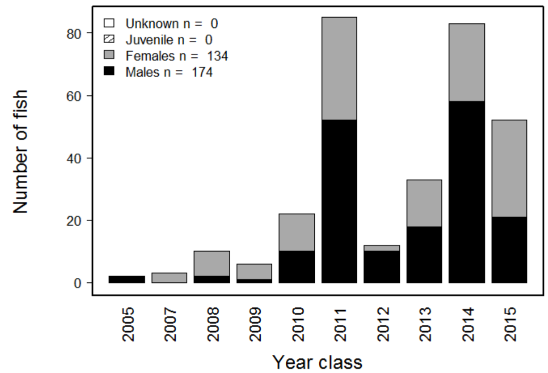


Figure 10.3: Year-class frequency distribution for spotted seatrout collected for ageing in 2015. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

10.3.3 Year class

Of the 308 fish aged with otoliths, 10 age classes (0 to 8, and 10) were represented (Table 10.2). The average age was 2.5 years, and the standard deviation and standard error were 2.1 and 0.12, respectively. Year-class data show that the fishery was comprised of 10 year-classes: fish from the 2005, and 2007 to 2015 year-classes, with fish primarily from the year classes of 2011, 2014, and 2015 with 27.6%, 26.9%, and 16.9%, respectively. The ratio of males to females was 1:0.77 in the sample collected (Figure 10.3).

10.3.4 Age-length key

We developed an age-length-key (Table 10.3) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

10.4 REFERENCES

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Quinn, T. J. II, and R. B. Deriso. 1999. Quantitative Fish Dynamics. Oxford Univeristy Press. New York.

R Core Team (2012). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL http://www.R-project.org.

Table 10.1: Number of spotted seatrout collected and aged in each 1-inch length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
8 - 8.99	5	0	0	5
9 - 9.99	5	0	0	5
10 - 10.99	5	0	0	5
11 - 11.99	8	1	1	7
12 - 12.99	21	29	29	0
13 - 13.99	17	19	18	0
14 - 14.99	15	16	16	0
15 - 15.99	23	24	24	0
16 - 16.99	31	31	31	0
17 - 17.99	33	31	31	2
18 - 18.99	27	27	27	0
19 - 19.99	27	13	13	14
20 - 20.99	22	25	25	0
21 - 21.99	11	16	12	0
22 - 22.99	12	13	12	0
23 - 23.99	10	14	10	0
24 - 24.99	9	11	10	0
25 - 25.99	7	11	8	0
26 - 26.99	6	8	6	0
27 - 27.99	7	9	8	0
28 - 28.99	6	7	6	0
29 - 29.99	5	8	6	0
30 - 30.99	5	9	9	0
31 - 31.99	5	4	4	1
32 - 32.99	5	1	1	4
33 - 33.99	5	0	0	5
34 - 34.99	5	1	1	4
35 - 35.99	5	0	0	5
Totals	342	328	308	57

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Table 10.2: The number of spotted seatrout assigned to each total length-at-age category for 308 fish sampled for otolith age determination in Virginia during 2015.

	Age										
Interval	0	1	2	3	4	5	6	7	8	10	Totals
11 - 11.99	1	0	0	0	0	0	0	0	0	0	1
12 - 12.99	29	0	0	0	0	0	0	0	0	0	29
13 - 13.99	6	12	0	0	0	0	0	0	0	0	18
14 - 14.99	3	13	0	0	0	0	0	0	0	0	16
15 - 15.99	2	21	1	0	0	0	0	0	0	0	24
16 - 16.99	2	19	7	0	3	0	0	0	0	0	31
17 - 17.99	5	13	9	2	2	0	0	0	0	0	31
18 - 18.99	2	3	8	4	8	2	0	0	0	0	27
19 - 19.99	1	1	2	1	7	0	1	0	0	0	13
20 - 20.99	1	1	3	3	14	3	0	0	0	0	25
21 - 21.99	0	0	2	1	9	0	0	0	0	0	12
22 - 22.99	0	0	0	0	10	1	0	1	0	0	12
23 - 23.99	0	0	1	0	5	3	0	1	0	0	10
24 - 24.99	0	0	0	1	8	1	0	0	0	0	10
25 - 25.99	0	0	0	0	7	1	0	0	0	0	8
26 - 26.99	0	0	0	0	5	0	0	0	0	1	6
27 - 27.99	0	0	0	0	5	2	0	0	0	1	8
28 - 28.99	0	0	0	0	2	3	1	0	0	0	6
29 - 29.99	0	0	0	0	0	5	1	0	0	0	6
30 - 30.99	0	0	0	0	0	1	2	5	1	0	9
31 - 31.99	0	0	0	0	0	0	0	3	1	0	4
32 - 32.99	0	0	0	0	0	0	1	0	0	0	1
34 - 34.99	0	0	0	0	0	0	0	0	1	0	1
Totals	52	83	33	12	85	22	6	10	3	2	308

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Table 10.3: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for spotted seatrout sampled for age determination in Virginia during 2015.

	Age									
Interval	0	1	2	3	4	5	6	7	8	10
11 - 11.99	1	0	0	0	0	0	0	0	0	0
12 - 12.99	1	0	0	0	0	0	0	0	0	0
13 - 13.99	0.33	0.67	0	0	0	0	0	0	0	0
14 - 14.99	0.19	0.81	0	0	0	0	0	0	0	0
15 - 15.99	0.08	0.88	0.04	0	0	0	0	0	0	0
16 - 16.99	0.06	0.61	0.23	0	0.1	0	0	0	0	0
17 - 17.99	0.16	0.42	0.29	0.06	0.06	0	0	0	0	0
18 - 18.99	0.07	0.11	0.3	0.15	0.3	0.07	0	0	0	0
19 - 19.99	0.08	0.08	0.15	0.08	0.54	0	0.08	0	0	0
20 - 20.99	0.04	0.04	0.12	0.12	0.56	0.12	0	0	0	0
21 - 21.99	0	0	0.17	0.08	0.75	0	0	0	0	0
22 - 22.99	0	0	0	0	0.83	0.08	0	0.08	0	0
23 - 23.99	0	0	0.1	0	0.5	0.3	0	0.1	0	0
24 - 24.99	0	0	0	0.1	0.8	0.1	0	0	0	0
25 - 25.99	0	0	0	0	0.88	0.12	0	0	0	0
26 - 26.99	0	0	0	0	0.83	0	0	0	0	0.17
27 - 27.99	0	0	0	0	0.62	0.25	0	0	0	0.12
28 - 28.99	0	0	0	0	0.33	0.5	0.17	0	0	0
29 - 29.99	0	0	0	0	0	0.83	0.17	0	0	0
30 - 30.99	0	0	0	0	0	0.11	0.22	0.56	0.11	0
31 - 31.99	0	0	0	0	0	0	0	0.75	0.25	0
32 - 32.99	0	0	0	0	0	0	1	0	0	0
34 - 34.99	0	0	0	0	0	0	0	0	1	0

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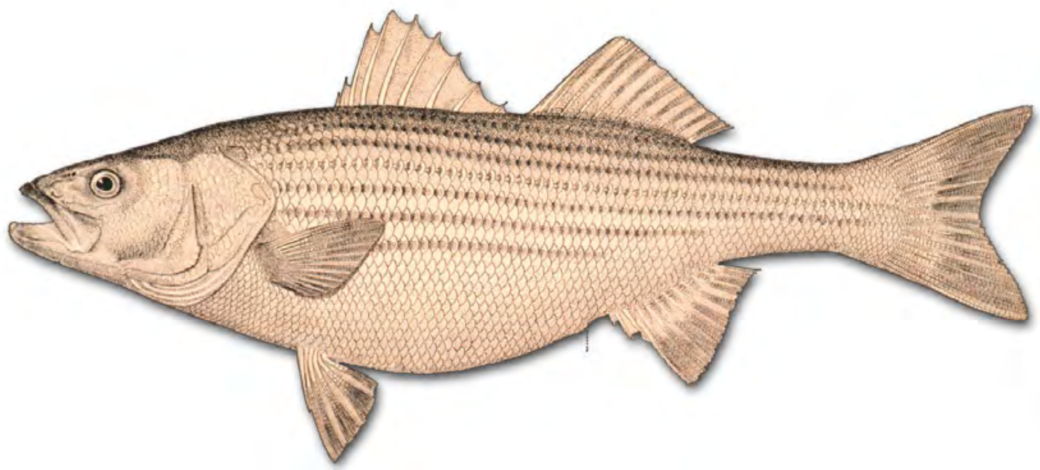
Chapter 11

STRIPED BASS *Morone saxatilis*



11.1 INTRODUCTION

We aged a total of 885 striped bass, *Morone saxatilis*, using their scales collected by the VMRC's Biological Sampling Program in 2015. Of 885 aged fish, 606 and 279 fish were collected in Chesapeake Bay (bay fish) and Virginia waters of the Atlantic Ocean (ocean fish), respectively. The average bay fish age was 8.5 years with a standard deviation of 4.3 and a standard error of 0.17. Twenty age classes (3 to 22) were represented in the bay fish, comprising fish from the 1993 to 2012 year classes. The bay fish sample in 2015 was dominated by the year class of 2011 with 22%. The average ocean fish age was 11.7 years with a standard deviation of 2.9 and a standard error of 0.17. Seventeen age classes (4, and 6 to 21) were represented in the ocean fish, comprising fish from the 1994 to 2009, and 2011 year classes. The ocean fish sample in 2015 was dominated by the year classes of 2004, 2005, and 2003 with 17%, 16%, and 16%, respectively. We also aged a total of 324 fish using their otoliths in addition to ageing their scales. The otolith ages were compared to the scale ages to examine how close both ages were to one another (see details in Results).

11.2 METHODS

11.2.1 Sample size for ageing

We estimated sample sizes for ageing striped bass collected in both Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015, respectively, using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 CV^2 + B_a/L} \tag{11.1}$$

where A is the sample size for ageing striped bass in 2015; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of striped bass used by VMRC to estimate length distribution of the catches from 2009 to 2013. θ_a, V_a, B_a, and CV were calculated using pooled age-length data of striped bass collected from 2009 to 2013 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (11.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2009 to 2013 catch. A_l is number of fish to be aged for length interval l in 2015.

11.2.2 Handling of collection

Sagittal otoliths (hereafter, referred to as "otoliths") and scales were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted based on date of capture. Their envelope labels were verified against VMRC's collection data, and each fish assigned a unique Age and Growth Laboratory identification number. All otoliths and scales were stored dry within their original labeled coin envelopes; otoliths were contained inside protective Axygen 2.0 ml microtubes.

11.2.3 Preparation

Scales

Striped bass scales were prepared for age and growth analysis by making acetate impressions of the scale microstructure. Due to extreme variation in the size and shape of scales from individual fish, we selected only those scales that had even margins and which were of uniform size. We selected a range of four to six preferred scales (based on overall scale size) from each fish, making sure that only non-regenerated scales were used. Scale impressions were made on extruded clear acetate sheets (25 mm x 75 mm) with a Carver Laboratory Heated Press (model "C"). The scales were pressed with the following settings:

Pressure: 15000 psi
Temperature: 77 °C (170 °F)
Time: 5 to 10 min

Striped bass scales that were the size of a quarter (coin) or larger, were pressed individually for up to twenty minutes. After pressing, the impressions were viewed with a Bell and Howell microfiche reader and checked again for regeneration and incomplete margins. Impressions that were too light, or when all scales were regenerated a new impression was made using different scales from the same fish.

Click here to obtain the protocol at the CQFE website on how to prepare scale impression for ageing striped bass.

Otoliths

We used our thin-section and bake technique to process spadefish sagittal otoliths (hereafter, referred to as "otoliths") for age determination. Otolith preparation began by randomly selecting either the right or left otolith. Each whole otolith was placed in a ceramic "Coors" spot plate well and baked in a Thermolyne 1400 furnace at 400 °C. Baking time was dependent on the otolith's size and gauged by color, with a light caramel color desired. Once a suitable color was achieved the baked otolith was embedded in epoxy resin with its distal surface orientated downwards and allowed to harden overnight. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet™ low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The otolith was positioned so the blades straddled each side of the otolith focus. It was crucial that this cut be perpendicular to the long axis of the otolith. Failure to do so resulted in broadening and distorted winter growth zones. A proper cut resulted in annuli that were clearly defined and delineated. Once cut, thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing striped bass.

11.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of reading the information contained in its otolith, the date of its capture, and the species-specific period when it deposits its annulus. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called annuli. Technically, an otolith annulus is the combination of both the opaque and the translucent bands. In practice, only the opaque bands are counted as annuli. The number of these visible dark bands replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the otolith section is examined for translucent growth. If no translucent growth is visible beyond the last dark annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last dark annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the dark band of the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of dark bands in the otolith. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday", but before the dark band deposition period, is interpreted as being toward the next age class.

For example, striped bass otolith deposition occurs between April and June (Secor et al. 1995). A striped bass captured between January 1 and June 30, before the end of the species' annulus formation period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of June 30, the period of annulus formation, which would be noted as 4+4.

Striped bass scales are also considered to have a deposition between April and June (Secor et al. 1995), and age class assignment using these hard-parts is conducted in the same way as otoliths.

All striped bass samples (scale pressings and sectioned otoliths) were aged by two different readers in chronological order based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish again without any knowledge of previously estimated ages or lengths, then assigned a final age to the fish. When the age readers were unable to agree on a final age, the fish was excluded from further analysis.

Scales

We determined fish age by viewing acetate impressions of scales (Figure 11.1) with a standard Bell and Howell R-735 microfiche reader equipped with 20 and 29 mm lenses. Annuli on striped bass scales are identified based on two scale microstructure features, "crossing over" and circuli disruption. Primarily, "crossing over" in the lateral margins near the posterior/anterior interface of the scale is used to determine the origin of the annulus. Here compressed circuli (annulus) "cross-over" the previously deposited circuli of the previous year's growth. Typically annuli of the first three years can be observed transversing this interface as dark bands. These bands

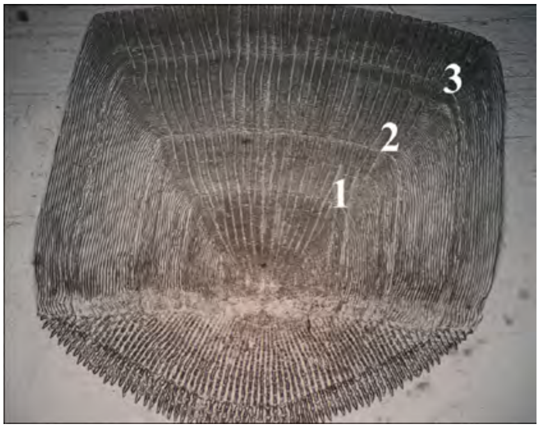


Figure 11.1: Scale impression of a 3 year-old striped bass.

remain consistent throughout the posterior field and rejoin the posterior/anterior interface on the opposite side of the focus. Annuli can also be observed in the anterior lateral field of the scale. Here the annuli typically reveal a pattern of discontinuous and suddenly breaking segmented circuli. This event can also be distinguished by the presence of concentric white lines, which are typically associated with the disruption of circuli.

Annuli can also be observed bisecting the perpendicular plain of the radial striations in the anterior field of the scale. Radii emanate out from the focus of the scale towards the outer corner margins of the anterior field. These radial striations consist mainly of segmented concave circuli. The point of intersection between radii and annuli results in a "straightening out" of the concave circuli. This straightening of the circuli should be consistent throughout the entire anterior field of the scale. This event is further amplified by the presence of concave circuli neighboring both directly above and below the annulus. The first year's annulus can be difficult to locate on some scales. It is typically best identified in the lateral field of the anterior portion of the scale. The distance from the focus to the first year's annulus is typically larger with respect to the following annuli. For the annuli two through six, summer growth generally decreases proportionally. For ages greater than six, a crowding effect of the annuli near the outer margins of the scale is observed. This crowding effect creates difficulties in edge interpretation. At this point it is best to focus on the straightening of the circuli at the anterior margins of the scale.

When ageing young striped bass, zero through age two, extreme caution must be taken as not to over age the structure. In young fish there is no point of reference to aid in the determination of the first year; this invariably results in over examination of the scale and such events as hatching or saltwater incursion marks (checks) may be interpreted as the first year.

Otoliths

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and darkfield polarization at between 8 and 20 times magnification (Figure 11.2). Each reader aged all of the otolith samples. By convention an annulus is identified as the narrow opaque zone, or winter growth. Typically the first year's annulus can be determined by first locating the focus of the otolith. The focus is generally located, depending on preparation, in the center of the otolith, and is visually well defined as a dark oblong region. The first year's annulus can be located directly below the focus, along the

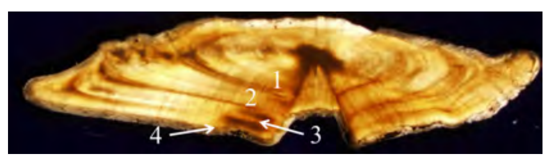


Figure 11.2: Otolith thin-section of a 4 year-old striped bass with the last annulus on the edge of the thin-section

outer ridge of the sulcal groove on the ventral and dorsal sides of the otolith. This insertion point along the sulcal ridge resembles a check mark (not to be confused with a false annulus). Here the annulus can be followed outwards along the ventral and dorsal surfaces where it encircles the focus. Subsequent annuli also emanate from the sulcal ridge; however, they do not encircle the focus, but rather travel outwards to the distal surface of the otolith. To be considered a true annulus, each annulus must be rooted in the sulcus and travel without interruption to the distal surface of the otolith. The annuli in striped bass have a tendency to split as they advance towards the distal surface. As a result, it is critical that reading path proceed in a direction down the sulcal ridge and outwards to the distal surface.

Click here to obtain the protocol at the CQFE website on how to age striped bass using their otolith thin-sections.

11.2.5 Comparison Tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for following comparisons: 1) between the two readers in the current year; 2) within each reader in the current year; 3) time-series bias between the current and previous years within each reader; and 4) between scale and otoliths ages. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2000 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

11.3 RESULTS

11.3.1 Sample size

We estimated a sample size of 573 bay striped bass in 2015, ranging in length interval from 17 to 50 inches (Table 11.1). This sample size provided a range in CV for age composition approximately from the smallest CV of 11% for age 7 to the largest CV of 23% for age 3 and 14 of the bay fish. We randomly selected and aged 606 fish from 784 striped bass collected by VMRC in Chesapeake Bay in 2015. We fell short in our over-all collections for this optimal length-class sampling estimate by 43 fish. We were short of few fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

We estimated a sample size of 496 ocean striped bass in 2015, ranging in length interval from 26 to 56 inches (Table 11.2). This sample size provided a range in CV for age composition approximately from the smallest CV of 1% for age 21 to the largest CV of 25% for age 6 of the ocean fish. We aged all 279 striped bass collected by VMRC in Virginia waters of the Atlantic Ocean in 2015. We fell short in our over-all collections for this optimal length-class sampling estimate by 231 fish. However, we were short of many fish from in the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would definitely be influenced significantly. Therefore, precaution should be used when developing ALK using these age data.

11.3.2 Scales

Reader 1 had high self-precision and Read 2 had moderate self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 60% (1 year or less agreement of 84%) and a *CV* of 4.7% (test of symmetry: $\chi^2 = 18$, $df = 15$, $P = 0.2627$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 38% (1 year or less agreement of 88%) and a *CV* of 6.4% (test of symmetry: $\chi^2 = 14.73$, $df = 16$, $P = 0.5442$). There was an evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 42% (1 year or less agreement of 75%) and a *CV* of 7% (test of symmetry: $\chi^2 = 256.54$, $df = 67$, $P < 0.0001$) (Figure 11.3).

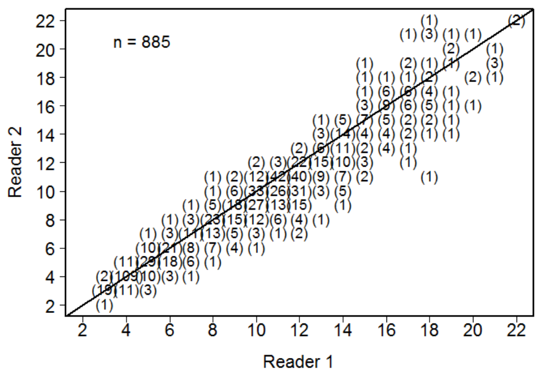


Figure 11.3: Between-reader comparison of scale age estimates for striped bass collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

Reader 1 had no time series bias while Read 2 does. Reader 1 had an agreement of 58% (1 year or less agreement of 95%) with ages of fish aged in 2000 with a *CV* of 4.9% (test of symmetry: $\chi^2 = 11.33$, $df = 13$, $P = 0.5829$). Reader 2 had an agreement of 50% (1 year or less agreement of 93%) with a *CV* of 5.9% (test of symmetry: $\chi^2 = 22.67$, $df = 12$, $P = 0.0307$).

Of the 606 bay striped bass aged with scales, 20 age classes (3 to 22) were represented (Table 11.3). The average age for the sample was 8.5 years. The standard deviation and standard error were 4.3 and 0.17, respectively. Year-class data (Figure 11.4) indicates that recruitment into the fishery in Chesapeake Bay begins at age 3, which corresponds to the 2012 year-class for striped bass caught in 2015. Striped bass in the sample in 2015 was dominated by the year class of 2011 with 22%. The sex ratio of male to female was 1:1.51 for the bay fish.

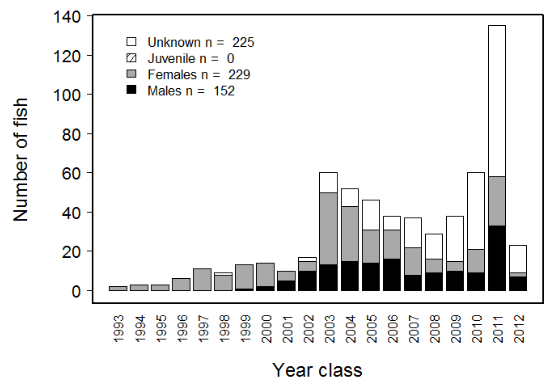


Figure 11.4: Year-class frequency distribution for striped bass collected in Chesapeake Bay, Virginia for ageing in 2015. Distribution is broken down by sex and estimated using scale ages. 'Unknown' represents the fish gonads that were not available for examination or were not examined for sex during sampling.

Of the 279 ocean striped bass aged with scales, 17 age classes (4, and 6 to 21) were represented (Table 11.4). The average age for the sample was 11.7 years. The standard deviation and standard error were 2.9 and 0.17, respectively. Year-class data (Figure 11.5) indicates that recruitment into the fishery in Virginia waters of Atlantic ocean begins at age 4, which corresponds to the 2011 year-class for striped bass caught in 2015. Striped bass in the sample in 2015 was dominated by the year classes of 2004, 2005, and 2003 with 17%, 16%, and 16%, respectively. The sex ratio of male to female was 1:2.24 for the ocean fish.

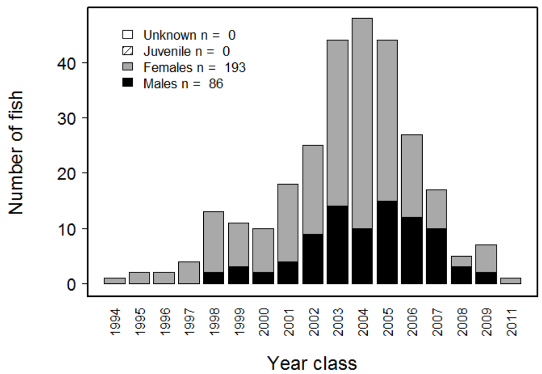


Figure 11.5: Year-class frequency distribution for striped bass collected in Virginia waters of the Atlantic Ocean for ageing in 2015. Distribution is broken down by sex and estimated using scale ages. 'Unknown' represents the fish gonads that were not available for examination or were not examined for sex during sampling.

11.3.3 Otoliths

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 94% and a *CV* of 0.4% (test of symmetry: $\chi^2 = 3$, $df = 3$, $P = 0.3916$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 94% and a *CV* of 0.3% (test of symmetry: $\chi^2 = 3$, $df = 3$, $P = 0.3916$). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 90% (1 year or less agreement of 99%) and a *CV* of 0.7% (test of symmetry: $\chi^2 = 15$, $df = 17$, $P = 0.5955$) (Figure 11.6).

There was no time-series bias for both readers. Reader 1 had an agreement of 90% with ages of fish aged in 2003 with a *CV* of 0.8% (test of symmetry: $\chi^2 = 4$, $df = 5$, $P = 0.5494$). Reader 2 had an agreement of 95% with a *CV* of 0.4% (test of symmetry:

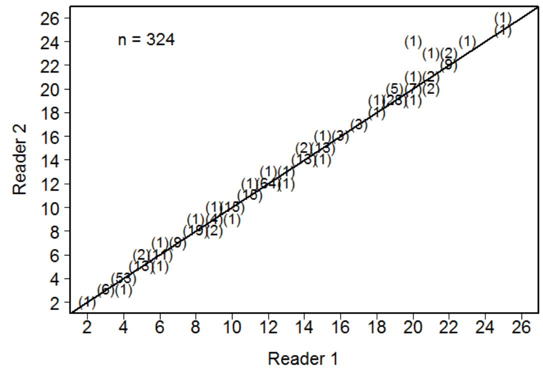


Figure 11.6: Between-reader comparison of otolith age estimates for striped bass collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

$\chi^2 = 1$, $df = 2$, $P = 0.6065$).

Of the 324 striped bass aged with otoliths, 24 age classes (2 to 23, and 25 to 26) were represented (Table 11.5). The average age for the sample was 11.3 years. The standard deviation and standard error were 5.7 and 0.32, respectively.

11.3.4 Comparison of scale and otolith ages

We aged 324 striped bass using scales and otoliths. There was an evidence of systematic disagreement between otolith and scale ages (test of symmetry: $\chi^2 = 130.63$, $df = 55$, $P < 0.0001$) with an average *CV* of 5.8%. There was an agreement of 48% between scale and otoliths ages whereas scales were assigned a lower and higher age than otoliths for 44% and 8% of the fish, respectively (Figure 11.7). There was also an evidence of bias between otolith and scale ages using an age bias plot (Figure 11.8), with scale generally assigned higher ages for younger fish and lower ages for older fish than otolith age estimates.

Figure 11.7: Comparison of scale and otolith age estimates for striped bass collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

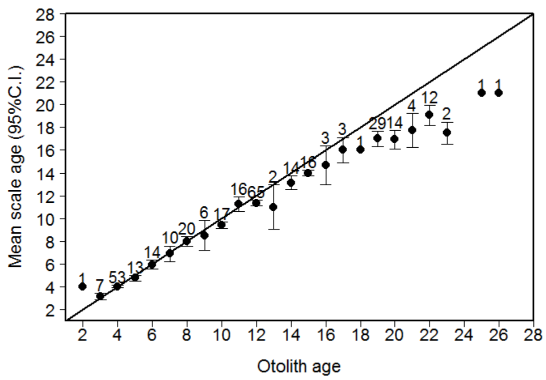


Figure 11.8: Age-bias plot for striped bass scale and otolith age estimates in 2015.

11.3.5 Age-Length-Key (ALK)

We developed an age-length-key for both bay (Table 11.6) and ocean fish (Table 11.7) using scale ages, separately. The ALK can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using scale ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

11.4 RECOMMENDATIONS

We recommend that VMRC and ASMFC use otoliths for ageing striped bass. Although preparation time is greater for otoliths compared to scales, nonetheless as the mean age of striped bass increases in the recovering fishery, otoliths should provide more reliable estimates of age (Liao et al. 2013; Secor et al. 1995). We will continue to compare the age estimates between otoliths and scales.

11.5 REFERENCES

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Table 11.1: Number of bay striped bass collected and aged in each 1-inch length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
17 - 17.99	5	0	0	5
18 - 18.99	9	18	14	0
19 - 19.99	20	39	27	0
20 - 20.99	22	51	31	0
21 - 21.99	28	46	29	0
22 - 22.99	29	41	30	0
23 - 23.99	30	37	30	0
24 - 24.99	32	43	30	2
25 - 25.99	28	39	28	0
26 - 26.99	27	29	27	0
27 - 27.99	23	33	28	0
28 - 28.99	18	22	18	0
29 - 29.99	15	23	19	0
30 - 30.99	14	26	14	0
31 - 31.99	16	31	17	0
32 - 32.99	20	31	30	0
33 - 33.99	20	25	25	0
34 - 34.99	24	21	21	3
35 - 35.99	29	22	21	8
36 - 36.99	34	31	31	3
37 - 37.99	31	23	23	8
38 - 38.99	17	26	18	0
39 - 39.99	12	16	11	1
40 - 40.99	11	25	18	0
41 - 41.99	8	10	8	0
42 - 42.99	9	12	9	0
43 - 43.99	7	11	8	0
44 - 44.99	5	18	13	0
45 - 45.99	5	15	9	0
46 - 46.99	5	13	12	0
47 - 47.99	5	4	4	1
48 - 48.99	5	2	2	3
49 - 49.99	5	1	1	4
50 - 50.99	5	0	0	5
Totals	573	784	606	43

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Table 11.2: Number of ocean striped bass collected and aged in each 1-inch length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
26 - 26.99	5	0	0	5
27 - 27.99	5	0	0	5
28 - 28.99	12	4	4	8
29 - 29.99	24	4	4	20
30 - 30.99	25	5	5	20
31 - 31.99	32	12	12	20
32 - 32.99	32	11	11	21
33 - 33.99	34	18	18	16
34 - 34.99	38	20	20	18
35 - 35.99	49	31	31	18
36 - 36.99	48	31	31	17
37 - 37.99	51	34	34	17
38 - 38.99	33	19	19	14
39 - 39.99	22	17	17	5
40 - 40.99	17	11	11	6
41 - 41.99	11	9	9	2
42 - 42.99	11	8	8	3
43 - 43.99	7	10	10	0
44 - 44.99	5	11	11	0
45 - 45.99	5	9	9	0
46 - 46.99	5	4	4	1
47 - 47.99	5	6	6	0
48 - 48.99	5	3	3	2
49 - 49.99	5	2	2	3
50 - 50.99	5	0	0	5
56 - 56.99	5	0	0	5
Totals	496	279	279	231

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Table 11.3: The number of striped bass assigned to each total length-at-age category for 606 fish sampled for scale age determination in Chesapeake Bay, Virginia during 2015.

	Age																				
Interval	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	Totals
18 - 18.99	7	4	3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	14
19 - 19.99	8	17	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	27
20 - 20.99	4	19	6	1	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	31
21 - 21.99	2	21	5	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	29
22 - 22.99	1	20	6	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	30
23 - 23.99	0	19	6	3	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	30
24 - 24.99	1	14	6	7	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	30
25 - 25.99	0	8	12	2	4	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	28
26 - 26.99	0	6	7	6	4	1	2	0	1	0	0	0	0	0	0	0	0	0	0	0	27
27 - 27.99	0	6	4	5	4	3	3	3	0	0	0	0	0	0	0	0	0	0	0	0	28
28 - 28.99	0	0	1	6	2	2	4	1	2	0	0	0	0	0	0	0	0	0	0	0	18
29 - 29.99	0	1	2	0	1	4	4	2	3	2	0	0	0	0	0	0	0	0	0	0	19
30 - 30.99	0	0	0	0	2	4	4	1	2	1	0	0	0	0	0	0	0	0	0	0	14
31 - 31.99	0	0	0	3	4	1	2	3	1	2	1	0	0	0	0	0	0	0	0	0	17
32 - 32.99	0	0	0	2	1	5	2	3	6	7	2	1	1	0	0	0	0	0	0	0	30
33 - 33.99	0	0	0	1	0	2	6	3	2	4	5	1	0	1	0	0	0	0	0	0	25
34 - 34.99	0	0	0	0	0	5	2	5	6	1	1	0	1	0	0	0	0	0	0	0	21
35 - 35.99	0	0	0	0	0	3	1	8	3	2	3	1	0	0	0	0	0	0	0	0	21
36 - 36.99	0	0	0	0	1	4	2	7	9	7	0	1	0	0	0	0	0	0	0	0	31
37 - 37.99	0	0	0	0	1	0	2	5	6	7	2	0	0	0	0	0	0	0	0	0	23
38 - 38.99	0	0	0	0	0	0	2	2	5	9	0	0	0	0	0	0	0	0	0	0	18
39 - 39.99	0	0	0	0	0	0	0	3	2	5	1	0	0	0	0	0	0	0	0	0	11
40 - 40.99	0	0	0	0	0	0	1	0	3	7	2	1	3	1	0	0	0	0	0	0	18
41 - 41.99	0	0	0	0	0	0	0	0	1	4	0	2	0	0	0	0	1	0	0	0	8
42 - 42.99	0	0	0	0	0	0	0	0	0	2	0	1	0	1	3	2	0	0	0	0	9
43 - 43.99	0	0	0	0	0	0	0	0	0	0	0	2	2	1	1	0	1	1	0	0	8
44 - 44.99	0	0	0	0	0	0	0	0	0	0	0	0	3	3	2	1	1	1	1	1	13
45 - 45.99	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	4	1	0	2	0	9
46 - 46.99	0	0	0	0	0	0	0	0	0	0	0	0	2	3	2	3	0	1	0	1	12
47 - 47.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	0	0	0	4
48 - 48.99	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	2
49 - 49.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1
Totals	23	135	60	38	29	37	38	46	52	60	17	10	14	13	9	11	6	3	3	2	606

(Go back to text)

Table 11.4: The number of striped bass assigned to each total length-at-age category for 279 fish sampled for scale age determination in Virginia waters of Atlantic ocean during 2015.

	Age																
Interval	4	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	Totals
28 - 28.99	0	3	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	4
29 - 29.99	1	0	0	2	0	0	0	0	0	0	1	0	0	0	0	0	0	4
30 - 30.99	0	3	0	1	0	1	0	0	0	0	0	0	0	0	0	0	0	5
31 - 31.99	0	1	1	3	3	3	1	0	0	0	0	0	0	0	0	0	0	12
32 - 32.99	0	0	2	3	1	2	0	3	0	0	0	0	0	0	0	0	0	11
33 - 33.99	0	0	1	2	4	5	2	2	0	1	1	0	0	0	0	0	0	18
34 - 34.99	0	0	0	3	2	8	4	3	0	0	0	0	0	0	0	0	0	20
35 - 35.99	0	0	0	1	5	10	6	5	3	0	0	1	0	0	0	0	0	31
36 - 36.99	0	0	1	0	3	6	5	8	5	2	0	0	1	0	0	0	0	31
37 - 37.99	0	0	0	2	6	5	14	4	2	1	0	0	0	0	0	0	0	34
38 - 38.99	0	0	0	0	1	3	4	6	1	2	1	0	1	0	0	0	0	19
39 - 39.99	0	0	0	0	1	1	5	6	3	0	1	0	0	0	0	0	0	17
40 - 40.99	0	0	0	0	0	0	3	0	2	2	1	3	0	0	0	0	0	11
41 - 41.99	0	0	0	0	0	0	1	4	2	1	0	1	0	0	0	0	0	9
42 - 42.99	0	0	0	0	0	0	1	0	5	1	0	1	0	0	0	0	0	8
43 - 43.99	0	0	0	0	0	0	2	0	0	3	2	0	1	0	0	2	0	10
44 - 44.99	0	0	0	0	0	0	0	1	1	3	1	1	4	0	0	0	0	11
45 - 45.99	0	0	0	0	0	0	0	1	1	1	1	2	1	2	0	0	0	9
46 - 46.99	0	0	0	0	0	0	0	1	0	1	0	2	0	0	0	0	0	4
47 - 47.99	0	0	0	0	0	0	0	0	0	0	1	0	3	2	0	0	0	6
48 - 48.99	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	1	3
49 - 49.99	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	2
Totals	1	7	5	17	27	44	48	44	25	18	10	11	13	4	2	2	1	279

(Go back to text)

Table 11.5: The number of striped bass assigned to each total length-at-age category for 324 fish sampled for otolith age determination in Chesapeake Bay and Virginia waters of Atlantic Ocean during 2015.

	Age																							
Interval	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	25	26	Totals
18 - 18.99	0	3	6	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	11
19 - 19.99	0	2	11	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	15
20 - 20.99	0	1	10	2	0	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	15
21 - 21.99	0	1	7	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	9
22 - 22.99	0	0	6	0	3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	9
23 - 23.99	0	0	7	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	10
24 - 24.99	0	0	3	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	5
25 - 25.99	1	0	0	2	1	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	6
26 - 26.99	0	0	0	2	2	3	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	7
27 - 27.99	0	0	3	0	0	0	4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	7
28 - 28.99	0	0	0	0	3	0	2	1	2	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	9
29 - 29.99	0	0	0	1	0	0	2	0	1	0	2	0	1	0	0	0	0	0	0	0	0	0	0	0	7
30 - 30.99	0	0	0	0	0	2	2	1	2	0	3	0	0	0	0	0	0	0	0	0	0	0	0	0	10
31 - 31.99	0	0	0	0	1	0	0	1	2	0	2	0	0	0	0	0	0	0	0	0	0	0	0	0	6
32 - 32.99	0	0	0	0	1	0	2	1	0	3	3	0	1	0	0	0	0	0	0	0	0	0	0	0	11
33 - 33.99	0	0	0	0	0	0	1	1	0	2	1	0	2	2	1	0	0	0	0	0	0	0	0	0	10
34 - 34.99	0	0	0	0	0	0	1	0	2	0	3	0	1	0	0	0	0	1	0	0	0	0	0	0	8
35 - 35.99	0	0	0	0	0	0	2	0	2	1	2	2	1	0	0	0	0	0	0	0	0	0	0	0	10
36 - 36.99	0	0	0	0	0	0	3	0	0	1	5	0	0	0	1	0	0	0	0	1	0	0	0	0	11
37 - 37.99	0	0	0	0	0	0	1	1	4	1	6	0	0	0	0	0	0	0	0	0	0	0	0	0	13
38 - 38.99	0	0	0	0	0	0	0	0	1	2	14	0	0	1	0	0	0	0	0	0	0	0	0	0	18
39 - 39.99	0	0	0	0	0	0	0	0	1	2	8	0	3	0	0	0	0	0	0	0	0	0	0	0	14
40 - 40.99	0	0	0	0	0	0	0	0	0	2	10	0	1	4	0	0	1	2	1	0	0	0	0	0	21
41 - 41.99	0	0	0	0	0	0	0	0	0	1	4	0	0	4	1	0	0	1	0	0	0	0	0	0	11
42 - 42.99	0	0	0	0	0	0	0	0	0	0	1	0	4	0	0	1	0	0	1	1	0	2	0	0	10
43 - 43.99	0	0	0	0	0	0	0	0	0	1	0	0	0	3	0	0	0	2	3	1	1	0	0	0	11
44 - 44.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	8	1	0	2	0	0	1	14
45 - 45.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	3	4	0	2	0	1	0	10
46 - 46.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	6	3	0	3	0	0	0	13
47 - 47.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	2	0	1	2	0	0	0	6
48 - 48.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	3	1	0	1	0	0	0	5
49 - 49.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	2
Totals	1	7	53	13	14	10	20	6	17	16	65	2	14	16	3	3	1	29	14	4	12	2	1	1	324

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Table 11.6: Age-Length key, as proportion-at-age in each 1-inch length interval, based on scale ages for striped bass sampled in Chesapeake Bay, Virginia during 2015.

	Age																			
Interval	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22
18 - 18.99	0.5	0.29	0.21	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
19 - 19.99	0.3	0.63	0.07	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
20 - 20.99	0.13	0.61	0.19	0.03	0	0.03	0	0	0	0	0	0	0	0	0	0	0	0	0	0
21 - 21.99	0.07	0.72	0.17	0.03	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
22 - 22.99	0.03	0.67	0.2	0.03	0.03	0.03	0	0	0	0	0	0	0	0	0	0	0	0	0	0
23 - 23.99	0	0.63	0.2	0.1	0.07	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
24 - 24.99	0.03	0.47	0.2	0.23	0.07	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
25 - 25.99	0	0.29	0.43	0.07	0.14	0.04	0.04	0	0	0	0	0	0	0	0	0	0	0	0	0
26 - 26.99	0	0.22	0.26	0.22	0.15	0.04	0.07	0	0.04	0	0	0	0	0	0	0	0	0	0	0
27 - 27.99	0	0.21	0.14	0.18	0.14	0.11	0.11	0.11	0	0	0	0	0	0	0	0	0	0	0	0
28 - 28.99	0	0	0.06	0.33	0.11	0.11	0.22	0.06	0.11	0	0	0	0	0	0	0	0	0	0	0
29 - 29.99	0	0.05	0.11	0	0.05	0.21	0.21	0.11	0.16	0.11	0	0	0	0	0	0	0	0	0	0
30 - 30.99	0	0	0	0	0.14	0.29	0.29	0.07	0.14	0.07	0	0	0	0	0	0	0	0	0	0
31 - 31.99	0	0	0	0.18	0.24	0.06	0.12	0.18	0.06	0.12	0.06	0	0	0	0	0	0	0	0	0
32 - 32.99	0	0	0	0.07	0.03	0.17	0.07	0.1	0.2	0.23	0.07	0.03	0.03	0	0	0	0	0	0	0
33 - 33.99	0	0	0	0.04	0	0.08	0.24	0.12	0.08	0.16	0.2	0.04	0	0.04	0	0	0	0	0	0
34 - 34.99	0	0	0	0	0	0.24	0.1	0.24	0.29	0.05	0.05	0	0.05	0	0	0	0	0	0	0
35 - 35.99	0	0	0	0	0	0.14	0.05	0.38	0.14	0.1	0.14	0.05	0	0	0	0	0	0	0	0
36 - 36.99	0	0	0	0	0.03	0.13	0.06	0.23	0.29	0.23	0	0.03	0	0	0	0	0	0	0	0
37 - 37.99	0	0	0	0	0.04	0	0.09	0.22	0.26	0.3	0.09	0	0	0	0	0	0	0	0	0
38 - 38.99	0	0	0	0	0	0	0.11	0.11	0.28	0.5	0	0	0	0	0	0	0	0	0	0
39 - 39.99	0	0	0	0	0	0	0	0.27	0.18	0.45	0.09	0	0	0	0	0	0	0	0	0
40 - 40.99	0	0	0	0	0	0	0.06	0	0.17	0.39	0.11	0.06	0.17	0.06	0	0	0	0	0	0
41 - 41.99	0	0	0	0	0	0	0	0	0.12	0.5	0	0.25	0	0	0	0	0.12	0	0	0
42 - 42.99	0	0	0	0	0	0	0	0	0	0.22	0	0.11	0	0.11	0.33	0.22	0	0	0	0
43 - 43.99	0	0	0	0	0	0	0	0	0	0	0	0.25	0.25	0.12	0.12	0	0.12	0.12	0	0
44 - 44.99	0	0	0	0	0	0	0	0	0	0	0	0	0.23	0.23	0.15	0.08	0.08	0.08	0.08	0.08
45 - 45.99	0	0	0	0	0	0	0	0	0	0	0	0	0.11	0.11	0	0.44	0.11	0	0.22	0
46 - 46.99	0	0	0	0	0	0	0	0	0	0	0	0	0.17	0.25	0.17	0.25	0	0.08	0	0.08
47 - 47.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0.25	0.25	0.25	0.25	0	0	0
48 - 48.99	0	0	0	0	0	0	0	0	0	0	0	0	0.5	0	0	0	0.5	0	0	0
49 - 49.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0

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Table 11.7: Age-Length key, as proportion-at-age in each 1-inch length interval, based on scale ages for striped bass sampled in Virginia waters of the Atlantic Ocean during 2015.

	Age															
Interval	4	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21
28 - 28.99	0	0.75	0	0	0.25	0	0	0	0	0	0	0	0	0	0	0	0
29 - 29.99	0.25	0	0	0.5	0	0	0	0	0	0	0.25	0	0	0	0	0	0
30 - 30.99	0	0.6	0	0.2	0	0.2	0	0	0	0	0	0	0	0	0	0	0
31 - 31.99	0	0.08	0.08	0.25	0.25	0.25	0.08	0	0	0	0	0	0	0	0	0	0
32 - 32.99	0	0	0.18	0.27	0.09	0.18	0	0.27	0	0	0	0	0	0	0	0	0
33 - 33.99	0	0	0.06	0.11	0.22	0.28	0.11	0.11	0	0.06	0.06	0	0	0	0	0	0
34 - 34.99	0	0	0	0.15	0.1	0.4	0.2	0.15	0	0	0	0	0	0	0	0	0
35 - 35.99	0	0	0	0.03	0.16	0.32	0.19	0.16	0.1	0	0	0.03	0	0	0	0	0
36 - 36.99	0	0	0.03	0	0.1	0.19	0.16	0.26	0.16	0.06	0	0	0.03	0	0	0	0
37 - 37.99	0	0	0	0.06	0.18	0.15	0.41	0.12	0.06	0.03	0	0	0	0	0	0	0
38 - 38.99	0	0	0	0	0.05	0.16	0.21	0.32	0.05	0.11	0.05	0	0.05	0	0	0	0
39 - 39.99	0	0	0	0	0.06	0.06	0.29	0.35	0.18	0	0.06	0	0	0	0	0	0
40 - 40.99	0	0	0	0	0	0	0.27	0	0.18	0.18	0.09	0.27	0	0	0	0	0
41 - 41.99	0	0	0	0	0	0	0.11	0.44	0.22	0.11	0	0.11	0	0	0	0	0
42 - 42.99	0	0	0	0	0	0	0.12	0	0.62	0.12	0	0.12	0	0	0	0	0
43 - 43.99	0	0	0	0	0	0	0.2	0	0	0.3	0.2	0	0.1	0	0	0.2	0
44 - 44.99	0	0	0	0	0	0	0	0.09	0.09	0.27	0.09	0.09	0.36	0	0	0	0
45 - 45.99	0	0	0	0	0	0	0	0.11	0.11	0.11	0.11	0.22	0.11	0.22	0	0	0
46 - 46.99	0	0	0	0	0	0	0	0.25	0	0.25	0	0.5	0	0	0	0	0
47 - 47.99	0	0	0	0	0	0	0	0	0	0	0.17	0	0.5	0.33	0	0	0
48 - 48.99	0	0	0	0	0	0	0	0	0	0	0	0	0.33	0	0.33	0	0.33
49 - 49.99	0	0	0	0	0	0	0	0	0	0	0	0	0.5	0	0.5	0	0

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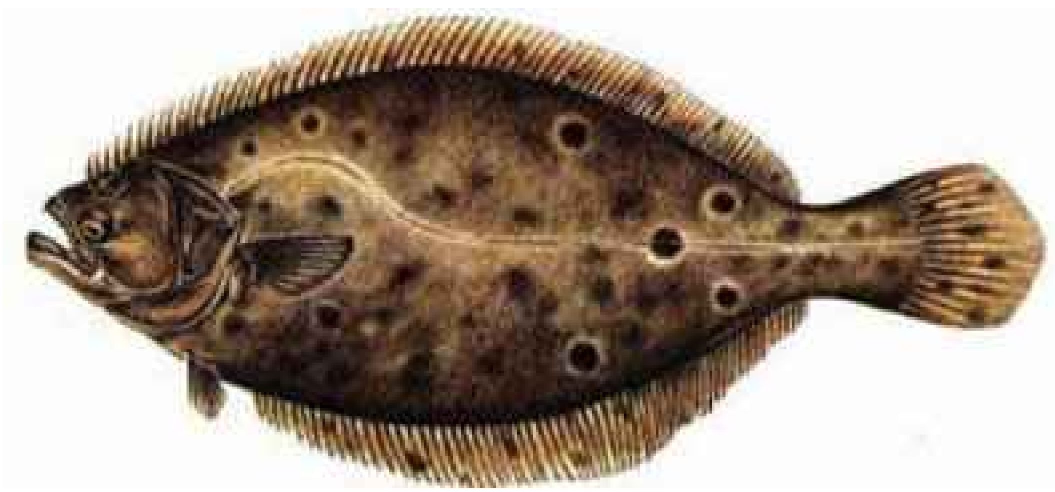
Chapter 12

SUMMER FLOUNDER *Paralichthys dentatus*



12.1 INTRODUCTION

We aged a total of 884 summer flounder,*Paralichthys dentatus*, using their scales collected by the VMRC's Biological Sampling Program in 2015. Of 884 aged fish, 371 and 513 fish were collected in Chesapeake Bay (bay fish) and Virginia waters of the Atlantic Ocean (ocean fish), respectively. The average bay fish age was 3.7 years with a standard deviation of 1.8 and a standard error of 0.09. Ten age classes (1 to 10) were represented in the bay fish, comprising fish from the 2005 to 2014 year classes. The bay fish sample in 2015 was dominated by the year class of 2012 with 33%. The average ocean fish age was 4.9 years with a standard deviation of 2 and a standard error of 0.09. Thirteen age classes (1 to 12, and 15) were represented in the ocean fish, comprising fish from the 2000, and 2003 to 2014 year classes. The ocean fish sample in 2015 was dominated by the year classes of 2010 and 2009 with 23% and 18%, respectively. We also aged a total of 294 fish using their otoliths in addition to ageing their scales. The otolith ages were compared to the scale ages to examine how close both ages were to one another (see details in Results).

12.2 METHODS

12.2.1 Sample size for ageing

We estimated sample sizes for ageing summer flounder collected in both Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015, respectively, using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 CV^2 + B_a/L} \qquad (12.1)$$

where A is the sample size for ageing summer flounder in 2015; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of summer flounder used by VMRC to estimate length distribution of the catches from 2009 to 2013. θ_a, V_a, B_a, and CV were calculated using pooled age-length data of summer flounder collected from 2009 to 2013 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (12.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2009 to 2013 catch. A_l is number of fish to be aged for length interval l in 2015.

12.2.2 Handling of collection

Sagittal otoliths (hereafter, referred to as "otoliths") and scales were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted based on date of capture, their envelope labels were verified against VMRC's collection data, and each fish assigned a unique Age and Growth Laboratory identification number. All otoliths and scales were stored dry within their original labeled coin envelopes; otoliths were contained inside protective Axygen 2.0 ml microtubes.

12.2.3 Preparation

Scales

Summer flounder scales were prepared for age and growth analysis by making acetate impressions of the scale microstructure. Due to extreme variation in the size and shape of scales from individual fish, we selected only those scales that had even margins and which were of uniform size. We selected a range of four to six preferred scales (based on overall scale size) from each fish, making sure that only non-regenerated scales were used. Scale impressions were made on extruded clear acetate sheets (25 mm x 75 mm) with a Carver Laboratory Heated Press (model "C"). The scales were pressed with the following settings:

Pressure: 15000 psi
Temperature: 77 °C (170 °F)
Time: 5 to 10 min

summer flounder scales that were the size of a quarter (coin) or larger, were pressed individually for up to twenty minutes. After pressing, the impressions were viewed with a Bell and Howell microfiche reader and checked again for regeneration and incomplete margins. Impressions that were too light, or when all scales were regenerated a new impression was made using different scales from the same fish.

Click here to obtain the protocol at the CQFE website on how to prepare scale impression for ageing summer flounder.

Otoliths

We used our thin-section and bake technique to process spadefish sagittal otoliths (hereafter, referred to as "otoliths") for age determination. Otolith preparation began by randomly selecting either the right or left otolith. Each whole otolith was placed in a ceramic "Coors" spot plate well and baked in a Thermolyne 1400 furnace at 400 °C. Baking time was dependent on the otolith's size and gauged by color, with a light caramel color desired. Once a suitable color was achieved the baked otolith was embedded in epoxy resin with its distal surface orientated downwards and allowed to harden overnight. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet™ low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The otolith was positioned so the blades straddled each side of the otolith focus. It was crucial that this cut be perpendicular to the long axis of the otolith. Failure to do so resulted in broadening and distorted winter growth zones. A proper cut resulted in annuli that were clearly defined and delineated. Once cut, thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing summer flounder.

12.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of reading the information contained in its otolith, the date of its capture, and the species-specific period when it deposits its annulus. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called annuli. Technically, an otolith annulus is the combination of both the opaque and the translucent bands. In practice, only the opaque bands are counted as annuli. The number of these visible dark bands replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the otolith section is examined for translucent growth. If no translucent growth is visible beyond the last dark annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last dark annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the dark band of the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of dark bands in the otolith. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday", but before the dark band deposition period, is interpreted as being toward the next age class.

For example, summer flounder otolith deposition occurs between January and April (Bolz et al. 2000). A summer flounder captured between January 1 and April 30, before the end of the species' annulus formation period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of June 30, the period of annulus formation, which would be noted as 4+4.

Summer flounder scales are also considered to have a deposition between January and June (Bolz et al. 1999 and modified by CQFE), and age class assignment using these hard-parts is conducted in the same way as otoliths.

All summer flounder samples (scale pressings and sectioned otoliths) were aged by two different readers in chronological order based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish again without any knowledge of previously estimated ages or lengths, then assigned a final age to the fish. When the age readers were unable to agree on a final age, the fish was excluded from further analysis.

Scales

We determined fish age by viewing acetate impressions of scales (Figure 12.1) with a standard Bell and Howell R-735 microfiche reader equipped with 20 and 29 mm lenses. Annuli on summer flounder scales are identified based on two scale microstructure features, "crossing over" and circuli disruption. Primarily, "crossing over" in the lateral margins near the posterior/anterior interface of the scale is used to determine the origin of the annulus. Here compressed circuli (annulus) "cross-over" the previously deposited circuli of the previous

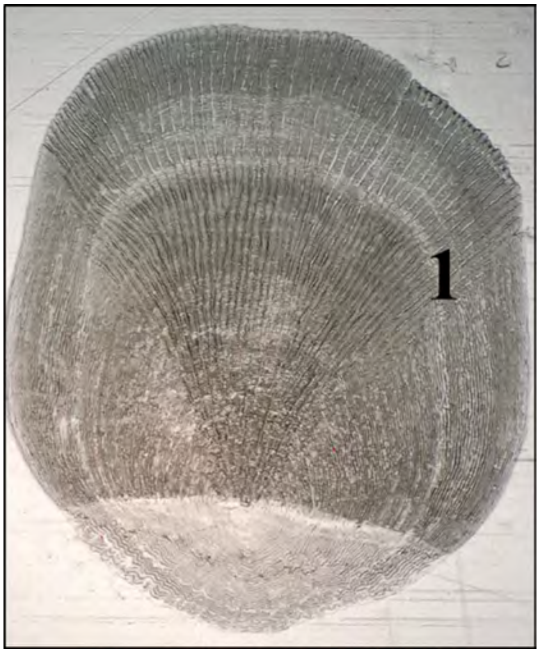


Figure 12.1: Scale impression of a 1 year-old summer flounder

year's growth. Typically annuli of the first three years can be observed transversing this interface as dark bands. These bands remain consistent throughout the posterior field and rejoin the posterior/anterior interface on the opposite side of the focus. Annuli can also be observed in the anterior lateral field of the scale. Here the annuli typically reveal a pattern of discontinuous and suddenly breaking segmented circuli. This event can also be distinguished by the presence of concentric white lines, which are typically associated with the disruption of circuli.

Annuli can also be observed bisecting the perpendicular plain of the radial striations in the anterior field of the scale. Radii emanate out from the focus of the scale towards the outer corner margins of the anterior field. These radial striations consist mainly of segmented concave circuli. The point of intersection between radii and annuli results in a "straightening out" of the concave circuli. This straightening of the circuli should be consistent throughout the entire anterior field of the scale. This event is further amplified by the presence of concave circuli neighboring both directly above and below the annulus. The first year's annulus can be difficult to locate on some scales. It is typically best identified in the lateral field of the anterior portion of the scale. The distance from the focus to the first year's annulus is typically larger with respect to the following annuli. For the annuli two through six, summer growth generally decreases proportionally. For ages greater than six, a crowding effect of the annuli near the outer margins of the scale is observed. This crowding effect creates difficulties in edge interpretation. At this point it is best to focus on the straightening of the circuli at the anterior margins of the scale.

When ageing young summer flounder, zero through age two, extreme caution must be taken as not to over age the structure. In young fish there is no point of reference to aid in the determination of the first year; this invariably results in over examination of the scale and such events as hatching or saltwater incursion marks (checks) may be interpreted as the first year.

Otoliths

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 12.2). Each reader aged all of the otolith samples. By conven-

Figure 12.2: Otolith thin-section of a 4 year-old summer flounder with the last annulus on the edge of the thin-section

tion an annulus is identified as the narrow opaque zone, or winter growth. Typically the first year's annulus can be determined by first locating the focus of the otolith. The focus is generally located, depending on preparation, in the center of the otolith, and is visually well defined as a dark oblong region. The first year's annulus can be located directly below the focus, along the outer ridge of the sulcal groove on the ventral and dorsal sides of the otolith. This insertion point along the sulcal ridge resembles a check mark (not to be confused with a false annulus). Here the annulus can be followed outwards along the ventral and dorsal surfaces where it encircles the focus. Subsequent annuli also emanate from the sulcal ridge; however, they do not encircle the focus, but rather travel outwards to the distal surface of the otolith. To be considered a true annulus, each annulus must be rooted in the sulcus and travel without interruption to the distal surface of the otolith. The annuli in summer flounder have a tendency to split as they advance towards the distal surface. As a result, it is critical that reading path proceed in a direction down the sulcal ridge and outwards to the distal surface.

Click here to obtain the protocol at the CQFE website on how to age summer flounder using their otolith thin-sections.

12.2.5 Comparison Tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (*CV*) analysis were used to detect any systematic difference and precision on age readings, respectively, for following comparisons: 1) between the two readers in the current year; 2) within each reader in the current year; 3) time-series bias between the current and previous years within each reader; and 4) between scale and otoliths ages. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2000 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

12.3 RESULTS

12.3.1 Sample size

We estimated a sample size of 381 bay summer flounder in 2015, ranging in length interval from 12 to 29 inches (Table 12.1). This sample size provided a range in *CV* for age composition approximately from the smallest *CV* of 6% for age 2 to the largest *CV* of 20% for age 6 of the bay fish. We randomly selected and aged 371 fish from 476 summer flounder collected by VMRC in Chesapeake Bay in 2015. We fell short in our over-all collections for this optimal length-class sampling estimate by 37 fish. We were short of few fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

We estimated a sample size of 433 ocean summer flounder in 2015, ranging in length interval from 11 to 33 inches (Table 12.2). This sample size provided a range in *CV* for age composition approximately from the smallest *CV* of 8% for age 3 to the largest *CV* of 22% for age 8 of the ocean fish. We randomly selected and aged 513 fish from 615 summer flounder collected by VMRC in Virginia waters of the Atlantic Ocean in 2015. We fell short in our over-all collections for this optimal length-class sampling estimate by 32 fish. However, we were short of no fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

12.3.2 Scales

Both readers had moderate self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 64% (1 year or less agreement of 96%) and a *CV* of 5.9% (test of symmetry: $\chi^2 = 9.8$, $df = 9$, $P = 0.3669$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 64% (1 year or less agreement of 92%) and a *CV* of 8.2% (test of symmetry: $\chi^2 = 12.67$, $df = 10$, $P = 0.2429$). There was an evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 63% (1 year or less agreement of 91%) and a *CV* of 8.2% (test of symmetry: $\chi^2 = 89.98$, $df = 32$, $P < 0.0001$) (Figure 12.3).

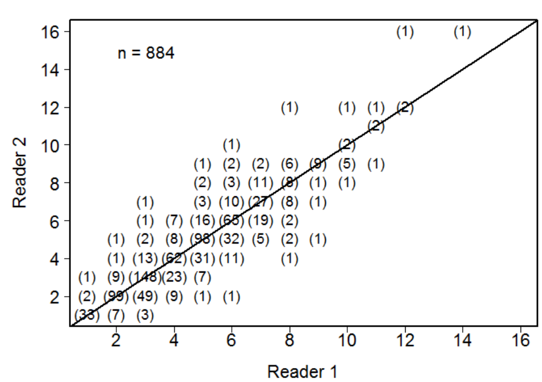


Figure 12.3: Between-reader comparison of scale age estimates for summer flounder collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

There was no time-series bias for both readers. Reader 1 had an agreement of 84% (1 year or less agreement of 100%) with ages of fish aged in 2000 with a *CV* of 3% (test of symmetry: $\chi^2 = 2.67$, $df = 3$, $P = 0.4459$). Reader 2 had an agreement of 68% (1 year or less agreement of 88%) with a *CV* of 12.8% (test of symmetry: $\chi^2 = 16$, $df = 9$, $P = 0.0669$).

Of the 371 bay summer flounder aged with scales, 10 age classes (1 to 10) were represented (Table 12.3). The average age for the sample was 3.7 years. The standard deviation and standard error were 1.8 and 0.09, respectively. Year-class data (Figure 12.4) indicates that recruitment into the fishery in Chesapeake Bay begins at age 1, which corresponds to the 2014 year-class for summer flounder caught in 2015. Summer flounder in the sample in 2015 was dominated by the year class of 2012 with 33%. The sex ratio of male to female was 1:48 for the bay fish.

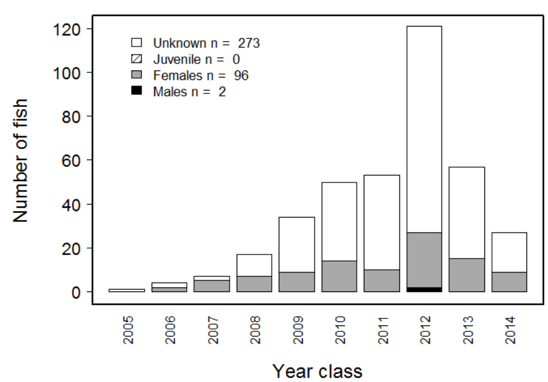


Figure 12.4: Year-class frequency distribution for summer flounder collected in Chesapeake Bay, Virginia for ageing in 2015. Distribution is broken down by sex and estimated using scale ages. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

Of the 513 ocean summer flounder aged with scales, 13 age classes (1 to 12, and 15) were represented (Table 12.4). The average age for the sample was 4.9 years. The standard deviation and standard error were 2 and 0.09, respectively. Year-class data (Figure 12.5) indicates that recruitment into the fishery in Virginia waters of Atlantic ocean begins at age 1, which corresponds to the 2014 year-class for summer flounder caught in 2015. Summer flounder in the sample in 2015 was dominated by the year classes of 2010 and 2009 with 23% and 18%, respectively. The sex ratio of male to female was 1:1.15 for the ocean fish.

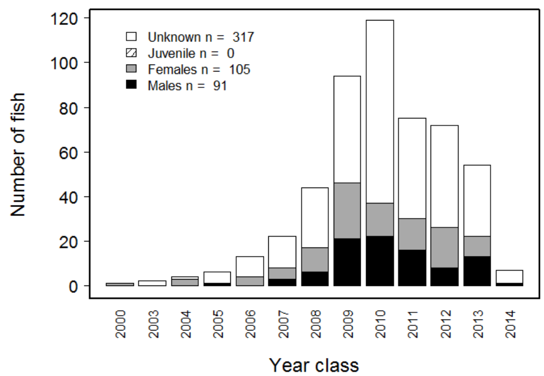


Figure 12.5: Year-class frequency distribution for summer flounder collected in Virginia waters of the Atlantic Ocean for ageing in 2015. Distribution is broken down by sex and estimated using scale ages. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

12.3.3 Otoliths

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 90% and a *CV* of 1.6% (test of symmetry: $\chi^2 = 5$, $df = 5$, $P = 0.4159$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 88% and a *CV* of 1.6% (test of symmetry: $\chi^2 = 6$, $df = 5$, $P = 0.3062$). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 88% (1 year or less agreement of 99%) and a *CV* of 1.8% (test of symmetry: $\chi^2 = 14.14$, $df = 13$, $P = 0.3639$) (Figure 12.6).

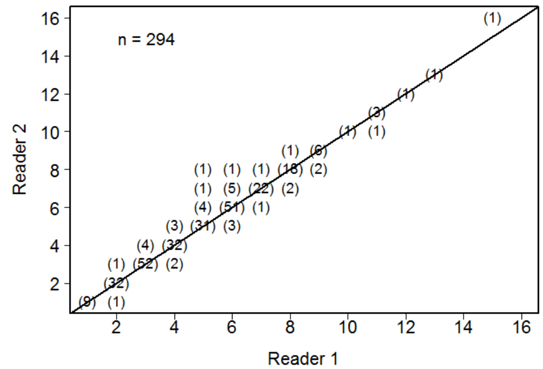


Figure 12.6: Between-reader comparison of otolith age estimates for summer flounder collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

There was no time-series bias for both readers. Reader 1 had an agreement of 92% with ages of fish aged in 2003 with a *CV* of 1.9% (test of symmetry: $\chi^2 = 4$, $df = 4$, $P = 0.406$). Reader 2 had an agreement of 94% with a *CV* of 0.8% (test of symmetry: $\chi^2 = 3$, $df = 3$, $P = 0.3916$).

Of the 294 summer flounder aged with otoliths, 14 age classes (1 to 13, and 15) were represented (Table 12.5). The average age for the sample was 4.9 years. The standard deviation and standard error were 2.3 and 0.13, respectively.

12.3.4 Comparison of scale and otolith ages

We aged 293 summer flounder using scales and otoliths (Excluding 1 fish with otolith-age only). There was an evidence of systematic disagreement between otolith and scale ages (test of symmetry: $\chi^2 = 33.91$, $df = 20$, $P = 0.0267$) with an average *CV* of 7.3%. There was an agreement of 61% between scale and otoliths ages whereas scales were assigned a lower and higher age than otoliths for 28% and 11% of the fish, respectively (Figure 12.7). There was also an evidence of bias between otolith and scale ages using an age bias plot(Figure 12.8), with scale generally assigned higher ages for younger fish and lower ages for older fish than otolith age estimates.

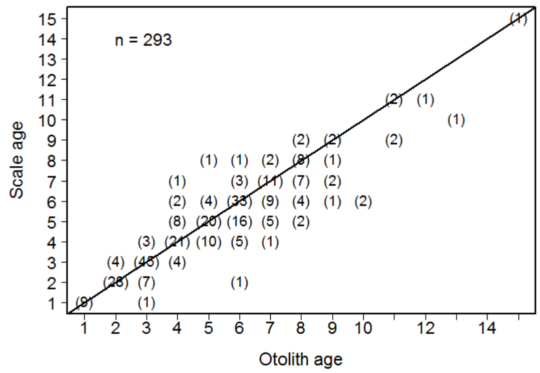


Figure 12.7: Comparison of scale and otolith age estimates for summer flounder collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

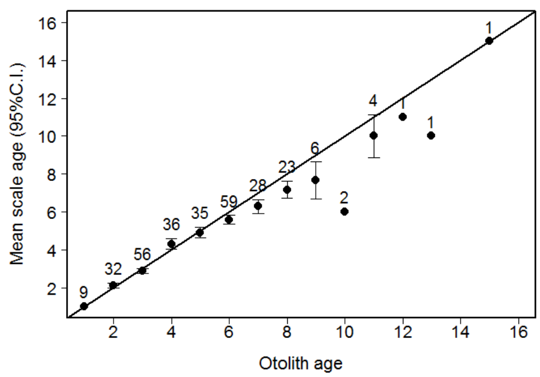


Figure 12.8: Age-bias plot for summer flounder scale and otolith age estimates in 2015.

12.3.5 Age-Length-Key (ALK)

We developed an age-length-key for both bay (Table 12.6) and ocean fish (Table 12.7) using scale ages, separately. The ALK can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using scale ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

12.4 REFERENCES

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Table 12.1: Number of bay summer flounder collected and aged in each 1-inch length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
12 - 12.99	5	0	0	5
13 - 13.99	6	0	0	6
14 - 14.99	70	86	75	0
15 - 15.99	58	70	60	0
16 - 16.99	45	64	45	0
17 - 17.99	39	73	39	0
18 - 18.99	31	53	36	0
19 - 19.99	28	45	31	0
20 - 20.99	21	29	29	0
21 - 21.99	22	19	19	3
22 - 22.99	15	14	14	1
23 - 23.99	11	9	9	2
24 - 24.99	5	9	9	0
25 - 25.99	5	2	2	3
26 - 26.99	5	2	2	3
27 - 27.99	5	0	0	5
28 - 28.99	5	1	1	4
29 - 29.99	5	0	0	5
Totals	381	476	371	37

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Table 12.2: Number of ocean summer flounder collected and aged in each 1-inch length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
11 - 11.99	5	0	0	5
12 - 12.99	5	0	0	5
13 - 13.99	5	1	1	4
14 - 14.99	34	54	43	0
15 - 15.99	56	97	72	0
16 - 16.99	58	81	62	0
17 - 17.99	47	71	64	0
18 - 18.99	34	45	45	0
19 - 19.99	24	26	26	0
20 - 20.99	23	32	29	0
21 - 21.99	17	30	19	0
22 - 22.99	21	37	27	0
23 - 23.99	21	35	30	0
24 - 24.99	17	30	26	0
25 - 25.99	14	25	18	0
26 - 26.99	11	23	23	0
27 - 27.99	10	13	13	0
28 - 28.99	6	8	8	0
29 - 29.99	5	4	4	1
30 - 30.99	5	2	2	3
31 - 31.99	5	0	0	5
32 - 32.99	5	1	1	4
33 - 33.99	5	0	0	5
Totals	433	615	513	32

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Table 12.3: The number of summer flounder assigned to each total length-at-age category for 371 fish sampled for scale age determination in Chesapeake Bay, Virginia during 2015.

	Age										
Interval	1	2	3	4	5	6	7	8	9	10	Totals
14 - 14.99	24	22	20	7	1	1	0	0	0	0	75
15 - 15.99	3	20	28	7	2	0	0	0	0	0	60
16 - 16.99	0	7	28	2	5	2	1	0	0	0	45
17 - 17.99	0	5	16	6	8	3	1	0	0	0	39
18 - 18.99	0	1	12	8	9	3	2	1	0	0	36
19 - 19.99	0	1	10	7	6	4	3	0	0	0	31
20 - 20.99	0	1	4	10	8	4	1	0	1	0	29
21 - 21.99	0	0	3	2	4	5	3	1	1	0	19
22 - 22.99	0	0	0	0	5	6	2	1	0	0	14
23 - 23.99	0	0	0	3	2	3	1	0	0	0	9
24 - 24.99	0	0	0	1	0	2	2	3	1	0	9
25 - 25.99	0	0	0	0	0	0	0	1	1	0	2
26 - 26.99	0	0	0	0	0	1	1	0	0	0	2
28 - 28.99	0	0	0	0	0	0	0	0	0	1	1
Totals	27	57	121	53	50	34	17	7	4	1	371

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Table 12.4: The number of summer flounder assigned to each total length-at-age category for 513 fish sampled for scale age determination in Virginia waters of Atlantic ocean during 2015.

	Age													
Interval	1	2	3	4	5	6	7	8	9	10	11	12	15	Totals
13 - 13.99	0	1	0	0	0	0	0	0	0	0	0	0	0	1
14 - 14.99	5	14	10	4	7	3	0	0	0	0	0	0	0	43
15 - 15.99	1	21	18	13	7	8	3	1	0	0	0	0	0	72
16 - 16.99	1	6	14	16	17	5	2	0	0	1	0	0	0	62
17 - 17.99	0	6	12	13	18	10	3	1	0	0	0	1	0	64
18 - 18.99	0	3	5	6	14	12	4	1	0	0	0	0	0	45
19 - 19.99	0	3	6	4	7	2	1	2	0	1	0	0	0	26
20 - 20.99	0	0	6	4	9	4	2	1	2	1	0	0	0	29
21 - 21.99	0	0	0	3	9	2	4	1	0	0	0	0	0	19
22 - 22.99	0	0	0	6	7	9	3	1	0	1	0	0	0	27
23 - 23.99	0	0	1	4	7	8	5	2	2	1	0	0	0	30
24 - 24.99	0	0	0	1	5	12	3	1	3	0	1	0	0	26
25 - 25.99	0	0	0	0	6	6	4	2	0	0	0	0	0	18
26 - 26.99	0	0	0	0	5	8	5	2	3	0	0	0	0	23
27 - 27.99	0	0	0	1	0	3	4	3	1	0	1	0	0	13
28 - 28.99	0	0	0	0	1	1	0	2	1	1	1	0	1	8
29 - 29.99	0	0	0	0	0	1	1	2	0	0	0	0	0	4
30 - 30.99	0	0	0	0	0	0	0	0	1	0	0	1	0	2
32 - 32.99	0	0	0	0	0	0	0	0	0	0	1	0	0	1
Totals	7	54	72	75	119	94	44	22	13	6	4	2	1	513

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Table 12.5: The number of summer flounder assigned to each total length-at-age category for 294 fish sampled for otolith age determination in Chesapeake Bay and Virginia waters of Atlantic Ocean during 2015.

	Age														
Interval	1	2	3	4	5	6	7	8	9	10	11	12	13	15	Totals
14 - 14.99	8	8	5	2	1	0	0	1	0	0	0	0	0	0	25
15 - 15.99	1	13	12	4	1	5	2	2	1	1	0	0	0	0	42
16 - 16.99	0	5	12	8	5	4	3	0	0	0	0	0	1	0	38
17 - 17.99	0	5	11	4	7	4	4	0	0	0	0	0	0	0	35
18 - 18.99	0	0	5	4	2	8	3	2	1	1	0	0	0	0	26
19 - 19.99	0	1	8	1	0	2	1	2	0	0	0	0	0	0	15
20 - 20.99	0	0	3	4	6	3	1	0	0	0	0	0	0	0	17
21 - 21.99	0	0	1	2	3	3	5	0	1	0	0	0	0	0	15
22 - 22.99	0	0	0	2	5	7	1	2	0	0	0	0	0	0	17
23 - 23.99	0	0	0	3	3	7	2	0	0	0	0	0	0	0	15
24 - 24.99	0	0	0	2	2	5	2	2	1	0	1	0	0	0	15
25 - 25.99	0	0	0	0	0	4	0	3	1	0	0	0	0	0	8
26 - 26.99	0	0	0	0	0	4	2	2	0	0	0	0	0	0	8
27 - 27.99	0	0	0	0	0	2	2	3	0	0	0	1	0	0	8
28 - 28.99	0	0	0	0	0	1	0	2	1	0	2	0	0	1	7
29 - 29.99	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1
30 - 30.99	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1
32 - 32.99	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1
Totals	9	32	57	36	35	59	28	23	6	2	4	1	1	1	294

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Table 12.6: Age-Length key, as proportion-at-age in each 1-inch length interval, based on scale ages for summer flounder sampled in Chesapeake Bay, Virginia during 2015.

	Age									
Interval	1	2	3	4	5	6	7	8	9	10
14 - 14.99	0.32	0.29	0.27	0.09	0.01	0.01	0	0	0	0
15 - 15.99	0.05	0.33	0.47	0.12	0.03	0	0	0	0	0
16 - 16.99	0	0.16	0.62	0.04	0.11	0.04	0.02	0	0	0
17 - 17.99	0	0.13	0.41	0.15	0.21	0.08	0.03	0	0	0
18 - 18.99	0	0.03	0.33	0.22	0.25	0.08	0.06	0.03	0	0
19 - 19.99	0	0.03	0.32	0.23	0.19	0.13	0.1	0	0	0
20 - 20.99	0	0.03	0.14	0.34	0.28	0.14	0.03	0	0.03	0
21 - 21.99	0	0	0.16	0.11	0.21	0.26	0.16	0.05	0.05	0
22 - 22.99	0	0	0	0	0.36	0.43	0.14	0.07	0	0
23 - 23.99	0	0	0	0.33	0.22	0.33	0.11	0	0	0
24 - 24.99	0	0	0	0.11	0	0.22	0.22	0.33	0.11	0
25 - 25.99	0	0	0	0	0	0	0	0.5	0.5	0
26 - 26.99	0	0	0	0	0	0.5	0.5	0	0	0
28 - 28.99	0	0	0	0	0	0	0	0	0	1

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Table 12.7: Age-Length key, as proportion-at-age in each 1-inch length interval, based on scale ages for summer flounder sampled in Virginia waters of the Atlantic Ocean during 2015.

	Age												
Interval	1	2	3	4	5	6	7	8	9	10	11	12	15
13 - 13.99	0	1	0	0	0	0	0	0	0	0	0	0	0
14 - 14.99	0.12	0.33	0.23	0.09	0.16	0.07	0	0	0	0	0	0	0
15 - 15.99	0.01	0.29	0.25	0.18	0.1	0.11	0.04	0.01	0	0	0	0	0
16 - 16.99	0.02	0.1	0.23	0.26	0.27	0.08	0.03	0	0	0.02	0	0	0
17 - 17.99	0	0.09	0.19	0.2	0.28	0.16	0.05	0.02	0	0	0	0.02	0
18 - 18.99	0	0.07	0.11	0.13	0.31	0.27	0.09	0.02	0	0	0	0	0
19 - 19.99	0	0.12	0.23	0.15	0.27	0.08	0.04	0.08	0	0.04	0	0	0
20 - 20.99	0	0	0.21	0.14	0.31	0.14	0.07	0.03	0.07	0.03	0	0	0
21 - 21.99	0	0	0	0.16	0.47	0.11	0.21	0.05	0	0	0	0	0
22 - 22.99	0	0	0	0.22	0.26	0.33	0.11	0.04	0	0.04	0	0	0
23 - 23.99	0	0	0.03	0.13	0.23	0.27	0.17	0.07	0.07	0.03	0	0	0
24 - 24.99	0	0	0	0.04	0.19	0.46	0.12	0.04	0.12	0	0.04	0	0
25 - 25.99	0	0	0	0	0.33	0.33	0.22	0.11	0	0	0	0	0
26 - 26.99	0	0	0	0	0.22	0.35	0.22	0.09	0.13	0	0	0	0
27 - 27.99	0	0	0	0.08	0	0.23	0.31	0.23	0.08	0	0.08	0	0
28 - 28.99	0	0	0	0	0.12	0.12	0	0.25	0.12	0.12	0.12	0	0.12
29 - 29.99	0	0	0	0	0	0.25	0.25	0.5	0	0	0	0	0
30 - 30.99	0	0	0	0	0	0	0	0	0.5	0	0	0.5	0
32 - 32.99	0	0	0	0	0	0	0	0	0	0	1	0	0

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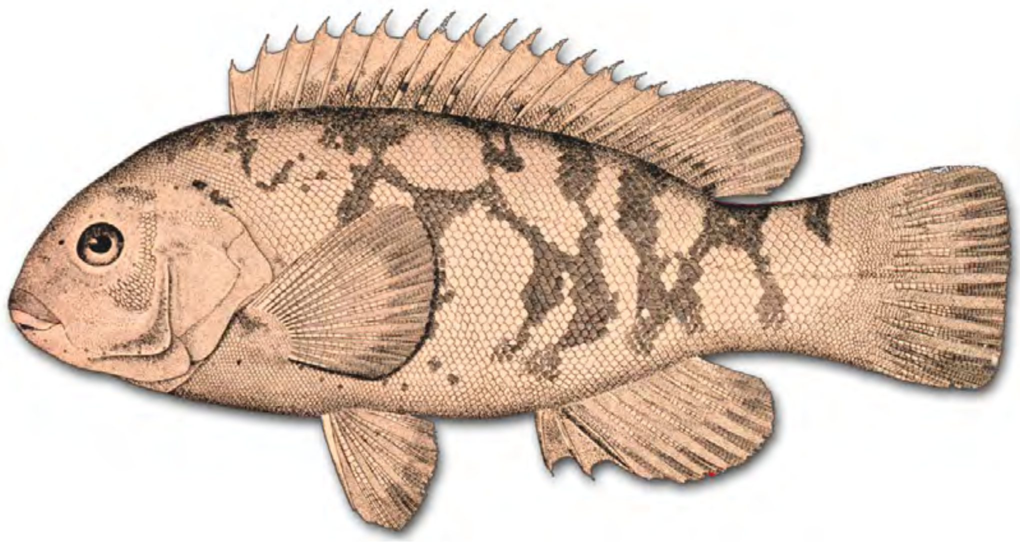
Chapter 13

TAUTOG *Tautoga onitis*



13.1 INTRODUCTION

We aged a total of 277 tautog, *Tautoga onitis*, using their opercula collected by the VMRC's Biological Sampling Program in 2015. Of 277 aged fish, 220 and 57 fish were collected in Chesapeake Bay (bay fish) and Virginia waters of the Atlantic Ocean (ocean fish), respectively. The average age for the bay fish was 6.1 years with a standard deviation of 1.6 and a standard error of 0.11. Eleven age classes (3 to 10, 13 to 14, and 18) were represented in the bay fish, comprising fish from the 1997, 2001 to 2002, and 2005 to 2012 year classes. The bay fish sample in 2015 was dominated by the year class of 2009 with 46%. The average age for the ocean fish was 10.4 years with a standard deviation of 6.7 and a standard error of 0.89. Eighteen age classes (4 to 10, 12 to 13, 15, 17 to 18, 20, 22 to 24, 27, and 31) were represented in the ocean fish, comprising fish from the 1984, 1988, 1991 to 1993, 1995, 1997 to 1998, 2000, 2002 to 2003, and 2005 to 2011 year classes. The ocean fish sample in 2015 was dominated by the year class of 2009 with 25%. We also aged a total of 273 fish using their otoliths in addition to ageing their opercula. The otolith ages were compared to the operculum ages to examine how close both ages were to one another (see details in Results).

13.2 METHODS

13.2.1 Sample size for ageing

We estimated sample sizes for ageing tautog collected in both Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015, respectively, using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 CV^2 + B_a/L} \tag{13.1}$$

where A is the sample size for ageing tautog in 2015; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of tautog used by VMRC to estimate length distribution of the catches from 2009 to 2013. θ_a, V_a, B_a, and CV were calculated using pooled age-length data of tautog collected from 2009 to 2013 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (13.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2009 to 2013 catch. A_l is number of fish to be aged for length interval l in 2015.

13.2.2 Handling of collection

Sagittal otoliths (hereafter, referred to as "otoliths") and opercula were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted based on date of capture. Their envelope labels were verified against VMRC's collection data, and each fish assigned a unique Age and Growth Laboratory identification number. All otoliths and opercula were stored dry within their original labeled coin envelopes; otoliths were contained inside protective Axygen 2.0 ml microtubes.

13.2.3 Preparation

Opercula

Tautog opercula were boiled for several minutes to remove any attached skin and connective tissue. After boiling, opercula were inspected for damage. If there were no obvious flaws, the opercula was dried and then stored in a new, labeled envelope.

Click here to obtain the protocol at the CQFE website on how to prepare operculum for ageing tautog.

Otoliths

We used our thin-section and bake technique to process spadefish sagittal otoliths (hereafter, referred to as "otoliths") for age determination. Otolith preparation began by randomly selecting either the right or left otolith. Each whole otolith was placed in a ceramic "Coors" spot plate well and baked in a Thermolyne 1400 furnace at 400 °C. Baking time was dependent on the otolith's size and gauged by color, with a light caramel color desired. Once a suitable color was achieved the baked otolith was embedded in epoxy resin with its distal surface orientated downwards and allowed to harden overnight. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a permanent marker across the epoxy resin surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet™ low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). The otolith was positioned so the blades straddled each side of the otolith focus. It was crucial that this cut be perpendicular to the long axis of the otolith. Failure to do so resulted in broadening and distorted winter growth zones. A proper cut resulted in annuli that were clearly defined and delineated. Once cut, thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-section.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing tautog.

13.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of reading the information contained in its otolith, the date of its capture, and the species-specific period when it deposits its annulus. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called annuli. Technically, an otolith annulus is the combination of both the opaque and the translucent bands. In practice, only the opaque bands are counted as annuli. The number of these visible dark bands replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the otolith section is examined for translucent growth. If no translucent growth is visible beyond the last dark annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last dark annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the dark band of the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of dark bands in the otolith. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday", but before the dark band deposition period, is interpreted as being toward the next age class.

For example, tautog otolith deposition occurs between May and July (Hostetter and Munroe 1993). A tautog captured between January 1 and July 31, before the end of the species' annulus formation period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of June 30, the period of annulus formation, which would be noted as 4+4.

Tautog opercula are also considered to have a deposition period of May through July (Hostetter and Munroe 1993), and age class assignment using these hard-parts is conducted in the same way as otoliths.

All tautog samples (opercula and sectioned otoliths) were aged by two different readers in chronological order based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish again without any knowledge of previously estimated ages or lengths, then assigned a final age to the fish. When the age readers were unable to agree on a final age, the fish was excluded from further analysis.

Opercula

All prepared opercula were aged by two different readers in chronological order based on collection date, without knowledge of previously estimated ages or the specimen lengths. Opercula were aged on a light table with no magnification (Figure 13.1).

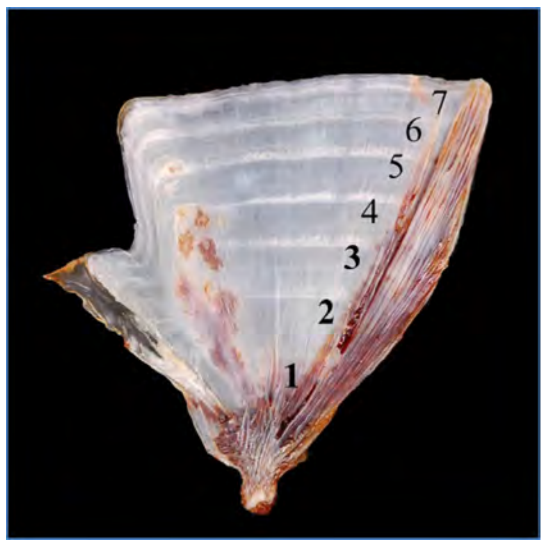


Figure 13.1: Operculum of a 7 year-old tautog

Otoliths

All otolith thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 13.2). Each reader aged all of the otolith samples.

Click here to obtain the protocol at the CQFE website on how to age tautog using their otolith thin-sections.

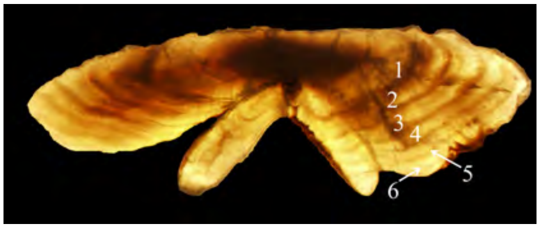


Figure 13.2: Otolith thin-section of 6 year-old tautog

13.2.5 Comparison Tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for following comparisons: 1) between the two readers in the current year; 2) within each reader in the current year; 3) time-series bias between the current and previous years within each reader; and 4) between operculum and otoliths ages. The readings from the entire sample for the current year were used to examine the difference between two readers. A random sub-sample of 50 fish from the current year was selected for second readings to examine the difference within a reader.Fifty otoliths randomly selected from fish aged in 2000 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

13.3 RESULTS

13.3.1 Sample size

We estimated a sample size of 414 bay tautog in 2015, ranging in length interval from 12 to 27 inches (Table 13.1). This sample size provided a range in CV for age composition approximately from the smallest CV of 1% for age 16 to the largest CV of 22% for age 2 of the bay fish. We aged all 220 tautog collected by VMRC in Chesapeake Bay in 2015. We fell short in our overall collections for this optimal length-class sampling estimate by 194 fish. However, we were short of many fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would definitely be influenced significantly. Therefore, precaution should be used when developing ALK using these age data.

We estimated a sample size of 398 ocean tautog in 2015, ranging in length interval from 11 to 30 inches (Table 13.2). This sample size provided a range in CV for age composition approximately from the smallest CV of 10% for age 4 to the largest CV of 21% for age 13 of the ocean fish. We aged all 57 tautog collected by VMRC in Virginia waters of the Atlantic Ocean in 2015. We fell short in our over-all collections for this optimal length-class sampling estimate by 342 fish. However, we were short of many fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would definitely be influenced significantly. Therefore, precaution should be used when developing ALK using these age data.

13.3.2 Opercula

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 74% (1 year or less agreement of 96%) and a CV of 3.1% (test of symmetry: χ^2 = 5.8, df = 6, P = 0.446), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 64% (1 year or less agreement of 94%) and a CV of 4.5% (test of symmetry: $\chi^2 = 8.33$, df = 10, P = 0.5963). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 70% (1 year or less agreement of 97%) and a *CV* of 3.5% (test of symmetry: $\chi^2 = 26.73$, $df = 20$, $P = 0.143$) (Figure 13.3).

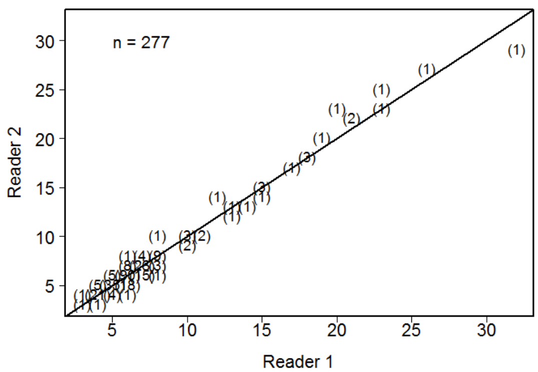


Figure 13.3: Between-reader comparison of operculum age estimates for tautog collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

There was no time-series bias for both readers. Reader 1 had an agreement of 68% (1 year or less agreement of 90%) with ages of fish aged in 2000 with a *CV* of 5.3% (test of symmetry: $\chi^2 = 6$, $df = 10$, $P = 0.8153$). Reader 2 had an agreement of 80% (1 year or less agreement of 98%) with a *CV* of 2.9% (test of symmetry: $\chi^2 = 6$, $df = 6$, $P = 0.4232$).

Of the 220 bay tautog aged with opercula, 11 age classes (3 to 10, 13 to 14, and 18) were represented (Table 13.3). The average age for the sample was 6.1 years. The standard deviation and standard error were 1.6 and 0.11, respectively. Year-class data (Figure 13.4) indicates that recruitment into the fishery in Chesapeake Bay begins at age 3, which corresponds to the 2012 year-class for tautog caught in 2015. Tautog in the sample in 2015 was dominated by the year class of 2009 with 46%. The sex ratio of male to female was 1:0.93 for the bay fish.

Of the 57 ocean tautog aged with opercula,

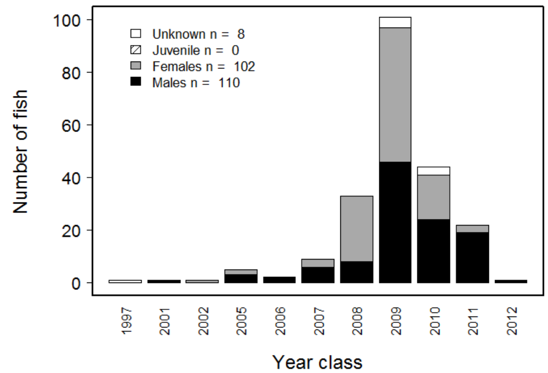


Figure 13.4: Year-class frequency distribution for tautog collected in Chesapeake Bay, Virginia for ageing in 2015. Distribution is broken down by sex and estimated using operculum ages. 'Unknown' represents the fish gonads that were not available for examination or were not examined for sex during sampling.

18 age classes (4 to 10, 12 to 13, 15, 17 to 18, 20, 22 to 24, 27, and 31) were represented (Table 13.4). The average age for the sample was 10.4 years. The standard deviation and standard error were 6.7 and 0.89, respectively. Year-class data (Figure 13.5) indicates that recruitment into the fishery in Virginia waters of Atlantic ocean begins at age 4, which corresponds to the 2011 year-class for tautog caught in 2015. Tautog in the sample in 2015 was dominated by the year class of 2009 with 25%. The sex ratio of male to female was 1:1.71 for the ocean fish.

13.3.3 Otoliths

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 90% and a *CV* of 0.8% (test of symmetry: $\chi^2 = 5$, $df = 5$, $P = 0.4159$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 88% and a *CV* of 1.2% (test of symmetry: $\chi^2 = 3.33$, $df = 4$, $P = 0.5037$). There was no evidence of systematic disagreement

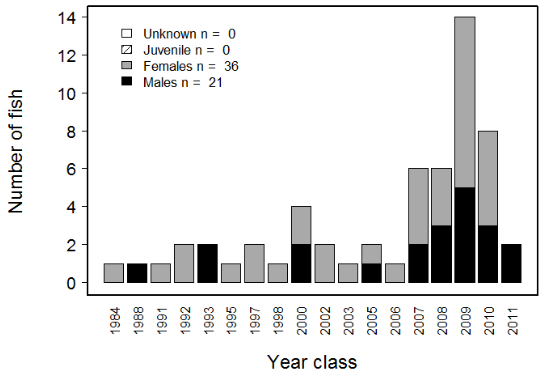


Figure 13.5: Year-class frequency distribution for tautog collected in Virginia waters of the Atlantic Ocean for ageing in 2015. Distribution is broken down by sex and estimated using operculum ages. 'Unknown' represents the fish gonads that were not available for examination or were not examined for sex during sampling.

between Reader 1 and Reader 2 with an agreement of 88% (1 year or less agreement of 97%) and a *CV* of 1.2% (test of symmetry: χ^2 = 18.2, *df* = 20, *P* = 0.5742) (Figure 13.6). There was no time-series bias

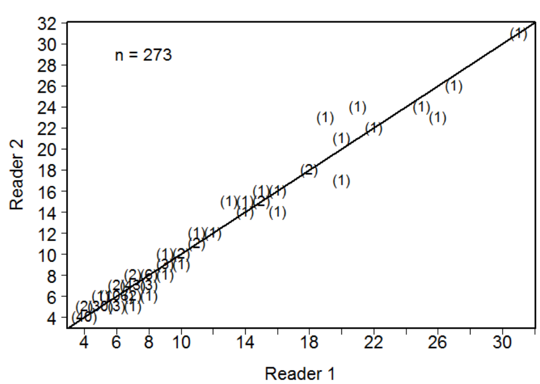


Figure 13.6: Between-reader comparison of otolith age estimates for tautog collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

for both readers. Reader 1 had an agreement of 94% with ages of fish aged in 2003 with a *CV* of 0.6% (test of symmetry: χ^2 = 3, *df* = 2, *P* = 0.2231). Reader 2 had an agreement of 94% with a *CV* of 0.7% (test of symmetry: χ^2 = 3, *df* = 2, *P* = 0.2231).

Of the 273 tautog aged with otoliths, 19 age classes (4 to 16, 18 to 19, 22 to 23, 25, and 31) were represented (Table 13.5). The average age for the sample was 6.9 years. The standard deviation and standard error were 3.8 and 0.23, respectively.

13.3.4 Comparison of operculum and otolith ages

We aged 273 tautog using opercula and otoliths. There was no evidence of systematic disagreement between otolith and operculum ages (test of symmetry: χ^2 = 38.46, *df* = 28, *P* = 0.0899) with an average *CV* of 5%. There was an agreement of 61% between operculum and otoliths ages whereas opercula were assigned a lower and higher age than otoliths for 18% and 21% of the fish, respectively (Figure 13.7). There was also little evidence of bias between otolith and operculum ages using an age bias plot(Figure 13.8), with no trend of either over-ageing younger or under-ageing older fish.

Figure 13.7: Comparison of operculum and otolith age estimates for tautog collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

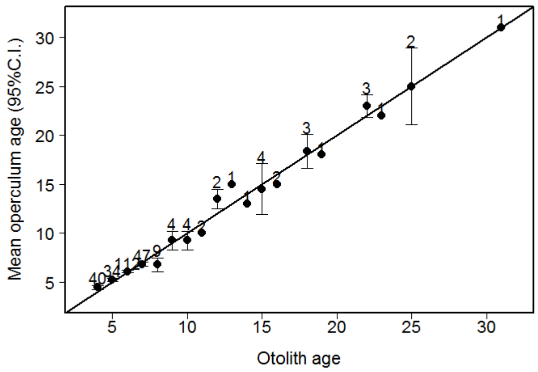


Figure 13.8: Age-bias plot for tautog operculum and otolith age estimates in 2015.

13.3.5 Age-Length-Key (ALK)

We developed an age-length-key for both bay (Table 13.6) and ocean fish (Table 13.7) using operculum ages, separately. The ALK can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using operculum ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

13.4 REFERENCES

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Table 13.1: Number of bay tautog collected and aged in each 1-inch length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
12 - 12.99	5	1	1	4
13 - 13.99	23	0	0	23
14 - 14.99	66	17	17	49
15 - 15.99	77	68	68	9
16 - 16.99	68	51	51	17
17 - 17.99	58	46	46	12
18 - 18.99	40	18	18	22
19 - 19.99	30	13	13	17
20 - 20.99	12	4	4	8
21 - 21.99	5	1	1	4
22 - 22.99	5	0	0	5
23 - 23.99	5	0	0	5
24 - 24.99	5	0	0	5
25 - 25.99	5	0	0	5
26 - 26.99	5	1	1	4
27 - 27.99	5	0	0	5
Totals	414	220	220	194

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Table 13.2: Number of ocean tautog collected and aged in each 1-inch length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
11 - 11.99	5	0	0	5
13 - 13.99	10	0	0	10
14 - 14.99	35	0	0	35
15 - 15.99	63	1	1	62
16 - 16.99	79	6	6	73
17 - 17.99	35	3	3	32
18 - 18.99	31	6	6	25
19 - 19.99	21	4	4	17
20 - 20.99	21	7	7	14
21 - 21.99	14	7	7	7
22 - 22.99	16	4	4	12
23 - 23.99	17	3	3	14
24 - 24.99	7	2	2	5
25 - 25.99	5	3	3	2
26 - 26.99	19	3	3	16
27 - 27.99	5	6	6	0
28 - 28.99	5	2	2	3
29 - 29.99	5	0	0	5
30 - 30.99	5	0	0	5
Totals	398	57	57	342

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Table 13.3: The number of tautog assigned to each total length-at-age category for 220 fish sampled for operculum age determination in Chesapeake Bay, Virginia during 2015.

	Age											
Interval	3	4	5	6	7	8	9	10	13	14	18	Totals
12 - 12.99	0	0	1	0	0	0	0	0	0	0	0	1
14 - 14.99	0	6	7	3	1	0	0	0	0	0	0	17
15 - 15.99	1	10	21	33	3	0	0	0	0	0	0	68
16 - 16.99	0	4	9	27	10	1	0	0	0	0	0	51
17 - 17.99	0	0	4	26	14	1	0	0	1	0	0	46
18 - 18.99	0	0	2	10	0	4	0	2	0	0	0	18
19 - 19.99	0	2	0	2	4	3	2	0	0	0	0	13
20 - 20.99	0	0	0	0	1	0	0	2	0	1	0	4
21 - 21.99	0	0	0	0	0	0	0	1	0	0	0	1
26 - 26.99	0	0	0	0	0	0	0	0	0	0	1	1
Totals	1	22	44	101	33	9	2	5	1	1	1	220

(Go back to text)

Table 13.4: The number of tautog assigned to each total length-at-age category for 57 fish sampled for operculum age determination in Virginia waters of Atlantic ocean during 2015.

	Age																	
Interval	4	5	6	7	8	9	10	12	13	15	17	18	20	22	23	24	27	31	Totals
15 - 15.99	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
16 - 16.99	1	4	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	6
17 - 17.99	0	0	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	3
18 - 18.99	1	1	2	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	6
19 - 19.99	0	0	3	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	4
20 - 20.99	0	2	3	2	0	0	0	0	0	0	0	0	0	0	0	0	0	0	7
21 - 21.99	0	0	3	1	2	0	1	0	0	0	0	0	0	0	0	0	0	0	7
22 - 22.99	0	0	2	0	2	0	0	0	0	0	0	0	0	0	0	0	0	0	4
23 - 23.99	0	0	0	0	0	0	1	1	0	1	0	0	0	0	0	0	0	0	3
24 - 24.99	0	0	0	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	2
25 - 25.99	0	0	0	0	0	0	0	0	0	2	0	1	0	0	0	0	0	0	3
26 - 26.99	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1	0	0	0	3
27 - 27.99	0	0	0	0	0	0	0	0	2	0	1	0	0	0	1	1	1	0	6
28 - 28.99	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	2
Totals	2	8	14	6	6	1	2	1	2	4	1	2	1	2	2	1	1	1	57

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Table 13.5: The number of tautog assigned to each total length-at-age category for 273 fish sampled for otolith age determination in Chesapeake Bay and Virginia waters of Atlantic Ocean during 2015.

	Age																		
Interval	4	5	6	7	8	9	10	11	12	13	14	15	16	18	19	22	23	25	31	Totals
12 - 12.99	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
14 - 14.99	8	5	4	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	17
15 - 15.99	18	17	27	5	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	67
16 - 16.99	7	7	35	7	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	57
17 - 17.99	2	4	27	13	0	0	1	0	0	0	0	1	0	0	0	0	0	0	0	48
18 - 18.99	2	0	12	8	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	24
19 - 19.99	2	0	6	4	2	1	1	0	0	0	0	0	0	0	0	0	0	0	0	16
20 - 20.99	0	1	1	5	1	0	1	1	1	0	0	0	0	0	0	0	0	0	0	11
21 - 21.99	0	0	0	3	3	1	0	1	0	0	0	0	0	0	0	0	0	0	0	8
22 - 22.99	0	0	0	2	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	4
23 - 23.99	0	0	0	0	0	0	1	0	0	0	0	1	1	0	0	0	0	0	0	3
24 - 24.99	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	1	0	0	2
25 - 25.99	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1	0	0	0	0	3
26 - 26.99	0	0	0	0	0	0	0	0	0	0	0	1	0	2	0	0	0	1	0	4
27 - 27.99	0	0	0	0	0	0	0	0	1	0	1	0	0	1	0	2	0	1	0	6
28 - 28.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	2
Totals	40	34	112	47	9	4	4	2	2	1	1	4	2	3	1	3	1	2	1	273

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Table 13.6: Age-Length key, as proportion-at-age in each 1-inch length interval, based on operculum ages for tautog sampled in Chesapeake Bay, Virginia during 2015.

	Age										
Interval	3	4	5	6	7	8	9	10	13	14	18
12 - 12.99	0	0	1	0	0	0	0	0	0	0	0
14 - 14.99	0	0.35	0.41	0.18	0.06	0	0	0	0	0	0
15 - 15.99	0.01	0.15	0.31	0.49	0.04	0	0	0	0	0	0
16 - 16.99	0	0.08	0.18	0.53	0.2	0.02	0	0	0	0	0
17 - 17.99	0	0	0.09	0.57	0.3	0.02	0	0	0.02	0	0
18 - 18.99	0	0	0.11	0.56	0	0.22	0	0.11	0	0	0
19 - 19.99	0	0.15	0	0.15	0.31	0.23	0.15	0	0	0	0
20 - 20.99	0	0	0	0	0.25	0	0	0.5	0	0.25	0
21 - 21.99	0	0	0	0	0	0	0	1	0	0	0
26 - 26.99	0	0	0	0	0	0	0	0	0	0	1

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Table 13.7: Age-Length key, as proportion-at-age in each 1-inch length interval, based on operculum ages for tautog sampled in Virginia waters of the Atlantic Ocean during 2015.

	Age																	
Interval	4	5	6	7	8	9	10	12	13	15	17	18	20	22	23	24	27	31
15 - 15.99	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
16 - 16.99	0.17	0.67	0	0.17	0	0	0	0	0	0	0	0	0	0	0	0	0	0
17 - 17.99	0	0	0.33	0.33	0.33	0	0	0	0	0	0	0	0	0	0	0	0	0
18 - 18.99	0.17	0.17	0.33	0.17	0.17	0	0	0	0	0	0	0	0	0	0	0	0	0
19 - 19.99	0	0	0.75	0	0	0.25	0	0	0	0	0	0	0	0	0	0	0	0
20 - 20.99	0	0.29	0.43	0.29	0	0	0	0	0	0	0	0	0	0	0	0	0	0
21 - 21.99	0	0	0.43	0.14	0.29	0	0.14	0	0	0	0	0	0	0	0	0	0	0
22 - 22.99	0	0	0.5	0	0.5	0	0	0	0	0	0	0	0	0	0	0	0	0
23 - 23.99	0	0	0	0	0	0	0.33	0.33	0	0.33	0	0	0	0	0	0	0	0
24 - 24.99	0	0	0	0	0	0	0	0	0	0.5	0	0	0	0.5	0	0	0	0
25 - 25.99	0	0	0	0	0	0	0	0	0	0.67	0	0.33	0	0	0	0	0	0
26 - 26.99	0	0	0	0	0	0	0	0	0	0	0	0.33	0.33	0	0.33	0	0	0
27 - 27.99	0	0	0	0	0	0	0	0	0.33	0	0.17	0	0	0	0.17	0.17	0.17	0
28 - 28.99	0	0	0	0	0	0	0	0	0	0	0	0	0	0.5	0	0	0	0.5

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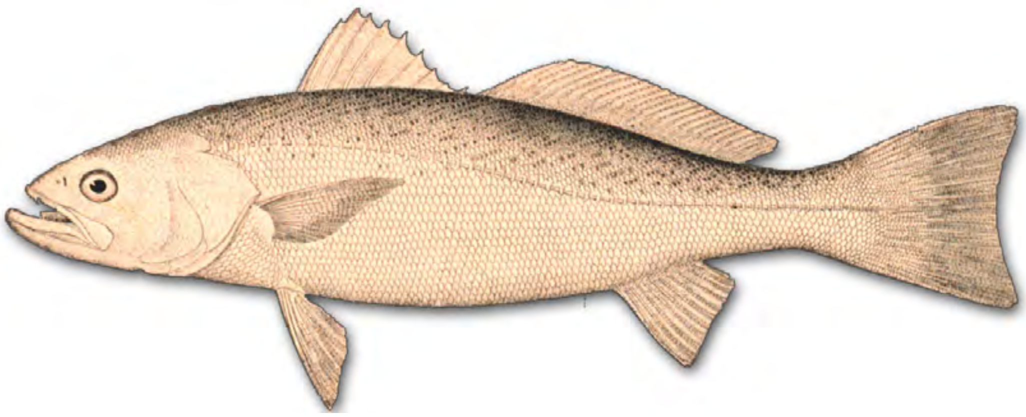
Chapter 14

WEAKFISH *Cynoscion regalis*



14.1 INTRODUCTION

We aged a total of 243 weakfish, *Cynoscion regalis*, collected by the VMRC's Biological Sampling Program for age and growth analysis in 2015. The weakfish ages ranged from 1 to 5 years old with an average age of 2.1, a standard deviation of 0.8, and a standard error of 0.05. Five age classes (1 to 5) were represented, comprising fish of the 2010 to 2014 year-classes. The sample was dominated by fish from the year-class of 2013 with 60.1%.

14.2 METHODS

14.2.1 Sample size for ageing

We estimated sample size for ageing weakfish in 2015 using a two-stage random sampling method (Quinn and Deriso 1999) to increase precision in estimates of age composition from fish sampled efficiently and effectively. The basic equation is:

$$A = \frac{V_a}{\theta_a^2 CV^2 + B_a/L} \tag{14.1}$$

where A is the sample size for ageing weakfish in 2015; θ_a stands for the proportion of age a fish in a catch. V_a and B_a represent variance components within and between length intervals for age a, respectively; CV is the coefficient of variation; L was the total number of weakfish used by VMRC to estimate length distribution of the catches from 2009 to 2013. θ_a, V_a, B_a, and CV were calculated using pooled age-length data of weakfish collected from 2009 to 2013 and using equations in Quinn and Deriso (1999). For simplicity, the equations are not listed here. The equation (14.1) indicates that the more fish that are aged, the smaller the CV (or higher precision) that will be obtained. Therefore, the criterion to age A (number) of fish is that A should be a number above which there is only a 1% CV reduction for the most major age in catch by aging an additional 100 or more fish. Finally, A_l is A multiplied by the proportion of length interval l from the length distribution of the 2009 to 2013 catch. A_l is number of fish to be aged for length interval l in 2015.

14.2.2 Handling of collections

Otoliths were received by the Age & Growth Laboratory in labeled coin envelopes, and were sorted by date of capture. Their envelope labels were verified against VMRC's collection data, and each fish was assigned a unique Age and Growth Laboratory identification number. All otoliths were stored dry in their original labeled coin envelopes.

14.2.3 Preparation

Sagittal otoliths, hereafter, referred to as "otoliths", were processed for age determination following the methods described in Lowerre-Barbieri et al. (1994) with a few modifications. The left or right otolith was randomly selected and attached, distal side down, to a glass slide with clear Crystalbond™ 509 adhesive. The otoliths were viewed by eye and, when necessary, under a stereo microscope to identify the location of the core, and the position of the core was marked using a pencil across the otolith surface. At least one transverse cross-section (hereafter, referred to as "thin-section") was then removed from the marked core of each otolith using a Buehler IsoMet™ low-speed saw equipped with two, 3-inch diameter, Norton diamond grinding wheels (hereafter, referred to as "blades"), separated by a stainless steel spacer of 0.5 mm (diameter 2.5"). Thin-sections were placed on labeled glass slides and covered with a thin layer of Flo-texx mounting medium that not only fixed the sections to the slide, but more importantly, provided enhanced contrast and greater readability by increasing light transmission through the thin-sections.

Click here to obtain the protocol at the CQFE website on how to prepare otolith thin-section for ageing weakfish.

14.2.4 Readings

The CQFE system assigns an age class to a fish based on a combination of number of annuli in a thin-section, the date of capture, and the species-specific period when the annulus is deposited. Each year, as the fish grows, its otoliths grow and leave behind markers of their age, called an annulus. Technically, an otolith annulus is the combination of both the opaque and the translucent band. In practice, only the opaque bands are counted as annuli. The number of annuli replaces "x" in our notation below, and is the initial "age" assignment of the fish.

Second, the thin-section is examined for translucent growth. If no translucent growth is visible beyond the last annulus, the otolith is called "even" and no modification of the assigned age is made. The initial assigned age, then, is the age class of the fish. Any growth beyond the last annulus can be interpreted as either being toward the next age class or within the same age class. If translucent growth is visible beyond the last annulus, a "+" is added to the notation.

By convention all fish in the Northern Hemisphere are assigned a birth date of January 1. In addition, each species has a specific period during which it deposits the annulus. If the fish is captured after the end of the species-specific annulus deposition period and before January 1, it is assigned an age class notation of "x+x", where "x" is the number of annuli in the thin-section. If the fish is captured between January 1 and the end of the species-specific annulus deposition period, it is assigned an age class notation of "x+(x+1)". Thus, any growth beyond the last annulus, after its "birthday" but before the end of annulus deposition period, is interpreted as being toward the next age class.

For example, weakfish annulus formation occurs between April and July (Lowerre-Barbieri et al. 1994 and modified by CQFE). A weakfish captured between January 1 and July 31, before the end of the species' annulus deposition period, with three visible annuli and some translucent growth after the last annulus, would be assigned an age class of "x+(x+1)" or 3+(3+1), noted as 3+4. This is the same age-class assigned to a fish with four visible annuli captured after the end of July 31, the period of annulus deposition, which would be noted as 4+4.

All thin-sections were aged by two different readers using a Nikon SMZ1000 stereo microscope under transmitted light and dark-field polarization at between 8 and 20 times magnification (Figure 14.1). Each reader aged all of the otolith samples. All samples were aged in chronological order, based on collection date, without knowledge of previously estimated ages or the specimen lengths. When the readers' ages agreed, that age was assigned to the fish. When the two readers disagreed, both readers sat down together and re-aged the fish, again

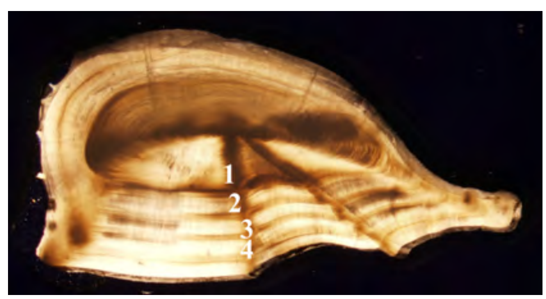


Figure 14.1: Otolith thin-section of 4 year-old weakfish

without any knowledge of previously estimated ages or lengths, and assigned a final age to the fish. When the readers were unable to agree on a final age, the fish was excluded from further analysis.

Click here to obtain the protocol at the CQFE website on how to age weakfish using their otolith thin-sections.

14.2.5 Comparison tests

A symmetry test (Hoenig et al. 1995) and coefficient of variation (CV) analysis were used to detect any systematic difference and precision on age readings, respectively, for the following comparisons: 1) between the two readers in the current year, 2) within each reader in the current year, and 3) time-series bias between the current and previous years within each reader. The readings from the entire sample for the current year were used to examine the difference between two readers. A random subsample of 50 fish from the current year was selected for second readings to examine the difference within a reader. Fifty otoliths randomly selected from fish aged in 2003 were used to examine the time-series bias within each reader. A figure of 1:1 equivalence was used to illustrate those differences (Campana et al. 1995). All statistics analyses were performed in R 3.2.2 (R Core Team 2012).

14.3 RESULTS

14.3.1 Sample size

We estimated a sample size of 318 for ageing weakfish in 2015, ranging in length interval from 6 to 35 inches (Table 14.1). This sample size provided a range in (CV) for age composition approximately from the smallest (CV) of 6% for age 2 to the largest (CV) of 11% for age 1. In 2015, we aged 243 of 283 weakfish (The rest of fish were either without otoliths or over-collected for certain length interval(s)) collected by VMRC. We fell short in our overall collections for this optimal length-class sampling estimate by 77 fish. However, we were short of no fish from the major length intervals (The interval requires 10 or more fish), as a result, the precision for the estimates of major age groups would not be influenced significantly.

14.3.2 Reading precision

Both readers had high self-precision. Specifically, there was no significant difference between the first and second readings for Reader 1 with an agreement of 96% and a CV of 1.9% (test of symmetry: $\chi^2 = 2$, $df = 1$, $P = 0.1573$), and there was no significant difference between the first and second readings for Reader 2 with an agreement of 96% and a CV of 1.9% (test of symmetry: $\chi^2 = 2$, $df = 1$, $P = 0.1573$). There was no evidence of systematic disagreement between Reader 1 and Reader 2 with an agreement of 100% (Figure 14.2).

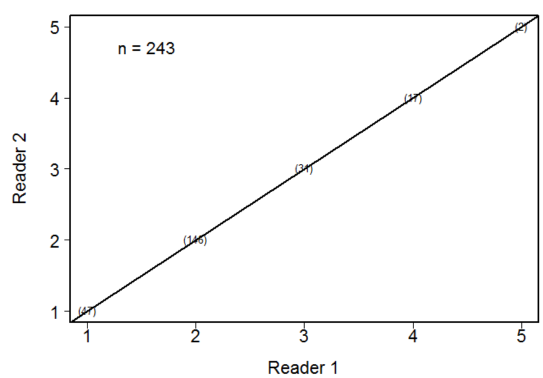


Figure 14.2: Between-reader comparison of otolith age estimates for weakfish collected in Chesapeake Bay and Virginia waters of the Atlantic Ocean in 2015.

There was no time-series bias for both readers. Reader 1 had an agreement of 98% with ages of fish aged in 2003 with a CV of

0.2% (test of symmetry: $\chi^2 = 1$, $df = 1$, $P = 0.3173$). Reader 2 had an agreement of 100%.

14.3.3 Year class

Of the 243 fish aged with otoliths, 5 age classes (1 to 5) were represented (Table 14.2). The average age was 2.1 years, and the standard deviation and standard error were 0.8 and 0.05, respectively. Year-class data show that the fishery was comprised of 5 year-classes: fish from the 2010 to 2014 year-classes, with fish primarily from the year-class of 2013 with 60.1%. The ratio of males to females was 1:4.06 in the sample collected (Figure 14.3).

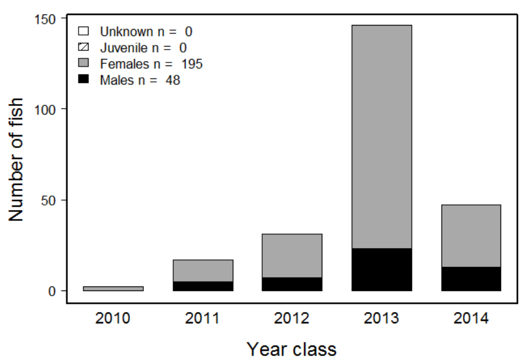


Figure 14.3: Year-class frequency distribution for weakfish collected for ageing in 2015. Distribution is broken down by sex. 'Unknown' represents gonads that were not available for examination or were not examined for sex during sampling.

14.3.4 Age-length key

We developed an age-length-key (Table 14.3) that can be used in the conversion of numbers-at-length in the estimated catch to numbers-at-age using otolith ages. The table is based on VMRC's stratified sampling of landings by total length inch intervals.

14.4 REFERENCES

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Table 14.1: Number of weakfish collected and aged in each 1-inch length interval in 2015. 'Target' represents the sample size for ageing estimated for 2015, and 'Need' represents number of fish shorted in each length interval compared to the optimum sample size for ageing and number of fish aged.

Interval	Target	Collected	Aged	Need
6 - 6.99	5	0	0	5
7 - 7.99	5	1	1	4
8 - 8.99	5	6	6	0
9 - 9.99	30	38	30	0
10 - 10.99	60	70	60	0
11 - 11.99	42	42	42	0
12 - 12.99	27	27	27	0
13 - 13.99	19	35	20	0
14 - 14.99	14	21	14	0
15 - 15.99	15	14	14	1
16 - 16.99	12	12	12	0
17 - 17.99	8	8	8	0
18 - 18.99	6	4	4	2
19 - 19.99	5	1	1	4
20 - 20.99	5	1	1	4
21 - 21.99	5	1	1	4
22 - 22.99	5	0	0	5
23 - 23.99	5	0	0	5
24 - 24.99	5	1	1	4
25 - 25.99	5	0	0	5
26 - 26.99	5	1	1	4
27 - 27.99	5	0	0	5
28 - 28.99	5	0	0	5
29 - 29.99	5	0	0	5
31 - 31.99	5	0	0	5
34 - 34.99	5	0	0	5
35 - 35.99	5	0	0	5
Totals	318	283	243	77

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Table 14.2: The number of weakfish assigned to each total length-at-age category for 243 fish sampled for otolith age determination in Virginia during 2015.

	Age					
Interval	1	2	3	4	5	Totals
7 - 7.99	1	0	0	0	0	1
8 - 8.99	1	5	0	0	0	6
9 - 9.99	5	24	1	0	0	30
10 - 10.99	6	50	4	0	0	60
11 - 11.99	14	27	1	0	0	42
12 - 12.99	6	14	5	2	0	27
13 - 13.99	9	4	5	2	0	20
14 - 14.99	1	8	5	0	0	14
15 - 15.99	2	3	5	3	1	14
16 - 16.99	1	4	3	3	1	12
17 - 17.99	1	5	1	1	0	8
18 - 18.99	0	2	1	1	0	4
19 - 19.99	0	0	0	1	0	1
20 - 20.99	0	0	0	1	0	1
21 - 21.99	0	0	0	1	0	1
24 - 24.99	0	0	0	1	0	1
26 - 26.99	0	0	0	1	0	1
Totals	47	146	31	17	2	243

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Table 14.3: Age-Length key, as proportion-at-age in each 1-inch length interval, based on otolith ages for weakfish sampled for age determination in Virginia during 2015.

	Age				
Interval	1	2	3	4	5
7 - 7.99	1	0	0	0	0
8 - 8.99	0.17	0.83	0	0	0
9 - 9.99	0.17	0.8	0.03	0	0
10 - 10.99	0.1	0.83	0.07	0	0
11 - 11.99	0.33	0.64	0.02	0	0
12 - 12.99	0.22	0.52	0.19	0.07	0
13 - 13.99	0.45	0.2	0.25	0.1	0
14 - 14.99	0.07	0.57	0.36	0	0
15 - 15.99	0.14	0.21	0.36	0.21	0.07
16 - 16.99	0.08	0.33	0.25	0.25	0.08
17 - 17.99	0.12	0.62	0.12	0.12	0
18 - 18.99	0	0.5	0.25	0.25	0
19 - 19.99	0	0	0	1	0
20 - 20.99	0	0	0	1	0
21 - 21.99	0	0	0	1	0
24 - 24.99	0	0	0	1	0
26 - 26.99	0	0	0	1	0

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